

Neotropical Diptera

Neotropical Diptera 3: 1-64 (April 15, 2009)
ISSN 1982-7121
www.neotropicaldiptera.info

Depto. de Biologia - FFCLRP
Universidade de São Paulo
Ribeirão Preto, SP, Brazil

Manual of Neotropical Diptera. Simuliidae

Sixto Coscarón

Facultad de Ciencias Naturales y Museo, Universidad Nacional de La Plata,
Paseo del Bosque, 1900 La Plata, República Argentina
Pesquisador Visitante do Departamento de Biologia, Faculdade de Filosofia, Ciências e Letras, Universidade de São
Paulo, Ribeirão Preto, SP, Brasil (Proc. FAPESP 2007/50877-5)
e-mail: sixtocos@netverk.com.ar

Cecilia L. Coscarón Arias

Research Associate

National Museum of Natural History, Smithsonian Institution
4210 Silver Hill Road
Suitland, Maryland 20746-2863, U. S. A.

&

Nelson Papavero

Museu de Zoologia, Universidade de São Paulo, São Paulo, SP, Brasil
Pesquisador Visitante do Departamento de Biologia, Faculdade de Filosofia, Ciências e Letras, Universidade de São
Paulo, Ribeirão Preto, SP, Brasil (Proc. FAPESP 2007/50878-1)

Introduction

The Neotropical Simuliidae have been most recently monographed by Coscarón & Coscarón Arias (2007), with the exception of the small genus *Pedrowygomyia* Coscarón & Miranda Esquivel, 1998 (revised by Coscarón & Miranda Esquivel, 1998) and the large genus *Gigantodax* Enderlein, 1915 (revised by Wygodzinsky & Coscarón, 1989). Basic information about black flies' bionomics (pp. 15-16), behavior (pp. 16-17), morphology (pp. 22-330), collection and preparation of material (pp. 34-35), methods of study (pp. 36-37) and bibliography (pp. 648-670) may be found in Coscarón & Coscarón Arias (2007). Some additional references are added in the ensuing paragraphs.

Some females are vicious biters and their saliva contains certain anticoagulant substances (Abebe, Cupp, Ramberg & Cupp, 1994; Abebe, Ribeiro, Cupp & Cupp, 1996). Due to their hematophagy and anthropophilic habits, simuliids have a great importance in public health and socioeconomy (Coscarón & Coscarón Arias, 2007: 17-18; see also Briceño Iragorry & Ortiz, 1957; Campos Gaona & Andrade, 1999; Coscarón, 1971; Lacey, 1981; Lacey & Charlwood, 1980; León & Wygodzinsky, 1953a, 1953b; Medeiros & Py-Daniel, 1999, 2002; Py-Daniel, Andreazze & Medeiros, 2002; Py-Daniel, Passos, Medeiros & Andreazze, 1999; Ramírez Pérez, Rassi, Conviti & Ramírez, 1976; Shelley, 1988a; Shelley, Dias, Maia-Herzog, Camargo, Costa, Garritano & Lowry, 2001; Shelley, Maia-Herzog, Dias, Camargo, Costa, Garritano & Lowry, 2001;

¹ This project was supported by FAPESP grants # 2003/10.274-9, 2007/50877-5, and 2007/50878-1.

Shelley, Maia-Herzog, Dias, Camargo & Garritano, 2001; Souza, 1984; Takaoka, 1982; Takaoka, Tada, Baba, Shimada, Lazos, Rumbea, Farias, Guderian & Amunarriz, 1988; Travis, Vargas V. & Swartzwelder, 1974; Vargas, 1941; Vieira, Brackenboro, Porter, Basáñez & Collins, 2002; Vieira, Brackenboro, Porter, Basáñez & Collins, 2005.

They are responsible for the transmission of several grave diseases to man and domestic animals, such as equine encephalitis (Sanmartín, Mackenzie, Trapido, Barreto, Mullenax, Gutiérrez & Lesmes, 1973), rabbit mixomatosis (Coscarón, 1963), vesicular stomatitis virus (Cupp, Mare, Cupp & Rambert, 1992; Howerth, Mead & Staalknecht, 2002; Mead, Gray, Noblet, Murphy, Howerth & Stalknecht, 2004), keratitis (Garrido & Campos, 2000), pemphigus foliaceus (fogo selvagem) (Aoki, Rivitti, Ito, Hans Filho & Diaz, 2005; Diaz, Sampaio, Rivitti, Marins, Cunha, Lombardi, Almeida, Castro, Macca, Lavrado, Hans-Filho, Borges, Chaul, Minelli, Empinotti, Friedman, Campbell, Labib & Anhalt, 1989), but especially of filarioses (Andrade, Medeiros, Pessoa & Py-Daniel, 2004) such as mansonellosis (Cerdeira, 1959; Kozek & Raccurt, 1983; Medeiros & Py-Daniel, 2002, 2003, 2004; Moraes, Shelley & Dias, 1985; Morales-Hojas, Post, Shelley, Maia-Herzog & Coscarón, 2001; Nathan, Tikasingh & Munroe, 1982; Shelley, 1975; Shelley, Dias & Moraes, 1980; Shelley, Maia-Herzog, Dias & Coscarón, 2002; Shelley & Shelley, 1976; Tidwell, Peterson, Ramírez Pérez & Lacey, 1980; Tidwell & Tidwell, 1982; Tidwell, Tidwell & Muñoz de Hoyos, 1980; Tidwell, Tidwell, Muñoz de Hoyos, Corredor & Barreto, 1980; Yarzábal, Basáñez, Ramírez Pérez, Ramírez, Botto & Yarzábal, 1985) and onchocerciasis (Andreazze & Py-Daniel, 2001; Andreazze, Py-Daniel & Medeiros, 2002; Anonymous, 1997; Arzube & Shelley, 1990; Barreto, Trapido & Lee, 1970; Basáñez & Boussinesq, 1999; Basáñez, Boussinesq, Proud'hon, Frontado, Villamizar, Medley & Anderson, 1994; Basáñez, Collins, Porter, Little & Brandling-Bennett, 2002; Basáñez, Remme, Alley, Bain, Shelley, Medley & Anderson, 1995; Basáñez, Rodríguez Pérez, Reyes Villanueva, Collins & Rodríguez, 1998; Basáñez, Townsend, Williams, Frontado, Villamizar & Anderson, 1996; Basáñez, Yarzábal, Frontado & Villamizar, 2000; Basáñez, Yarzábal, Takaoka, Suzuki, Noda & Tada, 1988; Bequaert, 1934; Botto, 1990; Botto, Escalona, Vivas Martínez, Behm, Delgado & Coronel, 2005; Botto, Gillespie, Vivas Martínez, Martínez, Planchart, Basáñez & Bradley, 1999; Branding-Bennett & Darsic, 1983; Calvão-Brito, Mokrabe, Maia-Herzog, Mello & Silva Jr., 1998; Campbell, Collins, Huong & Marroquin, 1980; Carabin, Escalona, Marshall, Vivas Martínez, Botto, Joseph & Basáñez, 2003; Charalambous, Lowell, Arzube & Lowry, 2005; Charalambous, Lowry, Lowell, Shelley & Arzube, 1997; Charalambous, Ready, Shelley, Arzube & Lowry, 1993; Charalambous, Shelley & Arzube, 1993; Charalambous, Shelley & Arzube, 1997; Charalambous, Shelley, Maia-Herzog & Dias, 1993; Charalambous, Shelley, Maia-Herzog & Dias, 1996a, 1996b; Collins, 1979a, 1979b; Collins, Campbell, Wilton & Newton, 1977; Collins, Lehmann, Vieira Garcia & Guderian, 1995; Collins, Ochoa, Cupp, González-Peralta & Porter, 1992; Corredor, Santiago Nicholls, Duque, Muñoz de Hoyos, Alvarez, Guderian, Lopez & Palma, 1998; Cupp, Chen & Cupp, 1997; Cupp, Cupp, Ochoa & Moulton, 1995; Dalmat, 1955; Davies, Oskam, Luján, Schoone, Kroon, López-Martínez & Paniagua-Álvarez, 1998; De León & Duke, 1966; Duke, 1970; Garms, 1975; Garms & Ochoa, 1979; Gómez-Priego, Mendoza & Rosa, 2005; Gowtage Sequeira, Higazi, Unnasch & Basáñez, 2002; Grillet, Basáñez, Vivas-Martínez, Villamizar, Frontado, Cortez, Coronel & Botto, 2001; Grillet, Botto, Basáñez & Barrera, 1994; Grillet, Villamizar, Cortez, Furtado, Escalona, Vivas-Martínez & Basáñez, 2005; Grillet, Vivas-Martínez, Villamizar, Frontado, Cortez, Coronel & Basáñez, 2002; Guderian, 1988; Guderian, Anselmi, Espinel, Mancero, Rivadaneira, Proano, Calvopina, Vieira & Cooper, 1997; Guderian, Anselmi, Espinel, Sandoval, Cooper, Rivadaneira & Guderian, 1997; Guderian, Beck, Stone, Isabel & Mackenzie, 1988; Guderian, Lovato, Anselmi, Mancero & Cooper, 1997; Guderian & Shelley, 1992; Guevara, Vieira, Lilley, López, Vieira, Rumbea, Collin, Katholi & Unnasch, 2003; Hashiguchi, Kawabata, Ito & Recinos, 1981; Hernandez, Shelley & Penn, 2002; Hoffmann, 1930, 1931; Lehmann, Cupp & Cupp, 1994a, 1994b, 1995a, 1995b; Lewis, 1963; Lewis & Ibáñez Aldecoa, 1962; Luz, Shelley & Maia-Herzog, 1996; Maia-Herzog, Shelley, Bradley, Dias, Calvão, Lowry, Camargos, Rubio, Post & Coelho, 1999; Marchon-Silva, Caér, Post, Maia-Herzog & Fernandes, 2004; Moraes, Shelley & Dias, 1986; Morales-Hojas, Post, Shelley, Maia-Herzog & Coscarón, 2001; Nettel, 1952; Ochoa, Castro, Barrios, Juárez & Tada, 1997; Omar & Garms, 1975, 1977; Porter & Collins, 1984, 1985, 1988a, 1988b; Porter, Collins & Brandling-Bennett, 1988; Post, Adams, Shelley, Maia-Herzog, Dias & Coscarón, 2003; Procunier, Shelley & Arzube, 1985; Py-Daniel, 1989, 1994a, 1994b; Py-Daniel, Andreazze & Medeiros, 2000, 2002; Py-Daniel, Passos, Medeiros & Andreazze, 1999; Py-Daniel & Py-Daniel, 1998; Ramírez Pérez, 1983, 1984, 1985; Ramírez-Ramírez, Sánchez-Tejeda, Méndez-Galván, Unnasch & Monroy-Ostria, 2006; Rassi, Lacerda & Guimaraes, 1976; Rassi, Lacerda, Guimaraes, Vulcano, Ramírez Pérez & Ramírez, 1975; Rodríguez Pérez, Danis-Lozano, Rodríguez & Bradley, 1999; Rodríguez Pérez, Danis-Lozano, Rodríguez, Unnasch & Bradley, 1999; Rodríguez Pérez, Katholi, Hassan & Unnasch, 2006; Rodríguez Pérez, Lilley, Domínguez-Vásquez, Segura-Arenas, Lizarazo-Ortega, Mendoza-Herrera, Reyes-Villanueva & Unnasch, 2004; Rodríguez Pérez, Núñez González, Lizarazo Ortega, Sánchez Varela, Wootren & Unnasch, 2006; Rodríguez Pérez & Reyes Villanueva, 1994; Rodríguez Pérez & Rivas Algalia, 1991; Rodríguez Pérez, Rodríguez, Margeli-Pérez & Rivas-Alcalá, 1995; Rodríguez Pérez, Valdivieso López & McCall, 2003; Rowe & Durand, 1998; Schiller, Petersen, Shirazian & Figueroa Marroquin, 1984; Shelley, 1988b, 1988c, 1991, 2000, 2001, 2002; Shelley & Arzube, 1985; Shelley, Arzube & Couch, 1989; Shelley, Charalambous & Arzube, 1990; Shelley, Dias, Maia-Herzog, Camargo, Costa, Garritano & Lowry, 2001; Shelley, Dias, Maia-Herzog, Procunier & Moraes, 1987; Shelley, Dias, Moraes & Procunier, 1987; Shelley, Dias, Moraes, Procunier & Couch, 1988; Shelley, Hernández, Maia-Herzog & Dias, 2002; Shelley, Lowry, Maia-Herzog, Dias & Moraes, 1997; Shelley, Maia-Herzog, Dias, Camargo, Costa, Garritano & Lowry, 2001; Shelley, Maia-Herzog, Dias & Coscarón, 2002; Shelley, Maia-Herzog, Lowry, Dias, Garritano, Camargo & Carter, 2000; Shelley, Maia-Herzog, Lowry, Dias, Garritano, Shelley, Camargo & Carter, 2000; Shelley, Mello & Rees, 1976; Shelley, Pinger, Moraes, Charlwood & Hayes, 1979; Shelley, Pinger, Moraes & Hayes, 1979; Shelley, Procunier & Arzube, 1986; Atallings, Cupp & Cupp, 2002; Tada, 1987; Takaoka, 1980, 1981, 1982; Takaoka, Ochoa, Juárez

& Hansen, 1982; Takaoka, Suzuki, Noda, Ochoa & Tada, 1984; Takaoka, Suzuki, Noda, Tada, Basáñez & Yarzábal, 1984; Takaoka, Tada, Baba, Shimada, Lazos, Rumbea, Farias D., Guderian & Amunarriz, 1988; Takaoka, Tada, Hasiguchi, Baba, Korenaga, Ochoa & Convit, 1986; Tanaka, Hashiguchi, Okazawa, Ochoa & Tada, 1980; Tidwell, Muñoz de Hoyos & Corredor, 1980; Tidwell, Peterson, Ramírez Pérez & Lacey, 1980; Tidwell, Tidwell, Muñoz de Hoyos & Corredor, 1980; Tidwell, Tidwell, Muñoz de Hoyos, Corredor & Barreto, 1980; Vargas, 1942, 1948a, 1948c, 1952; Vargas & Díaz Nájera, 1980; Vieira, Brackenboro, Porter, Basáñez & Collins, 2005; Vivas-Martínez, Basáñez, Botto, Rojas, García, Pacheco & Curtis, 2000; Vivas-Martínez, Basáñez, Botto, Villegas, García & Curtis, 2000; Wada, 1982; Yamagata, Suzuki & García-Manzo, 1986).

Studies about the biology and ecology of the larvae and their predation by some fishes have been published by Alencar, Ludwig, Soares & Hamada, 2001; Andrade & Py-Daniel, 2000; Andrade, Trivinho-Strixino, Py-Daniel & Medeiros, 2004; Crosskey, 1990; Kim & Merritt, 1988; Marino, 2003; Proculier, Shelley & Arzube, 1986; Py-Daniel & Py-Daniel, 1984; Py-Daniel & Jegu, 1998; Rodríguez Pérez, Reyes Villanueva & Rodríguez, 1995; Rodríguez Pérez, Valdivieso López & McCall, 2003; Sato, 1987; Strieder, 1986; Travis, Vargas V. & Fallas B., 1979; Travis, Vargas V. & Swartzwelder, 1974; Vargas, 1947, 1952; World Health Organization, 1982.

Larvae may be parasitized by a virus-like particle (Charpentier, Back, Garzon & Strykowski, 1986), Microsporidea (Ambrós Ginarte, Andrade & Gaona, 2003; Araújo-Coutinho, Nascimento, Figueiró & Becnel, 2004; Castello Branco Jr., 1999; Castello Branco Jr. & Andrade, 1993; Cordeiro & Castello-Branco Jr., 1988; García, 1990a, 1990b, 1990c, 1992; García, Hazard & Fukuda, 1989; Ginarte, Andrade & Gaona, 2003; Hamada, Costa & Darwich, 1997; Lutz & Splendore, 1908; Marino, Coscarón, Maurand, Loubés & Cabeza Meckert, 1980; Shelley (A. J.), 1983; Torres Fernández, Muñoz de Hoyos & Romero de Pérez, 1991; Shelley, 1983), Fungi (Torres Fernández, Muñoz de Hoyos & Romero de Pérez, 1991), especially Chytridiomycetes (López Lastra & García, 1990) and Trichomycetes: Lichtwardt, 1997; Lichtwardt, Ferrington Jr. & López Lastra, 1999; Lichtwardt, López-Lastra & Mazzuchelli, 2000; López Lastra, Scorsetti, Marti & Coscarón, 2005; McCreadie & Beard, 2003; Ríos-Velásquez & Hamada, 2002; Vojvodiæ, Nelder & McCreadie, 2006) and Nematoda (Mermithidae) (Ambrós Ginarte, Andrade & Gaona, 2003; Camino, 1985, 1986, 1988, 1990, 1991a, 1991b, 1992, 1993a, 1993b, 1994; Camino & Poinar Jr., 1988; Camino & Villalobos, 1997; Ginarte, Andrade & Gaona, 2003; Torres Fernández, Muñoz de Hoyos & Romero de Pérez, 1991; Villalobos & Camino, 1997).

Adults may be easily bred from pupae; mass rearings techniques, for various purposes, have been devised by Bernardo & Cupp, 1986; Edmon & Simmnons, 1985; Figueiró, Nascimento & Coutinho, 2002; Figueiró, Docile & Aranda, 2006; Hamada, Costa & Darwich, 1997 and Muirhead-Thomson, 1969.

Several control measures have been proposed (Coscarón & Coscarón Arias, 2007: 18-21; see also Andrade & Campos, 1995; Araújo Coutinho, Figueiró, Viviani, Nascimento & Cavados, 2005; Campos Gaona & Andrade, 2001; Castello Branco Jr. & Andrade, 1992; Cavados, Fonseca, Chaves, Araújo-Coutinho & Rabinovitch, 2005; Cavados, Majerovich, Chaves, Araújo-Coutinho & Rabinovitch, 2004; Cupp, Duke, Mackenzie, Guzmán, Vieira, Méndez-Galván, Castro, Richards, Sauerbrey, Domínguez, Eversole & Cupp, 2004; Cupp, Ochoa, Collins, Cupp, González-Peralta, Castro & Zea-Flores, 1992; Cupp, Ochoa, Collins, Ramberg & Zea, 1989; Elliot & Potter, 1978; Figueroa, Collins & Kozek, 1977; Gay, Adler & Noblet, 1997; Gray, Adler, Coscarón Arias & Noblet, 1999; Guderian, Anselmi, Espinel, Mancero, Rivadaneira, Proano, Calvopina, Vieira & Cooper, 1997; Jamnback, 1973; Kim & Merritt, 1988; Lacey & Undeen, 1988; Mardini, 2006; Mardini, Souza, Rabinovitch, Alves & Silva, 1999; Marino, 1993, 2003; Nettel, 1952; Ochoa, Castro, Barrios, Juárez & Tada, 1997; Overmyer, Armbrust & Noblet, 2003; Py-Daniel & Darwich, 1997; Rodríguez Pérez & Reyes Villanueva, 1994; Rodríguez Pérez, Reyes Villanueva, Barrera Saldaña, Domínguez Vásquez & Lizarazo Ortega, 2007; Rodríguez Pérez, Rodríguez, Margeli-Pérez & Rivas-Alcalá, 1995; Romaña & Ábalos, 1948; Ruas Neto, 1984a, 1984b; Ruas Neto & Silveira, 1989; Ruas Neto, Souza, Severino, Melo, Silveira & Fontes, 1985; Smith, 1973; Vargas, 1948b; Vieira, Brackenboro, Porter, Basáñez & Collins, 2002; Vivas-Martínez, Basáñez, Botto, Villegas, García & Curtis, 2000; Vivas-Martínez, Basáñez, Grillet, Weiss, Botto, García, Villamizar & Chavasse, 1998; Yamagata, Ochoa, Molina, Sato, Uemoto & Suzuki, 1987).

Illustration of the present publication have been extracted and modified from the original plates of papers published by the first author.

1. Key to the genera of Neotropical Simuliidae

1.1. Adults

1. Pedisulcus well developed (Figs. 8A-B: pe); basal cell invariably absent; length of basal section of R (as measured from humeral cross-vein) less than 1/3 distance from base of Rs to wing tip
..... *Simulium* Latreille, 1802 [For subgenera and species-groups, see below]
Pedisulcus absent (Figs. 8C-F); basal cell absent or present (Fig. 3A); length of basal section of R about 1/3 distance mentioned or more (Figs. 3A-B) 2
2. (1) Cu₂ and A₁ straight, basal cell absent (Fig. 3C). Arms of female genital fork with long cephalad directed apodemes

(Fig. 11K)	3
<i>Cu₂</i> and <i>A₁</i> curved, basal cell absent or present (Fig. 3A). Arms of female genital fork without apodemes (Figs. 11J, L-M) (except in <i>Cnesia</i>)	4
3. (2) First flagellomere shorter than wide; setae of basal flagellomeres and palpomeres shorter than segment (as in Fig. 2A); clypeus longer than wide (Fig. 1A); female mouthparts normal, functional; calcipala present (Figs. 8D-F); sub-basal tooth on claw well developed (Figs. 11D-E)	<i>Gigantodax</i> Enderlein, 1925 [For species-groups, see below]
First flagellomere as long as wide; setae of basal flagellomeres and palpomeres longer than segment (Fig. 2B); clypeus as long as wide; female mouthparts reduced, non functional; calcipala absent (Fig. 8C), sub-basal tooth on claw obsolescent (Fig. 11C)	<i>Pedrowygomyia</i> Coscarón & Miranda Esquivel, 1998
4(2) Upper portion of pleural membrane hairy (Fig. 2G); apical segment of maxillary palp more than twice as long as penultimate segment (as in Fig. 2C)	<i>Tlalocomya</i> Wygodzinsky & Díaz Nájera, 1970
Pleural membrane entirely bare; apical segment of maxillary palp generally less than twice as long as penultimate segment (Figs. 2D-F)	5
5(4) Antenna with 10 segments; arms of furcasternum with conspicuous projections (Fig. 8H); male terminalia with geniculate median sclerite, composed of a slender, basal, Y-shaped portion and 2 sub-parallel elongate apical arms (Fig. 17H); apical portion of endoparameres with numerous elongate denticles; genital fork of female almost entirely unpigmented, its stem very stout, not longer than arms (Fig. 11L)	<i>Paraustrosimulum</i> Wygodzinsky & Coscarón, 1964
Antenna with 11 segments (as in Fig. 2A); arms of furcasternum lacking conspicuous projections (Fig. 8G) (except <i>Lutzsimulium</i> ; Fig. 8S); median sclerite of male not as above, or apical portion of endoparameres obsolescent, lacking denticles (Figs. 16B, 17A); genital fork distinctly pigmented, with stem not as above (Figs. 11J, M)	6
6(5) <i>R_i</i> only with hair-like setae, spiniform setae not developed (as in Fig. 3E); basal tooth of female claws large (Fig. 11F)	<i>Cnesiamima</i> Wygodzinsky & Coscarón, 1973
<i>R_i</i> with hair-like and spiniform setae (Fig. 3D); if spiniform setae not very distinct, then basal tooth of female claw small (Figs. 11A-B)	7
7(6) Basal cell absent; mandible of female toothed only on internal margin (similar to Fig. 10M); female claws with small sub-basal tooth; male terminalia with teeth of endoparameral organ not perceptible (Fig. 16B)	8
Basal cell present (Fig. 3A); mandible of female as above or with well-developed teeth on internal and external margins (Fig. 10L); claws of female with larger basal tooth (Figs. 11A-B); male terminalia with teeth of endoparameral organ distinct (Fig. 17C)	9
8(7) Frons longer than wide, with median sulcus relatively short and without basal bifurcated branches (Fig. 1D); Sc with about 50 setae; furcasternal branches with large projections (Fig. 8J). Male gonocoxite about twice as long as gonostylus (Fig. 20A)	<i>Lutzsimulium</i> d'Andretta Jr. & d'Andretta, 1947
Frons about as long as wide, with median sulcus elongated and bifurcated, with long basal branches (Fig. 1E); Sc without setae; furcasternal branches with very small projections (Fig. 8I). Male gonocoxite 3 times as long as gonostylus (Fig. 20B)	<i>Kempfsimulium</i> Py-Daniel & Mello, 1982
9(7) Wings with a slight curvature in <i>Cu₂</i> ; <i>A₁</i> almost attaining wing margin (Fig. 3B); male terminalia with basal portion of endoparameres obsolescent, denticles of distal portion well developed but few in number, forming a tight group of characteristic arrangement; female with arms of genital fork bearing 2 forwardly directed apodemes (as in Fig. 11K)	<i>Cnesia</i> Enderlein, 1934
Wings with pronounced curvature of <i>Cu₂</i> ; <i>A₁</i> ending well before wing margin (Fig. 3A); endoparameres of male not as above; genital fork of female without apodemes (Figs. 11J)	10
10(9) Males	11
Females	13
11(10). Median sclerite with basal transverse satellite plate continuous distally with deep cleft and bifurcated apically (Fig. 17B); endoparameres lacking teeth (similar to Fig. 17A) ...	<i>Mayacnephia</i> Wygodzinsky & Coscarón, 1973
Median sclerite of complex geniculate, with distal half divided into 2 wide but short and divergent arms (Figs. 17C-D); endoparameres with numerous teeth	12

- 12(11) Ventral plate of terminalia sub-triangular, and basal arms narrow, straight (Figs. 18A); arms of median sclerite parallel (Fig. 17D); endoparameres with basal portion large and denticles of apical portion well developed
..... *Araucnephia* Wygodzinsky & Coscarón, 1973
- Ventral plate of terminalia sub-rectangular and basal arms stout, rounded at apex and curved (Fig. 18B); arms of median sclerite sharply diverging apically (Fig. 17E); endoparameres with basal portion small and denticles of apical portion obsolescent *Araucnephiooides* Wygodzinsky & Coscarón, 1973
- 13(10) Basal tooth of claws narrow, hook like (Fig. 11B) *Araucnephiooides* Wygodzinsky & Coscarón, 1973
Basal tooth of claws larger, sub-triangular or spatulate (as in Figs. 11A, F) 14
- 14(13) Frons very narrow (Fig. 1B), frontal angle approximately 50°; inner surface of spermatheca without spiculae.
Paraproct about half as long as its own width (Mesoamerica)
..... *Mayacnephia* Wygodzinsky & Coscarón, 1973
- Frons wider (Fig. 1C), frontal angle approximately 85°; inner surface of spermatheca with scattered spiculae.
Paraproct about as long as wide (Fig. 13A) (Southern South America)
..... *Araucnephia* Wygodzinsky & Coscarón, 1973

1.2. Pupae (Figs. 24-30)

1. Abdominal sterna VI and VII divided longitudinally along middle by a membranous, striate area (Fig. 30C); tergal hooks VI-VIII invariably simple; abdominal segments VIII and IX in many cases with strongly curved, twisted, looped, or grapnel-like strong setae (Figs. 30G-H) 2
Abdominal sterna VI and VII entire; in many cases some hooks on terga VI-VIII, bifid or trifid; abdominal terga VIII and IX without the above-mentioned setae *Simulium* Latreille, 1802
- 2(1) Terminal processes of abdomen short and pointed (Fig. 30G), or absent (Fig. 30D) 3
Terminal processes of abdomen elongate (Fig. 30H) 4
- 3(2) Cocoon reduced to a small pad, on which the terminal abdominal segments are inserted (Fig. 24A); gills in the shape of a thick stem, with a few thread-like filaments (Fig. 27A); abdomen strongly sclerotized, terga and sterna with a large number of supernumerary spines or hooks; apex of abdomen blunt; terminal processes absent (Fig. 30D) *Tlalocomyia* Wygodzinsky & Díaz Nájera, 1970
Cocoon well developed, of definite shape, covering whole body of pupa except gills, each one of the latter in the shape of a twisted, pseudo-segmented lamella (Fig. 24D); abdomen less sclerotized than thorax; abdomen without supernumerary hooks, and with short terminal processes (Fig. 30G)
..... *Cnesiamima* Wygodzinsky & Coscarón, 1973 and *Paraustrosimulum* Wygodzinsky & Coscarón, 1964
- 4(2) Abdominal terga with combs of spines; tergum VIII without large hooks 5
Abdominal terga without combs of spines; tergum VIII with 4 large hooks *Cnesia* Enderlein, 1934
- 5(4) Facial trichomes (Fig. 26D), thoracic trichomes (Figs. 29B-C) and setae of eighth and ninth abdominal segments tightly looped apically (Fig. 30H) 6
At least facial and thoracic trichomes not looped apically 7
- 6(5) Gill with 12-22 branches; frontoclypeus and thorax with tubercles *Lutzsimulum* d'Andretta Jr. & d'Andretta, 1947
Gill with 12 branches; frontoclypeus and thorax without tubercles (Fig. 26D)
..... *Kempf simulum* Py-Daniel & Mello, 1982
- 7(5) Head sclerite with frontal, facial, epicranial, lateral, and genal hair trichome-like (Figs. 26A)
..... *Araucnephia* Wygodzinsky & Coscarón, 1973
Head sclerite without all the trichomes mentioned above; facial trichomes hair-like or spine-like 8
- 8(7) Thorax with tubercles arranged in circles; basal portion of gills with minute plate-like cuticular structures; thoracic trichomes hair-like *Araucnephiooides* Wygodzinsky & Coscarón, 1973
Thorax with tubercles not arranged in circles; basal portion of gills without the aforementioned cuticular structures; thoracic trichomes hair- or spine-like 9

- 9(8) Cephalic sclerite with 2+2 or 3+3 frontal trichomes; branches of gills more or less tubular
..... *Mayacnephia* Wygodzinsky & Coscarón, 1973
Cephalic sclerite with 1+1 or without frontal trichomes; gill branches varied in shape 10

10(9) Frontal and frontoclypeal trichomes absent (Fig. 26C); clypeus comparatively narrow *Gigantodax* Enderlein, 1925
Frontal and frontoclypeal trichomes present (Fig. 26B); clypeus comparatively wide
..... *Pedrowygomyia* Coscarón & Miranda Esquivel, 1998

1.3. Larvae (Figs. 31-36)

1. Anal sclerite with accessory sclerite forming a complete ring around posterior end of body (Fig. 36A); each mandibles with 3 outer teeth (Fig. 34B) 2

Ring-shaped accessory sclerite not developed (Fig. 36B); each mandible with 2 or 4 outer teeth 3

2(1) Hypostomium with first intermediate tooth generally projecting beyond level of lateral tooth (Fig. 35C); cephalic larval apotome with basal spots *Pedrowygomyia* Coscarón & Miranda Esquivel, 1998

Hypostomium with lateral tooth, projecting beyond level of first intermediate tooth (Figs. 35D-E); cephalic apotome without basal spots *Gigantodax* Enderlein, 1925

3(1) Teeth of hypostomium arranged into 3 conspicuous groups (as in Fig. 35A); mandible with numerous (over 5) marginal serrations 4

Teeth of hypostomium more evenly distributed, not arranged into conspicuous groups (Fig. 35B); mandible with marginal serrations less numerous, not more than 5, generally only 2 (as in Fig. 34H) 7

4(3) Antennae approximately as long as stem of cephalic fan; hypostomium with either 13 or 17 teeth 5

Antennae much shorter than stem of cephalic fan; hypostomium with 15 teeth *Araucnephia* Wygodzinsky & Coscarón, 1973

5(4). Cervical sclerites very small, sub-ovoidal; mandibles with accessory teeth at level of inner teeth, with second preapical tooth minute and with short, slender, basal setae, only faintly dentate; hypostomium with 17 teeth *Araucnephoides* Wygodzinsky & Coscarón, 1973

Cervical sclerites wide or transverse, sub-rectangular, or transverse and fused to the upper ends of the postocciput; mandibles without accessory serrations at level of inner teeth, with second preapical tooth as long as first or only slightly shorter and with short basal setae bearing conspicuous elongated denticles; hypostomium with 13 teeth 6

6(5). Proximal antennal segment more than half as long as medial segment *Mayacnephia* Wygodzinsky & Coscarón, 1973

Proximal antennal segment at most half as long as medial segment *Tlalocomyia* Wygodzinsky & Díaz Nájera, 1970

7(3). Antenna much longer than stem of cephalic fan; distal antennal segment distinctly longer than proximal and medial segments combined; backwardly directed struts underlying main body of anal sclerite *Cnesiamima* Wygodzinsky & Coscarón, 1973 and *Paraustrosimulum* Wygodzinsky & Coscarón, 1964

Antenna not longer than stem of cephalic fan (Fig. 32A); distal segment not or only slightly longer than proximal and medial segments combined (Figs. 33F-L); anal sclerite without struts 8

8(7). Postgenal cleft very shallow (Fig. 31C); hypostomium with 17 teeth, the 4+4 intermediate ones similar to the remaining teeth (Fig. 35B); mandibles with 4 outer teeth *Cnesia* Enderlein, 1934

Postgenal cleft generally well developed (Figs. 31D-F); hypostomium with at most 13 teeth, the 1+1 lateral ones, when perceptible, somewhat similar to lateral serrations; mandible with 2 outer teeth 9

9(8). Preapical teeth of mandible sub-equal in size (Figs. 34F-H) or decreasing in size from first to third (Fig. 34D-E, G) *Simulium* Latreille, 1802

Preapical teeth of mandible with the second tooth smaller than either first or third 10

- 10(9). Body integument with or without setae; without cylindrical projections; rays of cephalic fan with longer setae alternately disposed among smaller setae; antenna with basal segments not overly wide; antenna slightly shorter than or as long as stem of cephalic fan; ninth abdominal segment with 1+1 ventral tubercles; anal ring with about 75 rows and 13 hooks each *Lutzsimulium* d'Andretta Jr. & d'Andretta, 1947
 Body integument without setae; cylindrical projections present; rays of cephalic fan with setae arranged homogeneously, without larger setae (Fig. 33N); antenna with basal segments very wide; antenna shorter than stem of cephalic fan; ninth abdominal segment without ventral tubercles; anal ring with 110-130 rows and about 22 hooks each *Kempfisimulium* Py-Daniel & Mello, 1982

2. Key to species groups of *Gigantodax*

2.1. Adults

1. Sub-basal tooth of claw hook-shaped in females (as in Fig. 11B); aedeagal membrane with spinules (Figs. 17F-H), ventral plates, distally, with a concavity in males (Figs. 17F, 18C) 2
 Sub-basal tooth of claw sub-triangular or sub-rhomboidal (Fig. 11D); aedeagal membrane without spinules, and ventral plate without a distal concavity in males 3
- 2(1). Gonapophyses with distal border elongated (Fig. 12E), spermatheca of normal size, mandible with teeth only on inner side (as in Fig. 10M); spinules of aedeagus well developed, some of them sclerotized basally and fused like a shield (Figs. 17F-G) *igniculus*-group
 Gonapophyses with distal border not elongate, only lobulate (Fig. 12B-C); spermatheca large, distinctly developed (Fig. 12B); mandible with teeth on both sides (as in Fig. 10L); spinules of aedeagus poorly developed (Fig. 17I) *minor*-group
- 3 (1). Sensorial vesicle of maxillary palp poorly developed (Fig. 2E); gonapophyses with distal border lobulate; calcipala nearly as long as second tarsomere (Fig. 8F); sub-basal tooth of claw sub-triangular (Fig. 11E) ... *multifilis*-group
 Sensorial vesicle well developed (Fig. 2F); gonapophyses with distal border straight (Fig. 12D); calcipala about half as long as second tarsomere (Fig. 8E); sub-basal tooth of claw sub-rhomboidal or sub-triangular *brophyi*, *cilicinus*, *cormonsi* and *wrighti*-groups

2.2. Pupae

1. Gill branches arborescent, with over 100 terminal filaments (Figs. 24B, 27B) 2
 Gill branches with not more than 18 terminal filaments, or branches with tubular or globular shape 3
- 2(1). Terminal spines thin and pointed; abdominal tergite 1 with 3 + 3 (4 + 4) or 4 + 4 (5 + 5) hairy trichomes (as in Fig. 30B) ... *multifilis*-group
 Terminal hooks stout; abdominal tergite 1 with 5 + 5 or 7 + 7 trichomes (Figs. 30A) *igniculus*-group
- 3 (1). Gill filamentous branches not thicker basad and without tegumentary process (Figs. 27C-D) 4
 Gills with tubular or sub-globose branches, with cuticular processes; if filamentous, they are thickened basally and generally the 12th and 16th ones more elongated (Figs. 27F-J) 5
- 4 (3). 11-13 relatively flexuous gill branches (Fig. 27C); terminal hook thin and elongated, pointed distally, trichomes of frontoclypeus and thorax hooked distally (Fig. 29A) *minor*-group
 18 relatively resistant (not flexuous) gill branches (Figs. 24C, 27D); terminal spines stout (Fig. 30F); trichomes of frontoclypeus and thorax not hooked *brophyi*-group
- 5 (3). Gill branches with 17-18 tubular branches, generally with tegumentary processes (Figs. 27E, H-J) 6
 Gill branches reduced in number, globose, with cuticular processes, or membranous (Figs. 27K-L) *wrighti*-group
- 6 (5). Branches independent (Fig. 27E) *cilicinus*-group
 Branches often fused, making a shield-like formation (Figs. 27I-J) *cormonsi*-group

2.3. Larvae

1. Basal segment of antenna at most half as long as second (Fig. 33A); second antennal segment as long as third; median tooth of hypostomium longer than corner teeth, an imaginary line going from the medium tooth to the fourth lateral tooth does not intersect the other teeth (Fig. 35E); numerous mandibular serrations (Fig. 34A) *igniculus*-group
Second antennal segment not longer than first or third; median tooth of hypostomium not longer than corner teeth; an imaginary line passing over the cuspid of the medium tooth to the fourth lateral tooth intersecting the other teeth (Fig. 35D); generally with a few mandibular serrations present 2
- 2 (1). Third antennal segment longer than first and second together; second segment very short, less than one-third length of basal segment (Fig. 33B); anal scales reduced, not completing a ring *minor*-group
Third antennal segment shorter than first and second together; basal segment about twice (or more) size of second segment (Figs. 35C-E); anal scales well developed, forming a ring (Fig. 36A) 3
- 3 (2). Anal gills with numerous lobules (more than three, Fig. 36E); mandible with reduced marginal serrations *multifilis*-group
Anal gill with only three lobules; mandible with abundant marginal serrations (Fig. 34C) *brophyi*, *cilicinus*, *cormonsi* and *wrighti*-groups

3. Keys to *Simulium* subgenera and species-groups

3.1. Females

1. General coloration black or blackish-brown 2
General coloration brown (reddish-brown to yellowish) 22
- 2 (1). Body without special ornamentation; scutum hairs homogeneously distributed; claws with basal tooth large, sub-ovoid, elongated; basal portion of cibarium smooth, without reinforcement at sides *Eusimulium* Roubaud, 1906 and *Nevermannia* Enderlein, 1921
Body ornamented with light vittae or spots; scutal hairs distributed homogeneously or in groups; claws with or without sub-basal tooth, but if present, it is proportionally smaller and sub-triangular (except *Byssodon* with basal tooth) 3
- 3 (2). With setae on basal sector of R 4
Without setae on basal sector of R (Fig. 3F) 9
- 4 (3). Grayish pollinose coloration; scutum with light area limited to 1 median and 1+1 sub-lateral stripes (Fig. 4A); abdomen blackish or grayish with 1+1 silver transverse bands, or with bands on posterior border of tergites III-IV (Figs. 7A-C) *Pternaspatha* Enderlein, 1930 (part)
Blackish coloration; scutum with or without light sub-median stripes or anterior spots; abdomen blackish without silvery transverse bands on tergites III-IV 5
- 5 (4). Paraproct sub-triangular, about twice as long as wide at base (Fig. 13E); basal portion of cibarium smooth (Fig. 9A) *Chirostilbia* Enderlein, 1921
Paraproct not sub-triangular, curved distally, slightly shorter than wide at base (Fig. 13F); cibarium smooth or with teeth 6
- 6 (5). Basal portion of cibarium with teeth arranged in elevated median sub-conical group (Fig. 9B) *Inaequalium* Coscarón & Wygodzinsky, 1984
Cibarium generally smooth or with small teeth: if teeth present, these arranged in median elevated group, scutum with 1+1 light anterior spots or 3 black longitudinal stripes separated by nacreous stripes that change tone with different light positions 7
- 7 (6). Cibarium with very small teeth or without teeth (Figs. 9A, 10E-F); paraprocts relatively short, not lobulate, scarcely acuminate apically and directed downwards (Figs. 14B-C) *Aspathia* Wygodzinsky & Coscarón, 1973 (part)

- Cibarium with 1 row of teeth (Figs. 10I-J) or without teeth but with reinforced border (Fig. 10K); paraproct relatively long, lobulate and scarcely sclerotized, not acuminate (Figs. 14I-L) 8
- 8(7). Scutum with setae grouped, simulating scales, without anterior grayish sub-quadratae spots; cibarium smooth (Fig. 10K, or as in Fig. 10G) *Thrysopelma* Enderlein, 1934 (part)
- Scutum with setae homogeneously distributed; with anterior grayish sub-quadratae light spots (Fig. 6H); cibarium with teeth arranged in 1 row (Figs. 10I-J) *Trichodagmia* Enderlein, 1934 (part)
- 9(3). Cibarium without teeth 10
- Cibarium with teeth 16
- 10(9). Scutum with hairs grouped like scales (Fig. 2H); claw without sub-basal tooth or much reduced (Figs. 11G-H); fronto-ocular triangle absent (as in Fig. 11); paraprocts generally very thin distally (Figs. 13C-D) *Notolepria* Enderlein, 1930
- Scutum with hairs homogeneously distributed; claw with or without sub-basal tooth, fronto-ocular triangle generally well developed; paraprocts generally not thinner distally 11
- 11(10). Grayish pollinose species; abdomen grayish, with tergites III-V blackish or velvety brown, generally with an anteromedian or 1+1 silvery lateral spots; calcipala frequently reduced or absent 12
- Blackish species; abdomen blackish, without silver spots on tergites III-V; calcipala well developed 13
- 12(11). Claws with sub-basal tooth; scutum with blackish areas delimited by grayish median and sub-lateral stripes; abdominal tergites III-V blackish, with 1+1 sub-lateral posterior whitish spots (Fig. 7C); paraprocts sub-rectangular, a little shorter than wide (Fig. 13B) *Pternaspatha* Enderlein, 1930
- Claws without sub-basal tooth (Fig. 11I); scutum uniformly grayish, except for 1+1 light median anterior spots; tergites III-V blackish or velvety brown, with or without spots anteromedially (Figs. 7H-I); paraprocts sub-triangular (Fig. 14A), as long as wide *Psilopeltia* Enderlein, 1934 (*blancasi* species group)
- 13(11). Paraproct sub-triangular, with distal border arched (Figs. 14G); scutum with 1+1 anterior light submedian spots continued posteriorly with a thin stripe joining the grayish posterior area *Hemicnetha* Enderlein, 1934 (*oviedoii* species group)
- Paraproct not sub-triangular, with distal border truncate or acuminate posteriorly; scutum ornamentation variable 14
- 14(13). Paraprocts with distal edge truncate and slightly acute posteriorly (Figs. 14B-C); scutum with median and 1+1 sub-median black stripes delimited by nacreous vittae that change color according to incidence of light (Figs. 6A-B) or with only 1+1 anterior, submedian, nacreous spots (Fig. 6C) *Aspathia* Enderlein, 1935
- Paraprocts with distal edge truncate or rounded, but not acuminate posteriorly; scutum without contrasting silvery vittae (Figs. 6D-G) 15
- 15(14). Paraproct as long as wide at base, with external surface mostly nude and shiny, with depressions and distal border truncated (Fig. 14H); gonapophysis as long as wide (Fig. 12G) *Hearlea* Vargas, Martínez Palacios & Díaz Nájera, 1946
- Paraproct longer than wide at base, external surface mostly covered by microtrichia, without depressions and distal border curved (Figs. 14E-F), gonapophysis longer than wide (Fig. 12F) *Hemicnetha* Enderlein, 1934 (part, *brachycladum* species group)
- 16(9). Scutum with abundant gray pollinosity, showing, according to incidence of light, 3-5 longitudinal, light stripes; cibarium with a weak concavity covered by small, similar, acute teeth; paraprocts sub-triangular, about as long as wide; claws without teeth *Psilozia* Enderlein, 1936
- Scutum with sparse pollinosity, with or without 1+1 silvery longitudinal stripes; cibarium with or without teeth, generally with median depression; paraprocts not sub-triangular, with curved distal edge, generally longer than wide and sometimes thinner distally; claws with or without teeth 17
- 17(16). Claws with large, sub-ovoid, basal tooth; posterior edge of tergites III-V frequently with 1+1 transverse silvery bands *Byssodon* Enderlein, 1925
- Claws with a small, sub-triangular, sub-basal tooth or without tooth; tergites III-V without silvery transverse bands 18

- 18(17). Cibarium generally without teeth, but if teeth present, not in the median depression and at sides (Figs. 10E-F); scutum with 1+1 sub-median, longitudinal, silvery stripes generally reaching the posterior grayish area (Figs. 6A-B) or darker, according to incidence of light, or yet reduced to 1 + 1 anterior, sublateral, nacreous spots (Fig. 6C); fronto-ocular triangle well developed *Aspathia* Enderlein, 1935
 Cibarium with median depression sclerotized or not and with small teeth at sides (Figs. 9C-G, K, 10C-D); scutum with or without silvery longitudinal stripes that can reach to posterior silvery area; fronto-ocular triangle generally absent or reduced 19
- 19(18). Cibarium with central area well sclerotized and with small, rounded teeth (Figs. 9E-G); infra-frontal sutures well developed (Fig. 1F) *Coscaroniellum* Py-Daniel, 1983 (*quadrifidum* species group)
 Cibarium with central area not sclerotized, teeth acute (Figs. 9C-D); infra-frontal sutures not developed 20
- 20(19). Scutum black, with or without silvery sub-median stripe mostly frequent on anterior area and wedge shaped (Figs. 4B-D); when reaching posterior silvery area, without gray shadow spots on silvery stripe
 *Psaroniocompsa* Enderlein, 1934
 Scutum grayish to black with silvery stripe reaching posterior grayish area, generally with gray shadow spots on silvery stripes (Figs. 4F-I, 5A-B) 21
- 21.(20). Cibarium with small teeth of about the same size and absent on the median depression (Fig. 9D); fronto-ocular triangle very short (Fig. 1I); abdomen without silvery posterior transverse spots on tergites III-V or very short (Fig. 7D) *Cerqueirellum* Py-Daniel, 1983
 Cibarium with sub-median teeth larger than other teeth and with small teeth on the median area (Fig. 9H); fronto-ocular triangle relatively deep; abdomen with silvery posterior transverse spots on tergites III-V about 1/3 of the tergite's height (Fig. 7E); scutum with sub-median, silvery vittae narrow (Fig. 5C)
 *Coscaroniellum* Py-Daniel, 1983 (*quadriovittatum* species group)
- 22(1). Yellowish-reddish to greenish-gray species; if scutum black, margins yellowish (Figs. 5D-F); paraprocts sub-triangular (Figs. 13F-K), longer than wide at base and distally relatively thin (Fig. 14L) 23
 Reddish-brown to dark grayish species; scutum generally with lyre-shaped pattern (Fig. 6H); paraprocts sub-rectangular and distally not acuminate (Figs. 14I-J) 28
- 23(22). Scutum gray-greenish to brown-yellowish; cibarium smooth, margin not strongly sclerotized (Fig. 9A)
 *Chirostilbia* Enderlein, 1921 (*subpallidum* species group)
 Scutum orange to reddish or black, bordered by yellow, frequently with sub-median silvery stripes; cibarium with sclerotized margin or with small teeth 24
- 24(23). Scutum with 1+1 or more nacreous or silvery longitudinal stripes (Figs. 5H-I); abdomen generally with 3 longitudinal rows of sub-quadrata blackish spots (Figs. 7F-G); cibarium with median depression well sclerotized, bordered with weak lobules and with teeth on weakly elevated areas at sides (Figs. 10A-D); paraprocts thinner distally (Figs. 13J-K, 14L) *Psilopeltia* Enderlein, 1934 (*escomeli* species group)
 Scutum generally without 1+1 or more nacreous or silvery longitudinal stripes (Figs. 5D-G); abdomen without sub-quadrata blackish spots; cibarium without teeth or with teeth generally larger and with different disposition from above (Figs. 9I-O); paraprocts scarcely thin distally (Figs. 13G-I) (*Ectemnaspis* Enderlein, 1914) 25
- 25(24). Scutum yellow or black, in the latter case, totally with yellow margins (Figs. 5D-E); cibarium without or with teeth, in the latter case, with plates or small lobules at median area (Figs. 9I-L) 26
 Scutum yellow or reddish, cibarium with teeth but median area without plates 27
- 26(25). Cibarium without teeth, but with well sclerotized edge (Figs. 9I-J) ... *Ectemnaspis* Enderlein, 1914 (*bicoloratum* species group)
 Cibarium with teeth and median area with plates or lobules (Figs. 9K-L)
 *Ectemnaspis* Enderlein, 1914 (*romanae* species group)
- 27(25). Cibarium with well-developed teeth grouped on 1+1 weak elevations (Figs. 9M-N)
 *Ectemnaspis* Enderlein, 1914 (*perflavum* species group)
 Cibarium with very small teeth and with 1+1 sub-median processes generally with teeth on apex (Fig. 9O)
 *Ectemnaspis* Enderlein, 1914 (*dinelli* species group)

- 28(22). Gonapophysis sub-ovoid, shorter than wide (Fig. 12I); paraproct slightly acuminate distally (Fig. 14L); scutum with hairs grouped and arranged into longitudinal rows *Thrysopelma* Enderlein, 1934 (part)
Gonapophysis sub-triangular (Fig. 12F) or sub-ovoid (Figs. 12H-I), longer than wide or about as long as wide; paraprocts not acuminate distally (Figs. 14D, E-F, I-J); scutum with hairs homogenously distributed or in groups, but not arranged in longitudinal rows 29
- 29(28). Cibarium scarcely sclerotized on median area and without teeth or spiculae (Fig. 10G); paraprocts slightly longer than wide, sub-rectangular and with abundant microtrichia (Fig. 14D) *Hemicnetha* Enderlein, 1934 (*paynei* species group)
Cibarium well sclerotized on median area, with thin, acuminate teeth and scarcely sclerotized (Figs. 10H-J); paraprocts about twice as long as wide and with sparse microtrichia (Figs. 14E-F, I-J) 30
- 30(29). Gonapophysis more than twice as long as wide, acute apically (Fig. 12F); cibarium with small, disordered teeth (Fig. 10H) *Hemicnetha* Enderlein, 1934 (*brachycladum* species group)
Gonapophysis less than twice as long as wide, and blunt apically (Figs. 12H-I); cibarium with conspicuous teeth arranged along border to base of cornuae (Figs. 10I-J) *Trichodagmia* Enderlein, 1934

3.2. Males

1. General coloration black, sometimes abdomen with greenish tones; with or without setae on basal sector of R 2
General coloration variable, from dark brown to yellowish, generally with setae on basal sector of R 23
- 2(1). With setae on basal sector of R 3
Without setae on basal sector of R 11
- 3(2). Gonostylus longer than gonocoxite (Figs. 22D-E); ventral plate relatively short and with or without median process and carina (Figs. 19F-H) 4
Gonostylus shorter than gonocoxite; ventral plate of variable shape 5
- 4(3). Scutum with setae homogeneously arranged and without silvery spots; ventral plate distally with small and median, but not lateral, processes (Fig. 19E) *Trichodagmia* Enderlein, 1934
Scutum with setae disposed in groups, like scales, and with 1+1 silvery anterolateral spots sometimes continued sub-medially, reaching the posterior grayish area (Figs. 15H-I); ventral plate distally with large median and lateral processes (Figs. 19F-G) *Thrysopelma* Enderlein, 1934
- 5(3). Distal spur of gonostylus much reduced or absent (Figs. 20C-F) (*Chirostilbia* Enderlein, 1921) 6
Gonostylus with distal spur 7
- 6(5). Gonostylus without lateral crest (Figs. 20C-D) *Chirostilbia* Enderlein, 1921 (*pertinax* species group)
Gonostylus with lateral crest (Figs. 20E-F) *Chirostilbia* Enderlein, 1921 (*subpallidum* species group, part)
- 7(5). Scutum without ornamentation; gonostylus flattened and curved, with sub-triangular internal lobe
Eusimulium Roubaud, 1906 and *Nevermannia* Enderlein, 1921
Scutum, when illuminated anteriorly, generally ornamented with spots; gonostylus variously shaped 8
- 8(7). Gonostylus (microscope slide mounted) sub-quadrata, with distal concavity and sub-apical spur (Figs. 23A-D) (*Pternaspatha* Enderlein, 1930) 9
Gonostylus (microscope slide mounted) sub-triangular to sub-trapezoidal or sub-cylindrical and with apical spur 10
- 9(8). Gonostylus with tubercles (Figs. 23A-B) *Pternaspatha* Enderlein, 1930 (*nigristrigatum* species group)
Gonostylus without tubercles (Figs. 23C-D) *Pternaspatha* Enderlein, 1930 (*nemorale* species group)
- 10(8). Gonostylus sub-triangular or sub-trapezoidal, with 1 apical spur (Figs. 23G-H); ventral plate without lateral constriction (Figs. 18D-F) *Inaequalium* Coscarón & Wygodzinsky, 1984
Gonostylus sub-cylindrical, with weak distal curvature and 1 apical spur (Figs. 20I-K) (except *wolffhuegeli*, in which it is relatively short and has several apical spurs) (Fig. 20L); ventral plate with weak lateral constriction *Ectemnaspis* (*bicoloratum* species group; *romanoi* species group)

- 11(2). Gonostylus much shorter than gonocoxite 12
 Gonostylus as long as or longer than gonocoxite 19
- 12(11). Gonostylus without apical spur 13
 Gonostylus with apical spur 14
- 13(12). Scutum without ornamentation; gonostylus sub-ovoid, with reinforced edge (Figs. 23E-F) ... *Notolepria* Enderlein, 1930
 Scutum with 1+1 sub-median, anterior, silvery spots (Fig. 15A); gonostylus sub-trapezoidal, with sub-lateral crest (Figs. 20E-F) *Chirostilbia* Enderlein, 1921 (*subpallidum* species group)
- 14(12). Scutum with 1+1 sub-median silvery vittae, reaching or not the grayish posterior area (Figs. 15B-C); gonostylus sub-triangular or sub-trapezoidal 15
 Scutum without 1+1 sub-median silvery vittae, but with anterior sub-quadrangular or sub-triangular silvery spots (Figs. 15D-G); gonostylus sub-rectangular or sub-cylindrical 17
- 15(14). Gonostylus about twice as long as wide at base; apical spur spatuliform (Figs. 23K-L)
 *Coscaroniellum* Py-Daniel, 1983
 Gonostylus only slightly longer than wide at base; terminal spur sub-conical 16
- 16(15). Gonostylus sub-triangular with apical or sub-apical spur (Figs. 23J, M)
 *Byssodon* Enderlein, 1925, *Cerqueirellum* Py-Daniel, 1983, *Coscaroniellum* Py-Daniel, 1983 (*quadrivittatum* species group) and *Psaroniocompsa* Enderlein, 1934 (*siolii* species group)
 Gonostylus sub-quadrangular or sub-trapezoidal, with sub-median spur (Figs. 23I)
 *Psaroniocompsa* Enderlein, 1934 (*incrustatum* species group)
- 17(14). Gonostylus sub-cylindrical, generally with weak dorsal curvature and 1 apical spur (Figs. 16C, 20I-K, 21A-F, 22A)
 *Ectemnaspis* Enderlein, 1914 (*bicoloratum* species group, *romanai* species group), *Psilopeltmia* Enderlein, 1934 and *Hemicnetha* Enderlein, 1934 (*oviedoii* species group)
 Gonostylus sub-trapezoidal, with 2-15 marginal spurs 18
- 18(17). Ventral plate over twice as wide as long; gonostylus with 7-15 marginal spurs (Fig. 20L)
 *Ectemnaspis* Enderlein, 1915 (*romanai* species group)
 Ventral plate as wide as long; gonostylus with 2-4 sub-apical spurs *Psilozia* Enderlein, 1936
- 19(11). Gonostylus sub-cylindrical, generally less than half width of gonocoxite, distal edge wide and generally with basal process 20
 Gonostylus wider than half width of gonocoxite, distal edge wide and without basal process 21
- 20(19). Ventral plate generally becoming narrower apicomediately (Figs. 18J-K); gonostylus with basal process present (Figs. 21H-J) *Aspathia* Enderlein, 1935
 Ventral plate not becoming narrower apicomediately (Fig. 19B-C); gonostylus without basal process, but with small sub-basal carina (Fig. 22B) *Hearlea* Vargas, Martínez Palacios & Díaz Nájera, 1946
- 21(19). Gonostylus flattened and sinuous, scarcely narrowed distally (Figs. 21K-L); endoparameres with large teeth and well sclerotized base (Fig. 17J) (*Hemicnetha* Enderlein, 1934) 22
 Gonostylus sub-triangular, narrowed and well curved apically (Figs. 22D-E); endoparameres without teeth and base not well sclerotized *Thrysopelma* Enderlein, 1934
- 22(21). Ventral plate wider than long, with a large median process (Figs. 18B, 19A)
 *Hemicnetha* Enderlein, 1934 (*paynei* species group, *brachycladum* species group)
 Ventral plate about as long as wide, without median process but with a median carina (Figs. 19B)
 *Hemicnetha* Enderlein, 1934 (*mexicanum* species group)
- 23(1). Gonostylus shorter or slightly longer than gonocoxite 24
 Gonostylus almost twice as long as gonocoxite 28
- 24(23). Gonostylus with 3 to 6 or more spurs (Fig. 22C); endoparameres without teeth (Fig. 17K)

.....	<i>Trichodagmia</i> Enderlein, 1934 (part)
Gonostylus with 1 or no spurs; endoparameres generally with large teeth (Fig. 16C)	25
25(24). Basal sector of R without setae; scutum dull grayish; gonostylus with internal border straight (Fig. 21G)	
..... <i>Psilopemia</i> Enderlein, 1934 (<i>blancasi</i> species group)	
Basal sector of R with hair (Fig. 3F); scutum not grayish; gonostylus with internal border curved	26
26(25). Gonostylus without apical spur, with sub-lateral crest and not curved distally (Figs. 20E-F)	
..... <i>Chirostilbia</i> Enderlein, 1921 (<i>subpallidum</i> species group)	
Gonostylus with apical spur, without sub-lateral crest and slightly curved distally	27
27(26). Ventral plate generally longer than wide at base; gonostylus without distal curvature (Figs. 21A-B)	
..... <i>Ectemnaspis</i> Enderlein, 1914 (<i>perflavum</i> species group)	
Ventral plate generally shorter than wide at base (Figs. 18G-F); gonostylus generally with distal curvature (Figs. 21C-F)	
..... <i>Psilopelmia</i> Enderlein, 1934 and <i>Ectemnaspis</i> Enderlein, 1914 (<i>dinellii</i> species group)	
28(23). Scutum dark brown, with setae grouped like scales; gonostylus relatively thick distally (Figs. 22D-E)	
..... <i>Thrysopelma</i> Enderlein, 1934	
Scutum reddish-brown, with setae not grouped like scales; gonostylus relatively thin distally (Fig. 22C)	
..... <i>Trichodagmia</i> Enderlein, 1934	

3.3. Pupae

1. Gill arborescent, with relatively thick branches, apices acute, well sclerotized and spine-shaped, with 12-50 branches (Figs. 28H-I); cocoon compact and with ventral side projected anteriorly	
..... <i>Thrysopelma</i> Enderlein, 1934 (part)	
Gills with variable shape, but without acute, well-sclerotized, spine-shaped apices; cocoon of variable shape ...	2
2(1). Gills thick, asymmetrical, generally with curvature (Figs. 24G-H, 25G, 28E-F)	3
Gills generally filiform, symmetrical, without lateral curvature	6
3(2). Frontoclypeus and exposed portion of thorax with simple trichomes (Fig. 26K); gills with striations (Figs. 28E-F); cocoon without dorsal projection (Fig. 25G)	
..... <i>Hearlea</i> Vargas, Martínez Palacios & Díaz Nájera, 1946	
Frontoclypeus and exposed portion of thorax with multibranched trichomes with 2-8 branches (Figs. 26F-G); gill without striations; cocoon generally with dorsal projection	4
4(3). Cocoon generally without dorsal projection, anterior edge straight (Figs. 24G-H)	
..... <i>Inaequalium</i> Coscarón & Wygodzinsky, 1984	
Cocoon projected dorsally (Figs. 24I, 25A)	5
5(4). Frontoclypeus and thorax with bifid trichomes (Figs. 26F)	
..... <i>Psaroniocompsa</i> Enderlein, 1934	
Frontoclypeus and thorax with 3-8 branched trichomes (Fig. 26G)	
..... <i>Ectemnaspis</i> Enderlein, 1914 and <i>Aspathia</i> Enderlein, 1935	
6(2). Cocoon with compact weave and anterior portion elevated, generally protecting gill base (Fig. 25D); gill filaments relatively thick	7
Cocoon without compact weave and generally with anterior portion not elevated or protecting gill; gill filaments relatively thin	13
7(6). Anterior opening of cocoon with festoons protecting gill base (Figs. 25D, H)	8
Anterior opening of cocoon without festoons protecting gill base	9
8(7). Gill with 8 branches, apices blunt and resistant (Figs. 28D); frontoclypeus generally reinforced at base and enlarged at facial area, with female and male having similar facial area (Figs. 26H-I)	
..... <i>Hemicnetha</i> Enderlein, 1934 (<i>paynei</i> species group)	
Gill with 10 branches, frequently with blunt membranous apices, not resistant (Fig. 28A); frontoclypeus not	

Manual of Neotropical Diptera. Simuliidae

- reinforced at base and not enlarged at facial area, male frons basally narrower than in female (Fig. 26E) *Chirostilbia* Enderlein, 1921 (part)
- 9(7). Gill with 6 filaments 10
Gill with more than 6 filaments 11
- 10(9). Frontoclypeus and thorax with numerous tubercles, some of them acute and like short spines (Figs. 26J); cocoon scarcely elevated anteriorly (Fig. 25F) *Hemicnetha* Enderlein, 1934 (*oviedoii* species group)
Frontoclypeus and thorax without tubercles (Fig. 26I); cocoon frequently very elevated anteriorly (Fig. 25E) *Hemicnetha* Enderlein, 1934 (*brachycladum* species group)
- 11(9). Gill with 18-20 branches arranged in a bunch (Fig. 28G) *Trichodagmia* Enderlein, 1934 (part)
Gill with 8-16 branches 12
- 12(11). Gill with 12 branches
..... *Hemicnetha* Enderlein, 1934 (*mexicanum* species group) and *Trichodagmia* Enderlein, 1934 (part)
Gill with 10 branches (Fig. 25E) *Hemicnetha* Enderlein, 1934 (*brachycladum* species group, part)
- 13(6). Gill with 4 filamentous branches 14
Gill with more than 4 filamentous branches 17
- 14(13). Frontoclypeus and thorax with acuminate tubercles (Fig. 29I) *Coscaroniellum* Py-Daniel, 1983 (part)
Frontoclypeus and thorax with tubercles not acuminate 15
- 15(14). Cocoon reduced ventrally (Fig. 25C); cephalic and thoracic trichomes simple (Fig. 29J)
..... *Psilopeltia* Enderlein, 1934 (*blancasi* species group)
Cocoon not reduced ventrally; cephalic and thoracic trichomes with 1-8 branches 16
- 16(15). Cephalic and thoracic trichomes generally bifid (Fig. 26F)
..... *Psaroniocompsa* Enderlein, 1934 and *Cerqueirellum* Py-Daniel, 1983
Cephalic and thoracic trichomes with 1-8 branches
..... *Eusimulum* Roubaud, 1906, *Byssodon* Enderlein, 1925, *Nevermannia* Enderlein, 1921 and *Psilopeltia* Enderlein, 1934
- 17(13). Gill with 6 filamentous branches 18
Gill with more than 6 filamentous branches 20
- 18(17). Over 10 thoracic trichomes on each side, frequently very modified (Figs. 29D-H)
..... *Pternaspatha* Enderlein, 1930 and *Aspathia* Enderlein, 1935 (part)
About 5 thoracic trichomes on each side 19
- 19(18). Cephalic and thoracic trichomes bifid (rarely single or trifid)
..... *Psaroniocompsa* Enderlein, 1934, *Cerqueirellum* Py-Daniel, 1983 and *Aspathia* Enderlein, 1935 (part)
Cephalic and thoracic trichomes generally with more than 3 branches, not bifid
..... *Byssodon* Enderlein, 1925, *Inaequalium* Coscarón & Wygodzinsky, 1984, *Notolepria* Enderlein, 1930,
Ectemnaspis Enderlein, 1914 (*bicoloratum* and *romanai* species groups) and *Psilopeltia* Enderlein, 1934
- 20(17). Gill with 8-10 branches 21
Gill with more than 10 branches *Psilozia* Enderlein, 1936, *Chirostilbia*
Enderlein, 1921, *Aspathia* Enderlein, 1935 and *Ectemnaspis* Endxerlein, 1914 (*perflavum* species group)
- 21(20). Over 10 thoracic trichomes, frequently much modified (Fig. 29D); gill with 8 branches *Pternaspatha* Enderlein, 1930
5 thoracic trichomes on each side; gill with 8-10 branches 22
- 22(21). Cocoon weave spongy, with elevated anterior margin and dorsal carina (Fig. 25B)
..... *Ectemnaspis* Enderlein, 1914 (*bicoloratum* and *romanai* species groups, part)
Cocoon weave compact, without elevated anterior margin or dorsal carina (Figs. 24E, J)
... *Chirostilbia* Enderlein, 1921 (*subpallidum* species group), *Notolepria* Enderlein, 1930, *Cerqueirellum* Py-Daniel,

1983, *Coscaroniellum* Py-Daniel, 1983, *Psilozia* Enderlein, 1934, *Byssodon* Enderlein, 1925, *Aspathia* Enderlein, 1935, *Ectemnaspis* Enderlein, 1914 (*perflavum* and *dinellii* species groups) and *Psilopelmia* Enderlein, 1934

3.4. Larvae

1. Anal ring with more than 150 rows of hooks 2
Anal ring with less than 120 rows of hooks 6
- 2(2). Body cuticle with lanceolate trichomes (Fig. 34I); hypostomium with median tooth and intermediate teeth not well differentiated (Figs. 35H-I); abdomen not dorsally flattened on distal portion (Fig. 31B); cervical sclerites enlarged, each more than twice as wide as long (Fig. 34J) *Thrysopelma* Enderlein, 1934
Body cuticle without lanceolate trichomes; hypostomium with median tooth and generally well differentiated intermediate teeth (Fig. 35F); abdomen dorsally flattened distally (Figs. 31A); cervical sclerites not enlarged, each less than twice wider than long (Figs. 32C, L) 3
- 3(2). Anal sclerite frequently with sclerotized reinforcement, in some cases with ventral branches enlarged and completing a ring (Figs. 36A); anterior margin of hypostomium generally arc-shaped, with lateral and intermediate teeth reduced (Fig. 35F) *Hearlea* Vargas, Martínez Palacios & Días Nájera, 1946
Anal sclerite without sclerotized reinforcement nor with ventral branches completing a ring (Fig. 36B); anterior margin of hypostomium of variable shape 4
- 4(3). Cephalic apotome darkened on distal half in form of triangle (Fig. 32F) *Chirostilbia* Enderlein, 1921
Cephalic apotome with different ornamentation 5
- 5(4). Hypostomium with lateral edges descending abruptly from lateral teeth, originating a plateau-like appearance (Fig. 35G); mandible with pre-apical tooth shorter than apical tooth (Fig. 34H)
..... *Hemicnetha* Enderlein, 1934 (*brachycladum* species group) and *Trichodagmia* Enderlein, 1934
Hypostomium with lateral borders gradually descending from the lateral teeth in a curved manner; mandible with pre-apical tooth as long as or longer than apical tooth (Fig. 34F)
..... *Hemicnetha* Enderlein, 1934 (*paynei* and *mexicanum* species groups)
- 6(1). Anal ring with 100-120 rows of hooks; cephalic fan rays with setae of similar size (Fig. 33N); mandible with a pair of large marginal teeth (Fig. 34G) *Hemicnetha* Enderlein, 1934 (*oviedoii* species group)
Anal ring with less than 100 rows of hooks; cephalic fan rays alternating between groups of small setae and some larger ones; mandible with only 1 marginal tooth, if 2, the second very small (Figs. 34D-E) 7
- 7(6). Cephalic apotome without ornamentation or with a small spot on the median basal area (Fig. 32E); with small, numerous and short setae on cephalic capsule; generally with dark band around first abdominal segment; antenna thin and surpassing apex of cephalic fan stem (Fig. 34E) 8
Cephalic apotome with evident ornamentation; without or with very scarce setae on cephalic capsule; antenna generally thick and surpassing or not apex of cephalic fan stem 9
- 8(7). Mandibles with a row of inferior supramarginal setae (Fig. 34D); medial antennal segment as long as or longer than proximal segment (Figs. 33J) *Cerqueirellum* Py-Daniel, 1983
Mandible without a row of inferior supramarginal setae; medial antennal segment about 1/2 to 1/4 length of proximal segment (Fig. 33K) *Coscaroniellum* Py-Daniel, 1983
- 9(7). Postgenal cleft scarcely developed, with postgenal bridge longer than hypostomium length (Fig. 31D); rectal papillae without diverticula on lobes (Fig. 36G) 10
Postgenal cleft well developed, with postgenal bridge as long as or shorter than hypostomium length (Fig. 31E); rectal papillae generally with diverticula on lobes (Fig. 36F) 13
- 10(9). Antennae with thin transverse striations and surpassing apex of cephalic fan stem; medial antennal segment longer than proximal segment (Fig. 33M) *Aspathia* Enderlein, 1935
Antennae without striations and not surpassing apex of cephalic fan; medial antennal segment shorter than proximal segment 11

Manual of Neotropical Diptera. Simuliidae

- 11(10). Ninth sternite without 1+1 tubercles *Psilopeltia* Enderlein, 1934 (*blancasi* species group)
Ninth sternite with 1+1 conspicuous tubercles 12
- 12(11). Antennae surpassing apex of cephalic fan stem *Eusimulum* Roubaud, 1906 and *Nevermannia* Enderlein, 1921
Antennae not surpassing apex of cephalic fan stem *Psilozia* Enderlein, 1936
- 13(9). Rectal papillae without diverticula on lobes (Fig. 36G) *Psilopeltia* Enderlein, 1934 (*blancasi* species group)
Rectal papillae with diverticula on lobes (Fig. 36F) 14
- 14(13). Cephalic apotome with isolated spots: anteromedian, posteromedian, anterolateral and posterolateral spots generally present (Figs. 32C-D) 15
Cephalic apotome with variable ornamentation, different from above 19
- 15(14). Anal sclerite without scales (Fig. 36B) 16
Anal sclerite with scales 17
- 16(15). Antenna surpassing apex of cephalic fan stem (Figs. 32C); medial antennal segment generally shorter than distal segment (Fig. 33I) *Psaroniocompsa* Enderlein, 1934
Antenna not surpassing apex of cephalic fan stem; medial antennal segment generally longer than distal segment (Fig. 33L) *Ectemnaspis* Enderlein, 1914 (*bicoloratum* and *perflavum* species groups)
- 17(15). Body length less than 4.5 mm *Notolepria* Enderlein, 1930
Body length over 5.0 mm 18
- 18(17). Medial antennal segment shorter than apical segment (Fig. 33G)
..... *Chirostilbia* Enderlein, 1921 (*subpallidum* species group)
Medial antennal segment longer than apical segment (Fig. 33F)
..... *Byssodon* Enderlein, 1925 and *Pternaspatha* Enderlein, 1930 (part)
- 19 (14). Cephalic apotome darkened on basal half, leaving a light window on the median sub-basal portion (Figs. 32G-J); anal sclerite without scales
... *Ectemnaspis* Enderlein, 1914 (*bicoloratum*, *romanai* and *dinelli* species groups) and *Psilopeltia* Enderlein, 1934
Cephalic apotome with variable ornamentation, different from above; anal sclerite with or without scales 20
- 20(19). Anal sclerite with scales; medial antennal segment longer than distal segment (Figs. 33F) ... *Pternaspatha* Enderlein, 1930
Anal sclerite without scales (Fig. 36B); medial antennal segment shorter than distal segment (Figs. 33G-H) 21
- 21 (20). Cephalic apotome with a dark, median, basal, diffuse spot, that becomes thinner anteriorly, delimiting two light elongated areas (Fig. 32B) *Inaequalium* Coscarón & Wygodzinsky, 1984
Cephalic apotome with strong, basal, triangle-shaped pigmentation (Fig. 32F)
..... *Chirostilbia* Enderlein, 1921 (*pertinax* species group)

References

- Abebe, M., M. S. Cupp, F. B. Ramberg & E. W. Cupp, 1994. Anticoagulant activity in salivary gland extracts of black flies (Diptera: Simuliidae). *J. med. Ent.* 31 (6): 908-911.
- Abebe, M., J. M. Ribeiro, M. S. Cupp & E. W. Cupp, 1996. Novel anticoagulant from salivary glands of *Simulium vittatum* (Diptera: Simuliidae) inhibits activity of coagulation factor V. *J. med. Ent.* 33 (1): 173-176.
- Alencar, Y. B. A., T. A. V. Ludwig, C. C. Soares & N. Hamada, 2001. Stomach content analyses of *Simulium perflavum* Roubaud, 1905 (Diptera: Simuliidae) larvae from stream in Central Amazonia, Brazil. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 96: 571-596.
- Ambrós Ginarte, C., C. F. S. Andrade & J. C. Gaona, 2003. Larvas de simulídeos (Diptera, Simuliidae) do centro-oeste, sudeste e sul do Brasil, parasitadas por microsporídeos (Protozoa) e mermitídeos (Nematoda). *Iheringia, Zool.*, Porto Alegre 93 (3): 323-334.
- Andrade, C. F. S. & J. Campos, 1995. Efetividade de Bactivec, a base de *Bacillus thuringiensis* H-14 no controle de *Simulium pertinax* (Diptera, Simuliidae). *Revta Patol. trop.* 24: 275-281.
- Andrade, H. T. A., J. F. de Medeiros, F. A. C. Pessoa & V. Py-Daniel, 2004. Considerações sobre simulídeos (Diptera – Simuliidae) e filarioses (oncocercose e mansonelose). *Carpe Diem*, Natal 2/3: 91-198.
- Andrade, H. T. A. & V. Py-Daniel, 2000. A fauna associada a Simuliidae (Diptera: Culicomorpha) em três bacias hidrográficas do estado do Rio Grande do Norte. *Revta Ecol. aquát. trop.*, Natal 10: 71-76.
- Andrade, H. T. A., S. Trivinho-Strixino, V. Py-Daniel & J. F. de Medeiros, 2004. Dinâmica sazonal de imaturos de *Hemicnetha brachyclada* (Lutz & Pinto, 1931) (Diptera, Simuliidae) em um criadouro permanente em área do semi-árido no Nordeste brasileiro, pp. 45-52, in Chellappa, N. T., S. Chellappa & J. Z. de O. Passavante, orgs. *Ecologia aquática tropical*. Natal.
- Andreazze, R. & V. Py-Daniel, 2001. Atividade hematofágica e infecção natural de *Psaroniocompsa incrustata* (Lutz, 1910) (Diptera, Culicomorpha, Simuliidae) vetor da *Onchocerca volvulus* (Leuckart, 1893) em Xitei/Xidea, área indígena Yanomami, Roraima, Brasil. *Entomología y Vectores*, Rio de Janeiro 8 (3): 317-329.
- Andreazze, R., V. Py-Daniel & J. F. de Medeiros, 2002. Influência dos fatores climáticos na atividade hematofágica de *Psaroniocompsa incrustata* (Lutz, 1910) (Diptera, Simuliidae) vetor de *Onchocerca volvulus* (Leuckart, 1893) em Xitei/Xidea, área indígena Yanomami, Roraima, Brasil. *Entomología y Vectores*, Rio de Janeiro 9 (4): 559-557.
- Anonymous, 1997. Onchocerciasis. Report from the Inter American Conference on Onchocerciasis in Oaxaca, Mexico. *Wkly epidemiol. Rec.* 72 (29): 215-218.
- Aoki, V., E. A. Rivitti, L. M. Ito, G. Hans-Filho & L. A. Diaz, 2005. Perfil histórico da imunopatogenia do pênfigo foliáceo endêmico (fogo selvagem). *An. bras. Dermatol.* 80 (3): 287-292.
- Araújo-Coutinho, C. J. P. C., R. Figueiró, A. P. Viviani, E. S. Nascimento & C. F. G. Cavados, 2005. Abioassay method for black flies (Diptera: Simuliidae) using larvicides. *Neotrop. Ent.* 34 (3): 511-513.
- Araújo-Coutinho, C. J. P. C., E. S. Nascimento, R. Figueiró & J. J. Becnel, 2004. Seasonality and prevalence rates of microsporidia in *Simulium pertinax* (Diptera: Simuliidae) larvae in the region of Serra dos Órgãos, Rio de Janeiro, Brazil. *J. invertebr. Pathol.* 85 (3): 188-191.
- Arzube, M. & A. J. Shelley, 1990. Seasonal variation in onchocerciasis transmission in the Santiago focus of Ecuador. *Trop. Med. Parasitol.* 41 (3): 286-188.
- Barreto, P., H. Trapido & V. H. Lee, 1970. Onchocerciasis in Colombia. Entomologic findings in the first observed focus. *Amer. J. trop. Med. Hyg.* 19 (3): 837-841.
- Basáñez, M. G. & M. Boussinesq, 1999. Population biology of human onchocerciasis. *Philos. Trans. r. Soc. London (B)* 354: 809-826.
- Basáñez, M. G., M. Boussinesq, J. Proud'hon, H. Frontado, N. J. Villamizar, G. F. Medley & R. M. Anderson, 1994. Density-dependent processes in the transmission of human onchocerciasis: Intensity of microfilariae in the skin and their uptake by the simuliid host. *Parasitology* 108 (1): 115-127.
- Basáñez, M. G., R. C. Collins, C. H. Porter, M. P. Little & D. Brandling-Bennett, 2002. Transmission intensity and the patterns of *Onchocerca volvulus* infection in human communities. *Amer. J. trop. Med. Hyg.* 67 (6): 669-679.
- Basáñez, M. G., J. H. F. Remme, E. S. Alley, O. Bain, A. J. Shelley, G. F. Medley & R. M. Anderson, 1995. Density-dependent processes in the transmission of human onchocerciasis: Relationship between the numbers of microfilariae ingested and successful larval development in the simuliid vector. *Parasitology* 110: 409-427.
- Basáñez, M. G., M. A. Rodríguez-Pérez, F. Reyes-Villanueva, R. C. Collins & M. H. Rodríguez, 1998. Determination of sample sizes for the estimation of *Onchocerca volvulus* (Filarioidea: Onchocercidae) infection rates in biting populations of *Simulium ochraceum* s. l. (Diptera: Simuliidae) and its application to ivermectin control programs. *J. med. Ent.* 35 (5): 745-757.
- Basáñez, M. G., H. Townsend, J. R. Williams, H. Frontado, N. J. Villamizar & R. M. Anderson, 1996. Density-dependent processes in the transmission of human onchocerciasis: Relationship between microfilarial intake and mortality of the simuliid vector. *Parasitology* 113: 331-355.
- Basáñez, M. G., L. Yarzábal, H. L. Frontado & N. J. Villamizar, 2000. *Onchocerca-Simulium* complexes in Venezuela: Can human onchocerciasis spread outside its present endemic areas? *Parasitology* 120 (2): 143-160.

- Basáñez, M. G., L. Yarzábal, H. Takaoka, H. Suzuki, S. Noda & I. Tada, 1988. The vectorial role of several blackfly species (Diptera: Simuliidae) in relation to human onchocerciasis in the Sierra Parima and Upper Orinoco regions of Venezuela. *Ann. trop. Med. Parasitol.* 82 (6): 597-611.
- Benchimol, J. L. & M. R. Sá, eds., 2006. *Adolpho Lutz. Obra completa. 4 (2). Entomologia*. Editora Fiocruz, Rio de Janeiro.
- Bequaert, J. C., 1934. Part III. Notes on the black-flies or Simuliidae, with special reference to those of the *Onchocerca* region of Guatemala, pp. 175-224, Figs. 93-99, in Strong, R. P., J. H. Sandground, J. C. Bequaert, & M. Muñoz Ochoa, Onchocerciasis, with special reference to the Central American form of the disease. *Contr. Dept. trop. Med. & Inst. trop. Biol. Med.* 6: 234 pp., 103 Figs., 2 tables, 1 map. Harvard University Press, Cambridge.
- Bernardo, M. J. & E. W. Cupp, 1986. Rearing black flies (Diptera: Simuliidae) in the laboratory: Mass-scale in vitro membrane feeding and its application to collection of saliva and to parasitological and repellent studies. *J. med. Ent.* 23 (6): 666-679.
- Botto, C., 1990. La oncocercosis el el norte y en el foco del Alto Orinoco en Venezuela: Un estudio comparativo. *Revta mexic. Parasitol.* 3: 413.
- Botto, C., E. Escalona, S. Vivas-Martínez, V. Behm, L. Delgado & P. Coronel, 2005. Geographical patterns of onchocerciasis in southern Venezuela: Relationships between environment and infection prevalence. *Parasitología* 47 (1): 145-150.
- Botto, C., A. J. Gillespie, S. Vivas-Martínez, N. Martínez, S. Planchart, M. G. Basáñez & J. E. Bradley, 1999. Onchocerciasis hyperendemic in the Unturán Mountains: The value of recombinant antigens in describing a new transmission area in southern Venezuela. *Trans. r. Soc. trop. Med. Hyg.* 93 (1): 25-30.
- Branding-Bennett, A. D. & R. F. Darsie, 1983. Distribution of microfilariae in Guatemalans with onchocerciasis. *Trans. r. Soc. trop. Med. Hyg.* 77 (2): 254-258.
- Briceño Iragorry, L. & I. Ortiz, 1957. Los simúlidos de Venezuela. (Importancia médica. Morfología y sistemática. Distribución geográfica). *Boln venezolano Labor. clín.* 2 (1-2): 23-57.
- Calvão-Brito, R. H. S., E. M. Mokrabe, M. Maia-Herzog, R. P. Mello & V. P. Silva Jr., 1998. Oncocercose eqüina: Diagnóstico e verificação da hematofagia por simulídeos e culicóides, prováveis vetores no estado do Rio de Janeiro, Brasil. *Revta bras. Zool.*, Curitiba 15 (3): 583-587.
- Camino, N. B., 1985. Estudio de cuatro especies del género *Mesomermis* Daday, 1911 (Nematoda: Mermithidae), parásitas de larvas de simúlidos (Diptera: Simuliidae). *Revta Mus. La Plata, Zool.* 14 (150): 1-19.
- Camino, N. B., 1986. Observaciones sobre el desarrollo embrionario y el estado infectante de *Mesomermis subandina* Camino, 1985 (Nematoda: Mermithidae), parásito de larvas de simúlidos. *Spheniscus* 4: 1-7.
- Camino, N. B., 1988. Descripción de *Octomyomermis bonaerensis* sp. n. (Nematoda: Mermithidae), parásito de larvas de *Simulium bonaerense* Coscarón & Wygodzinsky (Diptera: Simuliidae). *Revta ibér. Parasitol.* 48 (2): 183-186.
- Camino, N. B., 1990. *Mesomermis delponteiana* sp. n. (Nematoda: Mermithidae) parásito de larvas de *Simulium delponteianum* Wygodzinsky (Diptera: Simuliidae) em Argentina. *Revta ibér. Parasitol.* 50 (3-4): 273-276.
- Camino, N. B., 1991a. Presencia de tres especies de mermítidos (Nematoda) parasitando a larvas de *Simulium lahillei* Peterson & Shannon, y descripción de *Mesomermis nortensis* sp. n. en la provincia de Tucumán. *Neotropica*, La Plata 37 (97): 3-7.
- Camino, N. B., 1991b. *Gastromermis cordobensis* n. sp. (Nematoda: Mermithidae) parasitizing *Simulium lahillei* Paterson & Shannon (Diptera: Simuliidae) in Argentina. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 86 (2): 223-227.
- Camino, N. B., 1992. *Octomyomermis longispiculus* sp. n. (Nematoda: Mermithidae), parásita de *Simulium wolffhuegeli* (Enderlein) (Diptera: Simuliidae). *Neotropica*, La Plata 38 (100): 105-109.
- Camino, N. B., 1993a. Two new mermithids (Nematoda: Mermithidae) parasites of *Simulium wolffhuegeli* (Enderlein) and *S. jujuyense* (Paterson & Shannon) (Diptera: Simuliidae) in Argentina. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 88 (4): 571-575.
- Camino, N. B., 1993b. A new species of the genus *Bathymermis* Daday, 1911 (Nematoda: Mermithidae) parasitizing blackfly larvae (Diptera: Simuliidae) in Argentina. *Research and Reviews in Parasitology*, Valencia 53 (3-4): 125-127.
- Camino, N. B., 1994. *Mesomermis ornatissima* n. sp. (Nematoda: Mermithidae), a parasite of *Simulium bonaerense* Coscarón et Wygodzinsky (Diptera: Simuliidae) in Argentina. *Research and Reviews in Parasitology*, Valencia 54 (1): 29-31.
- Camino, N. B. & G. O. Poinar Jr., 1988. *Ditremamermis simuliae* gen. n., sp. n. (Nematoda: Mermithidae), parásita de *Simulium bonaerense* Coscarón & Wygodzinsky (Diptera: Simuliidae) en Argentina. *Neotropica*, La Plata 34 (92): 93-97.
- Camino, N. B. & L. C. de Villalobos, 1997. A new species of *Gastromermis* Micoletzky, 1923 (Nematoda: Mermithidae), a parasite of *Simulium pertinax* Kollar (Diptera: Simuliidae) in Argentina. *Nematol. mediterr.* 25: 105-108.
- Campbell, C. C., R. C. Collins, A. Y. Huong & H. E. Marroquin, 1980. Quantitative aspects of the infection of *Simulium ochraceum* by *Onchocerca volvulus*: The relation of skin microfilarial density to vector infection. *TropenMed. Parasitol.* 31 (4): 475-478.
- Campos Gaona, J. C. & C. F. S. Andrade, 1999. Aspectos da entomologia médica e veterinária dos borrachudos (Diptera, Simuliidae) – Biologia, importância e controle.

- Lecta*, Bragança Paulista 17 (1): 51-65.
- Campos Gaona, J. & C. F. S. Andrade, 2001. Considerações sobre os simulídeos (Diptera, Nematocera) e seu controle. *Entomología y Vectores*, Rio de Janeiro 8: 27-50.
- Carabin, H., M. Escalona, C. Marshall, S. Vivas Martínez, C. Botto, L. Joseph & M. G. Basáñez, 2003. Prediction of community prevalence of human onchocerciasis in the Amazonian onchocerciasis focus: Bayesian approach. *Bull. World Health Org.* 81: 482-490.
- Castello-Branco Jr., A., 1999. Effects of *Polydispyrenia similis* (Microspora; Duboscquiidae) on development of the gonads of *Simulium pertinax* (Diptera: Simuliidae). *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 94: 421-424.
- Castello-Branco Jr., A. & C. F. S. Andrade, 1992. Susceptibility of *Simulium (Chirostilbia) pertinax* Kollar, 1832 (Culicomorpha, Simuliidae) to *Bacillus thuringiensis* var. *israelensis* in an atypical breeding habitat. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 87: 317-318.
- Castello-Branco Jr., A. & C. F. S. Andrade, 1993. Studies on *Polydispyrenia simulii* (Microspora; Pleistophoridae) in *Simulium petrinax* (Diptera: Simuliidae) in Brazil. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 88: 167.
- Cavados, C. F., R. N. Fonseca, J. Q. Chaves, C. J. Araújo-Coutinho & L. Rabinovitch, 2005. A new black fly isolate of *Bacillus thuringiensis* autoagglutination strain highly toxic to *Simulium pertinax* (Kollar) (Diptera, Simuliidae) larvae. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 100 (7): 795-797.
- Cavados, C. F. G., S. Majerowicz, J. Q. Chaves, C. J. P. C. Araújo-Coutinho & L. Rabinovitch, 2004. Histopathological and ultrastructural effects of endotoxins of *Bacillus thuringiensis* Serovar *israelensis* in the midgut of *Simulium pertinax* larvae (Diptera, Simuliidae). *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 99 (5): 493-498.
- Cerdeira, N. L., 1959. *Sobre a transmissão da Mansonella ozzardi*, 29 pp. Instituto Nacional de Pesquisas da Amazônia, Manaus.
- Charalambous, M., S. Lowell, M. Arzube & C. A. Lowry, 2005. Isolation by distance and a chromosomal cline in the Cayapa cytospecies of *Simulium exiguum*, the vector of human onchocerciasis in Ecuador. *Genetica* 124 (1): 41-59.
- Charalambous, M., C. A. Lowry, S. Lowell, A. J. Shelley & M. Arzube, 1997. The value of larval head pattern for differentiating *Simulium exiguum* s. l. and *S. gonzalezi* (Diptera: Simuliidae) in the onchocerciasis focus of Ecuador. *Bull. ent. Res.* 87: 19-24.
- Charalambous, M., P. D. Ready, A. J. Shelley, M. Arzube & C. A. Lowry, 1993. Cytological and isoenzyme analysis of the Bucay and Quevedo cytotypes of the onchocerciasis vector *Simulium exiguum* (Diptera, Nematocera) in Ecuador. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 88: 39-48.
- Charalambous, M., A. J. Shelley & M. Arzube, 1993. Distribution and taxonomic status of chromosomal forms of the onchocerciasis vector *Simulium exiguum* (Diptera: Simuliidae) in Central Ecuador. *Med. vet. Ent.* 7: 299-303.
- Charalambous, M., A. J. Shelley & M. Arzube, 1997. The potential for dispersal of onchocerciasis in Ecuador in relation to the distribution of the vector *Simulium exiguum* (Diptera: Simuliidae). *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 92 (2): 153-156.
- Charalambous, M., A. J. Shelley, M. Maia-Herzog & A. P. A. L. Dias, 1993. Chromosome variation in the onchocerciasis vector blackfly *Simulium guianense* in Brazil. *Brit. Simuliid Group Bull.* 6: 13-15.
- Charpentier, G., C. Back, S. Garzon & H. Strykowski, 1986. Observation on a new intranuclear virus-like particle infecting larvae of a black fly *Simulium vittatum* (Diptera: Simuliidae). *Diseases aquat. Organisms* 1: 147-150.
- Charalambous, M., A. J. Shelley, M. Maia-Herzog & A. P. A. L. Dias, 1996a. Four new cytotypes of the onchocerciasis vector blackfly *Simulium guianense* in Brazil. *Med. vet. Ent.* 10: 111-120.
- Charalambous, M., A. J. Shelley, M. Maia-Herzog & A. P. A. L. Dias, 1996b. Cytotype of the blackfly *Simulium guianense*, vector of onchocerciasis in Brazil. *Med. vet. Ent.* 10: 120-131.
- Collins, R. C., 1979a. Onchocerciasis transmission potentials of four species of Guatemalan Simuliidae. *Amer. J. trop. Med. Hyg.* 28 (1): 72-75.
- Collins, R. C., 1979b. Development of *Onchocerca volvulus* in *Simulium ochraceum* and *Simulium metallicum*. *Amer. J. trop. Med. Hyg.* 28 (3): 491-495.
- Collins, R. C., C. C. Campbell, D. P. Wilton & L. Newton, 1977. Quantitative aspects of the infection of *Simulium ochraceum* by *Onchocerca volvulus*. *TropenMed. Parasitol.* 28 (2): 235-243.
- Collins, R. C., T. Lehmann, J. C. Vieira Garcia & R. H. Guderian, 1995. Vector competence of *Simulium exiguum* for *Onchocerca volvulus*: Implications for the epidemiology of onchocerciasis. *Amer. J. trop. Med. Hyg.* 52 (3): 213-218.
- Collins, R. C., J. O. Ochoa, E. W. Cupp, C. González-Peralta & C. H. Porter, 1992. Microepidemiology of onchocerciasis in Guatemala: Dispersal and survival of *Simulium ochraceum*. *Amer. J. trop. Med. Hyg.* 47 (2): 147-155.
- Cordeiro, N. S. & A. Castello-Branco Jr., 1988. Developmental cycle and histopathological studies of *Thelohania sp.* (Microsporida: Thelohaniidae) in blackfly larvae (Diptera: Simuliidae). *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 83: 232.
- Corredor, A., R. Santiago Nicholls, S. Duque, P. Muñoz de Hoyos, C. A. Alvarez, R. H. Guderian, H. H. Lopez & G. I. Palma, 1998. Current status of onchocerciasis in Colombia. *Amer. J. trop. Med. Hyg.* 58 (5): 594-598.
- Coscarón, S., 1963. Dípteros hematófagos de Tierra del Fuego, posibles vectores de la mixomatosis del conejo europeo *Oryctolagus cuniculus* (L.). *Revta Fac. Cienc. Veter.*, La Plata 5 (13): 135-139. [Simuliidae, p. 137].
- Coscarón, S., 1971. Estado actual de las enfermedades trans-

- mitidas por simúlidos. *Revta Soc. ent. Peru* 6(2): 80-84.
- Coscarón, S. & C. L. Coscarón Arias, 2007. *Neotropical Simuliidae (Diptera: Insecta)/ Simuliidae neotropicales (Diptera: Insecta)*, in Adis, J., J. R. Arias, G. Rueda-Delgado & K. M. Wantzen, eds., *Aquatic biodiversity in Latin America/ Biodiversidad acuática en América Latina*. Vol. 3. Pensoft Publishers, Sofia-Moscow.
- Coscarón, S. & Miranda Esquivel, 1998a. *Pedrowygomyia*, a new Neotropical Prosimuliini genus (Diptera: Simuliidae): *Gigantodax* s. lat. split into two genera. *Ent. scand.* 29 (2): 161-168, 1 fig.
- Crosskey, R. W., 1990. *The natural history of blackflies*, 711 pp. J. Wiley, Chichester.
- Cupp, E. W., B. O. Duke, C. D. Mackenzie, J. R. Guzmán, J. C. Vieira, J. Méndez-Galván, J. Castro, F. Richards, M. Sauerbrey, A. Domínguez, R. R. Eversole & M. S. Cupp, 2004. The effects of long-term community level treatment with ivermectin (Mctizan) on adult *Onchocerca volvulus* in Latin America. *Amer. J. trop. Med. Hyg.* 71 (5): 602-607.
- Cupp, E. W., C. J. Mare, M. S. Cupp & F. B. Ramberg, 1992. Biological transmission of vesicular stomatitis virus (New Jersey) by *Simulium vittatum* (Diptera: Simuliidae). *J. med. Ent.* 29 (2): 137-140.
- Cupp, E. W., J. O. Ochoa, R. C. Collins, M. S. Cupp, C. González-Peralta, J. Castro & G. Zea-Flores, 1992. The effects of repetitive community-wide ivermectin treatment on transmission of *Onchocerca volvulus* in Guatemala. *Amer. J. trop. Med. Hyg.* 47 (2): 170-180.
- Cupp, E. W., A. O. Ochoa, R. C. Collins, F. R. Ramberg & G. Zea, 1989. The effect of multiple ivermectin treatment on infection of *Simulium ochraceum* with *Onchocerca volvulus*. *Amer. J. trop. Med. Hyg.* 40 (5): 501-506.
- Cupp, M. S., Y. Chen & E. W. Cupp, 1997. Cellular hemolymph response of *Simulium vittatum* (Diptera: Simuliidae) to intrathoracic injection of *Onchocerca lienalis* (Filarioidea: Onchocercidae) microfilariae. *J. med. Ent.* 34 (1): 56-63.
- Cupp, M. S., E. W. Cupp, J. O. Ochoa & J. K. Moulton, 1995. Salivary apyrase in New World blackflies (Diptera: Simuliidae) and its relationship to onchocerciasis vector status. *Med. vet. Ent.* 9 (3): 325-330.
- Dalmat, H. T., 1955. The blackflies (Diptera, Simuliidae) of Guatemala and their role as vectors of onchocerciasis. *Smithson. misc. Collns* 125 (1): vii + 425 pp., 43 pls., 429 Figs., maps.
- Davies, J. B., L. Oskam, R. Luján, G. J. Schoone, C. C. Kroon, L. A. López-Martínez & A. J. Paniagua-Álvarez, 1998. Detection of *Onchocerca volvulus* DNA in pools of wild-caught *Simulium ochraceum* by use of the polymerase chain reaction. *Ann. trop. Med. Parasitol.* 92 (3): 295-304.
- De León, J. R. & B. O. Duke, 1966. Experimental studies on the transmission of Guatemalan and West African strains of *Onchocerca volvulus* by *Simulium ochraceum*, *S. metallicum* and *S. callidum*. *Trans. r. Soc. trop. Med. Hyg.* 60 (6): 735-752.
- Diaz, L. A., S. A. P. Sampaio, E. A. Rivitti, C. R. Martins, P. R. Cunha, C. Lombardi, F. A. Almeida, R. M. Castro, M. L. Macca, C. Lavrado, G. Hans-Filho, P. Borges, A. Chaul, L. Minelli, J. C. Empinotti, H. Friedman, I. Campbell, R. S. Labib & G. J. Anhalt, 1989. Endemic pemphigus foliaceus (fogo selvagem): II. Current and historic epidemiologic studies. *J. Invest. Dermatol.* 92: 4-12.
- Duke, B. O. L., 1970. *Onchocerca-Simulium* complexes. Experimental studies on the transmission of Guatemalan and West African strains of *Onchocerca volvulus* by *Simulium ochraceum*, *S. metallicum* and *S. callidum*. *Ann. trop. Med. Parasitol.* 64: 421-431.
- Edman, J. D. & K. R. Simmons, 1985. Simuliids (mainly *Simulium decorum* Walker), pp. 145-152, in Singh, P. & R. F. Moore, eds., *Handbook of insect rearing*. Vol. 2, 514 pp. Elsevier, Amsterdam.
- Elliot, M. J. & C. Potter, 1978. The future of pyrethroids in insect control. *A. Rev. Ent.* 23: 443-469.
- Figueiró, R., T. N. Docile & A. T. Aranda, 2006. An artificial breeding site for larvicide bioassay with blackfly (Diptera: Simuliidae) larvae and other macroinvertebrates from lotic systems. *Bioassay* 1 (1): 11.
- Figueiró, R., E. S. Nascimento & C. J. P. C. Coutinho, 2002. Avaliação da vazão da coluna d'água do criadouro artificial sobre a captação de partículas por larvas de *Simulium pertinax* (Diptera: Simuliidae). *Entomología y Vectores*, Rio de Janeiro 9 (2): 251-261.
- Figueroa, M. H., R. C. Collins & W. J. Kozek, 1977. Post-prandial transportation and maintenance of *Simulium ochraceum* infected with *Onchocerca volvulus*. *Amer. J. trop. Med. Hyg.* 26 (1): 75-79.
- García, J. J., 1990a. Patógenos de simúlidos netropicales (Diptera: Simuliidae): *Polydispyrenia simulii* (Lutz & Splendore, 1908) (Microspora). *Revta Soc. ent. Argent.* 48 (1-4): 85-90.
- García, J. J., 1990b. *Helmichia similiae* sp. nov. (Microspora: Thelohaniidae), una nueva especie de microsporidio patógeno de larvas de simúlidos de la República Argentina. *Neotropica*, La Plata 35 (94): 71-79.
- García, J. J., 1990c. Un nuevo microsporidio patógeno de larvas de simúlidos (Diptera, Simuliidae): *Ringueletium pillosa* gen. et sp. nov. (Microspora: Caudosporidae). *Neotropica*, La Plata 36 (96): 111-122.
- García, J. J., 1992. Patógenos de simúlidos neotropicales (Diptera: Simuliidae): *Amblyospora bracteata* (Strickland, 1913) (Microspora: Thelohaniidae). *Revta Soc. ent. argentina* 50 (1-4): 3-8.
- García, J. J., E. I. Hazard & T. Fukuda, 1989. Preliminary report of microsporidia in Simuliidae larvae from Argentina. *J. Amer. Mosquito Control Assoc.*, New Jersey 5 (1): 64-69.
- Garms, R., 1975. Observations on filarial infections and parous rates of anthropophilic blackflies in Guatemala, with reference to the transmission of *Onchocerca volvulus*. *TropenMed. Parasitol.* 26 (2): 169-182.
- Garms, R. & J. O. Ochoa, 1979. Further studies on the relative importance of Guatemalan blackfly species as vectors

- of *Onchocerca volvulus*. *TropenMed. Parasitol.* 30 (1): 120-128.
- Garrido, C. & M. Campos, 2000. First report of presumed parasitic keratitis in Indians from the Brazilian Amazon. *Cornea. The J. of Cornea ext. Dis.* 19 (6): 817-819.
- Gómez-Priego, A., R. Mendoza & J. L. de la Rosa, 2005. Prevalence of antibodies to *Onchocerca volvulus* in residents of Oaxaca, Mexico, treated for 10 years with ivermectin. *Clin. Diagn. Lab. Immunol.* 12 (1): 40-43.
- Gowtage Sequeira, S., T. Higazi, T. R. Unnasch & M. G. Basáñez, 2002. Estimating the prevalence and intensity of *Onchocerca volvulus* infection in *Simulium guianense* s. l. using the O-150 polymerase chain reaction assay. *Brit. Simuliid Group Bull.* 18: 13-15.
- Gray, E. W., P. H. Adler & R. Noblet, 1997. Economic impact of black flies (Diptera: Simuliidae) in South Carolina and development of a localized suppression program. *J. Am. Mosquito Control Ass.* 12: 676-678.
- Gray, E. W., P. H. Adler, C. L. Coscarón Arias, S. Coscarón & R. Noblet, 1999. Development of the first black fly (Diptera, Simuliidae) management program in Argentina and comparison with other programs. *J. Amer. Mosquito Control Ass.* 15 (3): 400-406.
- Grillet, M. E., M. G. Basáñez, S. Vivas-Martínez, N. Villamizar, H. Frontado, J. Cortez, P. Coronel & C. Botto, 2001. Human onchocerciasis in the Amazonian area of southern Venezuela: Spatial and temporal variations in biting and parity rates of black fly (Diptera: Simuliidae) vectors. *J. med. Ent.* 38 (4): 520-530.
- Grillet, M. E., C. Botto, M. G. Basáñez & R. Barrera, 1994. Vector competence of *Simulium metallicum* s. l. (Diptera: Simuliidae) in two endemic areas of human onchocerciasis in northern Venezuela. *Ann. trop. Med. Hyg.* 88 (1): 65-75.
- Grillet, M. E., N. J. Villamizar, J. Cortéz, H. L. Frontado, M. Escalona, S. Vivas-Martínez & M. G. Basáñez, 2005. Diurnal biting periodicity of parous *Simulium* (Diptera: Simuliidae) vectors in the onchocerciasis Amazonian focus. *Acta trop.* 94 (2): 139-158.
- Grillet, M. E., S. Vivas Martínez, N. Villamizar, H. Frontado, J. Cortez, P. Coronel & M. G. Basáñez, 2002. Spatial and seasonal variation of biting and parity rates of blackfly vectors in the Amazonian onchocerciasis focus. *Brit. Simuliid Group Bull.* 18: 19-21.
- Guderian, R. H., 1988. Effects of nodelectomy in onchocerciasis in Ecuador. *Trop. Med. Parasitol.* 39 (suppl. 4): 356-357.
- Guderian, R. H., M. Anselmi, M. Espinel, T. Mancero, G. Rivadaneira, R. Proano, H. M. Calvopina, J. C. Vieira & P. J. Cooper, 1997. Successful control of onchocerciasis with community-based ivermectin distribution in the Rio Santiago focus in Ecuador. *Trop. Med. Int. Health* 2 (10): 982-988.
- Guderian, R. H., M. Anselmi, M. Espinel, C. Sandoval, P. J. Cooper, G. Rivadaneira & R. H. Guderian, 1997. Onchocerciasis in Ecuador: Prevalence of infection on the Ecuador-Colombia border in the province of Esmeraldas. *Mems Inst. Oswaldo Cruz, Rio de Janeiro* 92 (2): 157-162.
- Guderian, R. H., B. J. Beck, D. J. Stone, K. Isabel & C. D. Mackenzie, 1988. Onchocerciasis in Ecuador: Recent observations in the province of Esmeraldas. *J. trop. Med. Hyg.* 91: 161-168.
- Guderian, R. H., R. Lovato, M. Anselmi, T. Mancero & P. J. Cooper, 1997. Onchocerciasis and reproductive health in Ecuador. *Trans. r. Soc. trop. Med. Hyg.* 91 (3): 315-317.
- Guderian, R. H. & A. J. Shelley, 1992. Onchocerciasis in Ecuador: The situation in 1989. *Mems Inst. Oswaldo Cruz, Rio de Janeiro* 87 (3): 405-415.
- Guevara, A. G., J. C. Vieira, B. G. Lilley, A. López, N. Vieira, J. Rumbea, R. Collin, C. R. Katholi & T. R. Unnasch, 2003. Entomological evaluation by pool screen polymerase chain reaction of *Onchocerca volvulus* transmission in Ecuador following mass Mectizan distribution. *Amer. J. trop. Med. Hyg.* 68 (2): 222-227.
- Hamada, N., W. L. S. Costa & S. M. Darwich, 1997. Notes on artificial substrates for black flies (Diptera: Simuliidae) larvae and microsporidian infection in Central Amazonia, Brazil. *An. Soc. ent. Brasil* 26 (3): 589-593.
- Hashiguchi, Y., M. Kawabata, S. Ito & M. M. Recinos, 1981. Limited fly load and development of *Onchocerca volvulus* microfilariae in Guatemalan *Simulium ochraceum*. *J. Helminthol.* 55 (3): 189-196.
- Hernandez, L. M., A. J. Shelley & M. Penn, 2002. Human onchocerciasis in Central America. Can it be dispersed to Belize? *Acta tropica* 83: 49-50.
- Hoffmann, C. C., 1930. Los simúlidos de la region onchocercosa [sic] de Chiapas (con descripción de nuevas especies). *An. Inst. Biol., México* 1 (4): 293-306, 15 figs.
- Hoffmann, C. C., 1931. Los simulidos de la región onchocercosa [sic] de Chiapas, segunda parte: Los estados larvales. *An. Inst. Biol., México* 2: 207-218.
- Howerth, E. W., D. G. Mead & D. E. Staalknecht, 2002. Immunolocalization of vesicular stomatitis virus in black flies (*Simulium vittatum*). *Ann. New York Acad. Sci.* 969: 340-345.
- Jamnback, H., 1973. Recent developments in control of blackflies. *A. Rev. Ent.*, Palo Alto 18: 281-304.
- Kim, K. C. & R. W. Merritt, eds., 1988. *Black flies: Ecology, population management and annotated world list*. Pennsylvania State University, University Park ("1987").
- Kozek, W. J. & C. Raccurt, 1983. Ultrastructure of *Mansonella ozzardi* microfilaria with a comparison of the South American (simuliid-transmitted) and the Caribbean (culicoid-transmitted) forms. *TropenMed. Parasitol.* 34 (1): 38-53.
- Lacey, L. A., 1981. Simulídeos antropofilicos no Parque Nacional da Amazônia (Tapajós), Brasil, com referência aos efeitos no homem. *Boln Ofic. sanit.panamer.* 90 (4): 326-338.
- Lacey, L. A. & J. D. Charlwood, 1980. On the biting activities of some anthropophilic Amazonian Simuliidae (Diptera). *Bull. Ent. Res.* 70: 495-509.
- Lacey, L. A. & A. H. Undeen, 1988. The biological control

- potential of pathogens and parasites of black flies, pp. 327-340, in Kim, K. C. & R. W. Merritt, eds., *q. v.*
- Lehmann, T., M. S. Cupp & W. E. Cupp, 1994a. *Onchocerca linealis*: Rapid clearance of microfilariae within the black fly, *Simulium vittatum*. *Amer. J. trop. Med. Hyg.* 53 (3): 243-247.
- Lehmann, T., M. S. Cupp & E. W. Cupp, 1994b. *Onchocerca lienalis*: A comparison of microfilarial loss in *Simulium jenningsi* and *Simulium vittatum*. *Experiment. Parasitol.* 79 (2): 195-197.
- Lehmann, T., S. M. Cupp & W. E. Cupp, 1995a. Chemical guidance of *Onchocerca linealis* microfilariae to the thorax of *Simulium vittatum*. *Parasitology* 110 (3): 329-337.
- Lehmann, T., M. S. Cupp & E. W. Cupp, 1995b. Analysis of migration success of *Onchocerca lienalis* microfilariae in the haemocoel of *Simulium vittatum*. *J. Helminthol.* 69 ((1)): 47-52.
- León, L. A. & P. Wygodzinsky, 1953a. Los simúlidos del Ecuador (Diptera). Su importancia en medicina humana. *Boln Inform. cient. nac.*, Quito 6 (57): 269-288, 7 Figs.
- León, L. A. & P. Wygodzinsky, 1953b. Los simúlidos del Ecuador y su importancia en medicina tropical (Diptera Simuliidae [sic!]). *Revta ecuator. Ent. Parasitol.* 1 (4): 23-39, 2 pls.
- Lewis, D. J., 1963. Simuliidae (Diptera) from the human onchocerciasis area of Venezuela. *Proc. r. ent. Soc. London* 32: 53-62.
- Lewis, D. J. & R. Ibáñez Aldecoa, 1962. Simuliidae and their relation to human onchocerciasis in northern Venezuela. *Bull. World Health Org.* 27: 449-464.
- Lichtwardt, R. W., 1997. Costa Rican gut fungi (Trichomycetes) infecting lotic insect larvae. *Revta Biol. trop.*, San José 45: 1349-1383.
- Lichtwardt, R. W., L. Ferrington Jr. & C. C. López Lastra, 1999. Trichomycetes in Argentinean aquatic insect larvae. *Mycologia*, Lawrence 91: 1069-1082.
- Lichtwardt, R. W., C. C. López-Lastra & M. G. Mazzucchelli, 2000. Fungi living in the guts of larval aquatic insects in northwestern Argentina. *Mycologia*, Lawrence 92: 332-340.
- López Lastra, C. C. & J. J. García, 1990. Primer registro de simúlidos (Diptera: Simuliidae) parasitados por *Coelomycidium simulii* Debaisieux (Chytridiomycetes. Chytridiales) em la República Argentina. *Revta Soc. ent. argent.* 48 (1-4): 91-96.
- López Lastra, C. C., A. C. Scorsetti, G. A. Martí & S. Coscarón, 2005. Trichomycetes living in the guts of aquatic insects of Misiones and Tierra del Fuego, Argentina. *Mycologia*, Lawrence 97 (2): 320-328.
- Lutz, A. & A. Splendore, 1908. Ueber Pebrine und verwandte Mikrosporidien. Zweite Mitteilung. *Centralblatt Bakteriol. ParasitKde Infektionskrankh.*, Berlin 46 (4): 311-315 [Nosema simulii, p. 312, Fig. 29]. [Fac-simile reproduction in Benchimol & Sá, 2006b: 901-905; Portuguese translation in Benchimol & Sá, 2006b: 907-911].
- Luz, S. L. B., A. J. Shelley & M. Maia-Herzog, 1996. The need for an integrated approach to the taxonomy of neotropical Simuliidae of the onchocerciasis focus of Brazil. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 91 (6): 721-722.
- Maia-Herzog, M., A. J. Shelley, J. E. Bradley, A. P. A. L. Dias, R. H. Calvão, C. Lowry, M. Camargo, J. M. Rubio, R. J. Post & G. E. Coelho, 1999. Discovery of a new focus of human onchocerciasis in Central Brazil. *Trans. r. Soc. trop. Med. Hyg.* 93 (3): 235-239.
- Marchon-Silva, V., J. C. Caér, R. J. Post, M. Maia-Herzog & O. Fernandes, 2004. Detection of *Onchocerca volvulus* (Nematoda: Onchocercidae) infection in vectors from Amazonian Brazil following mass Mectizan™ distribution. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 99: 1-6.
- Mardini, L. B. L. F., org., 2006. *Simulídeos. Programa Estadual Rio Grande do Sul – Brasil. Guia para orientação aos municípios sobre manejo integrado, controle e gestão de insetos da família Simuliidae (Diptera, Nematocera) no Rio Grande do Sul*. Centro Estadual de Vigilância em Saúde, Secretaria Estadual da Saúde do Rio Grande do Sul, Porto Alegre.
- Mardini, L. B. L. F., M. A. T. Souza, L. Rabinovitch, R. S. A. Alves & C. M. B. Silva, 1999. Field studies with the bacterial larvicide INPALBAC for *Simulium spp.* control in Rio Grande do Sul, Brazil. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 94 (5): 679-681.
- Marino, G. S. Coscarón, J. Maurand, C. Loubés & P. Cabeza Meckert, 1980. Estudios sobre microsporidios de la región neotropical. I. Sobre la presencia de *Pleistophora simulii* (Lutz & Splendore) en la región austral de América (Microspora). *Neotropica*, La Plata 25 (74): 127-132. ("1979").
- Marino, H. A., 1993. Estudios y control de simúlidos 'paquetas' en el valle de Uspallarta, departamento de Las Heras, provincia de Mendoza. *Pest Report* 2 (7): 12-14.
- Marino, H. A., 2003. Los simúlidos (Diptera – Simuliidae) en la cuenca del río Salado, Pcia. de Buenos Aires, Argentina: Ecología y control. *Entomología y Vectores*, Rio de Janeiro 10 (4): 613-620.
- McCreadie, J. W. & C. E. Beard, 2003. The microdistribution of the trichomycete *Smittium culisetae* in the hindgut of the black fly host *Simulium vittatum*. *Mycologia* 95: 998-1003.
- Mead, D. G., E. W. Gray, R. Noblet, M. D. Murphy, E. W. Howerth & D. E. Stallknecht, 2004. Biological transmission of vesicular stomatitis virus (New Jersey serotype) by *Simulium vittatum* (Diptera: Simuliidae) to domestic swine (*Sus scrofa*). *J. med. Ent.* 41 (1): 78-82.
- Medeiros, J. F. de & V. Py-Daniel, 1999. Atividade hematofágica e infecção natural de três espécies de Simuliidae (Diptera: Culicomorpha) em Xitei/Xidea, área indígena Yanomami, Roraima, Brasil. *Entomología y Vectores*, Rio de Janeiro 6 (3): 210-226.
- Medeiros, J. F. de & V. Py-Daniel, 2002. Estado fisiológico e preferência de picadas de *Cerqueirellum argentiscutum* (Shelley & Luna Dias) (Diptera: Simuliidae) vetor de *Mansonella ozzardi* (Manson) (Nematoda: Onchocercidae) no Brasil. *Entomología y Vectores* Rio de Janeiro 9 (4): 505-517.

- Medeiros, J. F. de & V. Py-Daniel, 2003. Atividade hematofágica diária e taxa de infecção parasitária de *Cerqueirellum argentiscutum* (Shelley & Luna Dias) (Diptera: Simuliidae) por *Mansonella ozzardi* (Manson) (Nematoda: Onchocercidae) em uma localidade do baixo rio Solimões, Amazonas, Brasil. *Entomología y Vectores*, Rio de Janeiro 10 (1): 9-20.
- Medeiros, J. F. de & V. Py-Daniel, 2004. Seasonality, parity rates and transmission indices of *Mansonella ozzardi* (Manson) (Nematoda: Onchocercidae) by *Cerqueirellum argentiscutum* (Shelley & Luna Dias) (Diptera: Simuliidae) in a lower Solimões river community, Amazonas, Brazil. *Acta amazonica*, Manaus 34 (2): 221-227.
- Moraes, A. A. P., A. J. Shelley & A. P. A. L. Dias, 1985. *Mansonella ozzardi* no Território Federal de Roraima, Brasil. Distribuição e achado de um novo vetor na área do rio Surumu. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 80 (4): 395-400.
- Moraes, M. A. P., A. J. Shelley & A. P. A. L. Dias, 1986. O foco brasileiro de oncocercose: Novas observações feitas nas áreas dos rios Mucajaí e Catrimâni, Território de Roraima. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 81 (1): 105-109.
- Morales-Hojas, R., R. J. Post, A. J. Shelley, M. Maia-Herzog & S. Coscarón, 2001. Characterisation of nuclear ribosomal DNA sequences from *Onchocerca volvulus* and *Mansonella ozzardi* (Nematoda: Filarioidea) and development of a PCR-based method for their detection in skin biopsies. *Intern. J. Parasitol.* 31 (2): 169-77. [Erratum in *Intern. J. Parasitol.* 31 (8): 850-851, 2001].
- Muirhead-Thomson, R. C., 1969. A laboratory technique for establishing *Simulium* larvae in an experimental channel. *Bull. ent. Res.* 59 (3): 533-536.
- Nathan, M. B., E. S. Tikasingh & P. Munroe, 1982. Filariasis in Amerindians of Western Guyana with observations of *Mandonella ozzardi* by a *Simulium* species of the *amazonicum* group. *Trop. Med. Parasitol.* 33: 219-222.
- Nettel, F. R., 1952. Revisión del problema entomológico de la oncocercosis y pián para la erradicación de *Simulium ochraceum* Walker. *Medicina*, México 32 (660): 438-441.
- Ochoa, J. O., J. C. Castro, V. M. Barrios, E. L. Juárez & I. Tada, 1997. Successful control of onchocerciasis vectors in San Vicente Pacaya, Guatemala, 1984-1989. *Ann. trop. Med. Parasitol.* 91 (5): 471-479.
- Omar, M. S. & R. Garms, 1975. The fate and migration of microfilariae of a Guatemalan strain of *Onchocerca volvulus* in *Simulium ochraceum* and *S. metallicum*, and the role of the buccopharyngeal armature in the destruction of microfilariae. *TropenMed. Parasitol.* 26: 183-190.
- Omar, M. S. & R. Garms, 1977. Lethal damage to *Simulium metallicum* following high intakes of *Onchocerca volvulus* microfilariae in Guatemala. *TropenMed. Parasitol.* 28 (1): 109-119.
- Overmyer, J. P., K. L. Armbrust & R. Noblet, 2003. Susceptibility of black fly larvae ((Diptera: Simuliidae) to lawn-care insecticides individually and as mixtures. *Environm. Toxicol. Chem.* 22 (7): 1582-1588.
- Porter, C. H. & R. C. Collins, 1984. Descriptive statistics for the larval stages of *Onchocerca volvulus* in host-seeking *Simulium ochraceum*. *Amer. J. trop. Med. Hyg.* 33 (2): 252-260.
- Porter, C. H. & R. C. Collins, 1985. The gonotrophic cycle of wild *Simulium ochraceum* and the associated development of *Onchocerca volvulus*. *Amer. J. trop. Med. Hyg.* 34 (2): 302-309.
- Porter, C. H. & R. C. Collins, 1988a. Seasonality of adult black flies and *Onchocerca volvulus* transmission in Guatemala. *Amer. J. trop. Med. Hyg.* 38 (1): 153-167.
- Porter, C. H. & R. C. Collins, 1988b. Transmission of *Onchocerca volvulus* by secondary vectors in Guatemala. *Amer. J. trop. Med. Hyg.* 39 (6): 559-566.
- Porter, C. H., R. C. Collins & A. D. Brandling-Bennett, 1988. Vector density, parasite prevalence, and transmission of *Onchocerca volvulus* in Guatemala. *Amer. J. trop. Med. Hyg.* 39 (6): 567-574.
- Post, R. J., Z. Adams, A. J. Shelley, M. Maia-Herzog, A. P. A. L. Dias & S. Coscarón, 2003. The morphological discrimination of microfilariae of *Onchocerca volvulus* from *Mansonella ozzardi*. *Parasitology* 127: 21-27.
- Procunier, W. S., A. J. Shelley & M. Arzube, 1985. Sibling species of *Simulium exiguum* (Diptera, Simuliidae), the primary vector of onchocerciasis in Ecuador. *Revta ecuat. Hig. Medna trop.* 35 (2): 19-59.
- Procunier, W. S., A. J. Shelley & M. Arzube, 1986. A new method for age grading in Simuliidae (Diptera). *Trans. r. Soc. trop. Med. Hyg.* 80: 841-845.
- Py-Daniel, L. H. R. & V. Py-Daniel, 1984. Observações sobre *Spatularicaria evansi* (Boulenger, 1892) (Osteichthyes; Loricariidae) e a sua predação em Simuliidae (Diptera: Culicomorpha). *Bolm Mus. paraense Emílio Goeldi, Zool.*, Belém 1 (2): 207-218.
- Py-Daniel, V., 1989. Oncocercose no Solimões. *Revta Saúde públ.*, São Paulo 23 (3): 260.
- Py-Daniel, V., 1994a. Oncocercose em expansão no Brasil. *Revta Saúde públ.*, São Paulo 28 (2): 173-174.
- Py-Daniel, V., 1994b. Algumas considerações sobre o programa de erradicação da oncocercose para as Américas. *Mems Centro amazón. Invest. Control Enferm. Trop.*, Puerto Ayacucho (Amazonas, Venezuela) 4 (1-2): 157-161.
- Py-Daniel, V., R. Andreazze & J. F. de Medeiros, 2000. Projeto piloto Xitei/Xidea (Roraima). I. Índices epidemiológicos da transmissão de *Onchocerca volvulus* para os anos de 1995-1996. *Entomología y Vectores*, Rio de Janeiro 7 (4): 389-444.
- Py-Daniel, V., R. Andreazze & J. F. de Medeiros, 2002. Influência dos fatores climáticos na atividade hematofágica de *Psaroniocompsa incrustata* (Lutz, 1910) (Diptera: Simuliidae) vetor de *Onchocerca volvulus* (Leuckart, 1893) em Xitei/Xidea, área indígena Yanomami, Roraima, Brasil. *Entomología y Vectores*, Rio de Janeiro 9 (4): 559-577.

- Py-Daniel, V. & S. M. Darwich, 1997. Simuliid "Borrachudo/Pium" control in Brazil. *Brit. Simuliid Group Bull.*, Liverpool 9: 7-14.
- Py-Daniel, V. & M. Jegu, 1998. Peixes (Serrasalminae) predadores dos imaturos de *Thrysopelma guianense* (Wise, 1911) (Diptera, Culicomorpha, Simuliidae), vetor da filária *Onchocerca volvulus* (Leuckart, 1893). *Entomología y Vectores*, Salta 3 (1): 27-29.
- Py-Daniel, V., M. C. V. Passos, F. J. de Medeiros & R. Andreatze, 1999. Dinâmica da atividade hematofágica (preferências horárias-tópicas) e estado reprodutivo das fêmeas de *Thrysopelma guianense* (Wise, 1911) (Diptera, Simuliidae, Culicomorpha) principal vetor da filária *Onchocerca volvulus* (Leuckart, 1893) no Brasil. *Entomología y Vectores*, Rio de Janeiro 6 (4): 339-360.
- Py-Daniel, V. & L. H. R. Py-Daniel, 1998. Dinâmica de dispersão, padrões de distribuição geográfica e aspectos relativos à transmissão de *Onchocerca volvulus* (Leuckart, 1893) por simulídeos na bacia hidrográfica do rio Amazonas. *Entomología y Vectores*, Rio de Janeiro 5 (5): 191-215.
- Ramírez Peres, J., 1983. Los jejenes de Venezuela. *Simposio de Oncocercosis americana, Centro amazón. Invest. Control Enferm. Trop.*, Puerto Ayacucho (Amazonas, Venezuela), 156 pp. [mimeographed].
- Ramírez Pérez, J., 1984. Vectores de la oncocercosis humana en Venezuela. *Boln Direcc. Malariol. Saneam. amb.* 24: 79-94.
- Ramírez Pérez, J., 1985. Vectores de la oncocercosis humana en la región neotropical. *Boln Ofic. Sanit. panamer.* 98 (2): 117-135.
- Ramírez Pérez, J., E. Rassi, J. Convit & A. Ramírez, 1976. Importancia epidemiológica de los grupos de edad en las poblaciones de *Simulium metallicum* (Diptera: Simuliidae) de Venezuela. *Boln Ofic. sanit. panamer.* 80 (2): 105-122.
- Ramírez-Ramírez, A., G. Sánchez-Tejeda, J. Méndez-Galván, T. R. Unnasch & A. Monroy-Ostria, 2006. Molecular studies of *Onchocerca volvulus* isolates from Mexico. *Infect. Genetics Evolution* 6 (3): 171-176.
- Rassi, E., N. Lacerda & J. A. Guimarães, 1976. Estudio de una zona de oncocercosis en Brasil: Encuesta realizada en residentes locales. *Boln Ofic. sanit. panamer.* 80 (4): 288-301.
- Rassi, E., N. Lacerda, J. A. Guimarães, M. A. Vulcano, J. Ramírez Pérez & A. Ramírez, 1975. Preliminary report on a new vector of onchocerciasis in the Americas: *Simulium amazonicum* (Goeldi, Lutz, 1910 and 1917 [sic]). *Bull. Pan-Amer. Health Org.* 9 (3): 10-12., Mexico. *J. Amer. Mosquito Control Assoc.* 10 ((3): 430-433.
- Ríos-Velásquez, C. & N. Hamada, 2002. Trichomycete fungi (Zygomycota) associated with the digestive tract of *Simulium goeldii* Cerqueira & Nunes de Mello and *Simulium ulyssesi* (Py-Daniel & Coscarón) (Diptera: Simuliidae) larvae, in Central Amazonia, Brazil. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 97 (3): 423-426.
- Rodríguez Pérez, M. A., R. Danis-Lozano, M. H. Rodríguez & J. E. Bradley, 1999. Comparison of serological and parasitological assessments of *Onchocerca volvulus* transmission after 7 years of mass ivermectin treatment in Mexico. *Trop. Med. int. Health* 4 (2): 98-104.
- Rodríguez Pérez, M. A., R. Danis-Lozano, M. H. Rodríguez, T. R. Unnasch & J. E. Bradley, 1999. Detection of *Onchocerca volvulus* infection in *Simulium ochraceum* sensu lat: Comparison of a PCR assay and fly dissection in a Mexican hypoendemic community. *Parasitology* 119 (6): 613-619.
- Rodríguez Pérez, M. A., C. R. Katholi, H. K. Hassan & T. R. Unnasch, 2006. Large-scale entomologic assessment of *Onchocerca volvulus* transmission by poolscreen PCR in Mexico. *Amer. J. trop. Med. Hyg.* 74 (6): 1026-1033.
- Rodríguez Pérez, M. A., B. G. Lilley, A. Domínguez-Vásquez, R. Segura-Arenas, C. Lizarazo-Ortega, A. Mendoza-Herrera, F. Reyes-Villanueva & T. R. Unnasch, 2004. Polymerase chain reaction monitoring of transmission of *Onchocerca volvulus* in two endemic states in Mexico. *Amer. J. trop. Med. Hyg.* 70 (1): 38-45.
- Rodríguez Pérez, M. A., C. A. Núñez González, C. Lizarazo Ortega, A. Sánchez Varela, M. C. Wootren & T. R. Unnasch, 2006. Analysis of genetic variation in ribosomal DNA internal transcribed spacer and the NADH dehydrogenase subunit 4 mitochondrial genes of the onchocerciasis vector *Simulium ochraceum*. *J. med. Ent.* 43 (4): 701-706.
- Rodríguez Pérez, M. A. & F. Reyes Villanueva, 1994. Efecto de la ivermectina sobre la transmisión de *Onchocerca volvulus* en el sur de México. *Salud públ. de México* 36 (3): 281-290.
- Rodríguez Pérez, M. A., F. Reyes Villanueva, H. A. Barrera Saldaña, A. Domínguez-Vázquez & C. Lizararo Ortega, 2007. Impacto del programa de tratamiento con ivermectina sobre la supresión e interrupción de la transmisión de *Onchocerca volvulus* en México. *Ciencia UANL* [Universidad Autónoma de Nuevo León] 8 (4): 493-489.
- Rodríguez Pérez, M. A., F. Reyes-Villanueva & M. H. Rodríguez, 1995. Estimating the gonotrophic cycle and survivorship of *Simulium ochraceum* (Diptera: Simuliidae) during routine vector surveillance in southern Mexico. *J. Amer. Mosquito Control Assoc.* 11 (3): 360-362.
- Rodríguez Pérez, M. A. & A. R. Rivas Algaria, 1991. Algunos problemas en la investigación para el control de la transmisión de *Onchocerca volvulus* en México. *Salud públ. de México* 33 (5): 493-503.
- Rodríguez Pérez, M. A., M. H. Rodriguez, H. M. Margeli-Pérez & A. R. Rivas-Alcalá, 1995. Effect of semiannual treatment of ivermectin on the prevalence and intensity of *Onchocerca volvulus* skin infection, ocular lesions, and infectivity of *Simulium ochraceum* populations in southern Mexico. *Amer. J. trop. Med. Hyg.* 52 (5): 429-434.
- Rodríguez Pérez, M. A., N. L. Valdivieso López & P. J. McCall, 2003. Aggregated oviposition in *Simulium ochraceum* s. l. (Diptera: Simuliidae), an important

- neotropical vector of *Onchocerca volvulus*. *Ann. trop. Med. Parasitol.* 97 (2): 203-207.
- Romaña, C. & J. W. Ábalos, 1948. Control larvario de simúlidos y culicídos con Gammexane y DDT. *An. Inst. Medna trop.*, Tucumán 2 (2): 107-146.
- Rowe, S. G. & M. Durand, 1998. Blackflies and whitewater: Onchocerciasis and the eye. *Intern. Ophthalmol. Clin.* 38 (1): 231-240.
- Ruas Neto, A. L., 1984a. Avaliação do uso de temephos para o controle de culicídeos e simulídeos no Rio Grande do Sul. *Bolm Saúde*, Porto Alegre 11: 27-31.
- Ruas Neto, A. L., 1984b. *Bacillus thuringiensis* var. *israelensis* como alternativa no controle de simulídeos no Rio Grande do Sul. 1. Susceptibilidade a campo. *Bolm Saúde*, Porto Alegre 11: 12-16.
- Ruas Neto, A. L. & S. M. Silveira, 1989. Uso de inseticidas bacterianos para o controle de culicídeos e simulídeos [sic] no Rio Grande do Sul. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 84: 39-45.
- Ruas Neto, A. L., M. A. T. Souza, S. Severino, J. L. B. Melo, S. M. Silveira & N. D. F. Fontes, 1985. Controle integrado do *Simulium (Chrirostilbia) pertinax* Kollar, 1822 – Utilização de *Bacillus thuringiensis* vcar. *israelensis* em três municípios do Rio Grande do Sul. *Bolm Saúde*, Porto Alegre 12: 17-20.
- Sanmartín, C., R. B. Mackenzie, H. Trapido, P. Barreto, C. H. Mullenax, E. Gutiérrez, & C. Lesmes, 1973. Encefalitis equina venezolana en Colombia, 1967. *Boln Ofic. sanit. panamer.* 74 (2): 108-137. [Abstract in *Rev. applied Ent.* (B) 61 (10): 564-565].
- Sato, G., 1987. Identificação de peixes predadores de larvas de simulídeos da região de Joinville/SC. *Ciência e Cultura*, São Paulo 39 (10): 962-966.
- Schiller, E. L., J. L. Petersen, D. Shirazian & H. Figueroa Marroquin, 1984. Morphogenesis of larval *Onchocerca volvulus* in the Panamanian black fly, *Simulium quadrivittatum*. *Amer. J. trop. Med. Hyg.* 33 (3): 419-413.
- Shelley, A. J., 1975. A preliminary survey of the prevalence of *Mansonella ozzardi* in some rural communities on the river Purus, state of Amazonas, Brazil. *Ann. trop. Med. Parasitol.* 69 (3): 407-412.
- Shelley, A. J., 1983. The identities of the Brazilian blackflies (Diptera: Simuliidae) reported by Lutz and Splendore as hosts of Microsporidia. *J. invert. Pathol.* 42: 280-281.
- Shelley, A. J., 1988a. Biosystematic and medical importance of the *Simulium amazonicum* group and *S. exiguum* complex in Latin America, pp. 203-220, in Service, M. W., ed., *Biosystematics of haematophagous insects*, 363 pp. [Systematics Association, Special volume 37], Oxford University Press, Oxford.
- Shelley, A. J., 1988b. Vector aspects of the epidemiology of onchocerciasis in Latin America. *Ann. Rev. Ent.* 30: 337-366.
- Shelley, A. J., 1988c. Biosystematics and distribution of simuliid vectors of human onchocerciasis in South America. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 83 (4): 399-403.
- Shelley, A. J., 1991. Simuliidae and the transmission and control of human onchocerciasis in Latin America. *Cadernos de Saúde públ.* 7 (3): 310-327.
- Shelley, A. J., 2000. The Simuliidae of the secondary onchocerciasis focus at Minaçu in Central Brazil. *Bull. nat. Hist. Mus.*, London 69 (2): 171-221.
- Shelley, A. J., 2001. Simuliidae and the transmission and control of human onchocerciasis in Latin America. *Cadernos Saúde públ.*, Rio de Janeiro 7 (3): 319-327.
- Shelley, A. J., 2002. Human onchocerciasis in Brazil: An overview. *Cadernos Saúde públ.*, Rio de Janeiro 18 (5): 1167-1177.
- Shelley, A. J. & M. Arzube, 1985. Studies on the biology of Simuliidae (Diptera) at the Santiago onchocerciasis focus in Ecuador, with special reference to the vectors and disease transmission. *Trans. r. Soc. trop. Med. Hyg.* 79: 328-338.
- Shelley, A. J., M. Arzube & M. Couch, 1989. Simuliidae (Diptera) of Santiago onchocerciasis focus of Ecuador. *Bull. Brit. Mus. nat. Hist., Ent.* 58: 79-130.
- Shelley, A. J., M. Charalambous & M. Arzube, 1990. *Onchocerca volvulus* development in four *Simulium exiguum* cytospecies in Ecuador. *Bull. Soc. française Parasitol.* 8: 1145.
- Shelley, A. J., A. P. A. L. Dias, M. Maia-Herzog, M. Camargo, E. G. Costa, P. Garritano & C. A. Lowry, 2001. Biting behaviour and potential vector status of anthropophilic simuliid species (Diptera: Simuliidae) in a new focus of human onchocerciasis at Minaçu, Central Brazil. *Med. vet. Ent.* 15 (1): 28-39.
- Shelley, A. J., A. P. A. L. Dias, M. Maia-Herzog, W. S. Procunier & M. A. P. Moraes, 1987. Identification of vector species (Diptera: Simuliidae) of human onchocerciasis in the Amazonia focus of Brazil and Venezuela. *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 82 (4): 461-465.
- Shelley, A. J., A. P. A. L. Dias & M. A. P. Moraes, 1980. *Simulium* species of the *amazonicum*-group as vectors of *Mansonella ozzardi* in the Brazilian Amazon. *Trans. r. Soc. trop. Med. Hyg.* 74: 784-788.
- Shelley, A. J., A. P. A. L. Dias, M. A. P. Moraes & W. S. Procunier, 1987. The status of *Simulium oyapockense* and *S. limbatum* as vectors of human onchocerciasis in Brazilian Amazonia. *Med. veter. Ent.* 1: 219-234.
- Shelley, A. J., A. P. A. L. Dias, M. A. P. Moraes, W. S. Procunier & C. A. Couch, 1988. Simuliidae and human onchocerciasis in the Amazonia focus, pp. 190-193, in Olejniewk, J., ed., *Medical and veterinary dipterology: Proceedings of the international conference, November 14-December 4, 1987*, Èeské Budìjovice. House of Technics ÈSVTS, Èeské Budìjovice.
- Shelley, A. J., L. M. Hernández, M. Maia-Herzog & A. P. A. L. Dias, 2002. Morphological variation in the larva of *Simulium guianense* s. l. Wise (Dipt., Simuliidae), a primary vector of human onchocerciasis in Brazil. *Entomologist's monthly Mag.* 138: 95-101.
- Shelley, A. J., C. A. Lowry, M. Maia-Herzog, A. P. A. L. Dias & M. A. P. Moraes, 1997. Biosystematic studies on the Simuliidae

- (Diptera) of the Amazonian onchocerciasis focus of Brazil. *Bull. nat. Hist. Mus., Ent.*, London 66 (1): 1-121.
- Shelley, J. A., M. Maia-Herzog, A. P. A. L. Dias, M. Camargo, E. G. Costa, P. Garritano & C. A. Lowry, 2001. Biting behaviour and potential vector status of anthropophilic blackflies in a new focus of human onchocerciasis at Minaçu, central Brazil. *Med. veter. Ent.* 18: 28-39.
- Shelley, A. J., M. Maia-Herzog, A. P. A. L. Dias, M. Camargo & P. Garritano, 2001. Biting behaviour and potential vector status of anthropophilic simuliid species (Diptera: Simuliidae) in a new focus of human onchocerciasis at Minaçu, Central Brazil. *Med. vet. Ent.* 15: 1-12.
- Shelley, A. J., M. Maia-Herzog, A. P. A. L. Dias & S. Coscarón, 2002. Simuliids as vectors of *Onchocerca volvulus* and *Mansonella ozzardi* in Latin America: Significance on onchocerciasis epidemiology and control. *Acta tropica* 83: 1-49.
- Shelley, A. J., M. Maia-Herzog, C. A. Lowry, A. P. A. L. Dias, P. Garritano, M. Camargo & H. G. Carter, 2000. The Simuliidae (Diptera) of the secondary onchocerciasis focus at Minaçu in Central Brazil. *Bull. nat. Hist. Mus., Ent.*, London 69 (2): 171-221.
- Shelley, A. J., M. Maia-Herzog, C. A. Lowry, A. P. L. Dias, P. R. Garritano, A. Shelley, M. Camargo & H. G. Carter, 2000. The Simuliidae (Diptera) of the secondary onchocerciasis focus at Minaçu in Central Brazil. *Bull. Mus. nat. Hist.*, London 69 (2): 171-221.
- Shelley, A. J., J. A. S. N. de Mello & R. G. O. Rees, 1976. Observações preliminares sobre a transmissão de oncocercose no rio Tootobi, Amazonas, Brasil. *Acta amazonica*, Manaus 6 (3): 327-334.
- Shelley, A. J., R. R. Pinger, M. A. P. Moraes, J. D. Charlwood & J. Hayes, 1979. Vectors of *Onchocerca volvulus* at the river Tootobi, Brazil. *J. Hemithol.* 53: 41-43.
- Shelley, A. J., R. R. Pinger, M. A. P. Moraes & J. Hayes, 1979. Concentration of microfilariae of *Onchocerca volvulus* by *Simulium sanguineum* during feeding; use in mapping parasite distribution in skin. *J. med. Ent.* 16: 48-51.
- Shelley, A. J., W. S. Proculier & M. Arzube, 1986. Direct incrimination of *Simulium exiguum* Cayapa form as a vector of *Onchocerca volvulus* in Ecuador. *Trans. r. Soc. trop. Med. Hyg.* 80: 845.
- Shelley, A. J. & A. Shelley, 1976. Further evidence for the transmission of *Mansonella ozzardi* by *Simulium amazonicum* in Brazil. *Ann. trop. Med. Parasitol.* 70 (2): 213-217.
- Smith, C. N., 1973. Pyrethrum for control of insects affecting man and animals, in Casida, J. E., ed., *Pyrethrum, the natural insecticide*, 329 pp. Academic Press, London.
- Souza, M. A., 1984. Atendimento médico por picadas de simulídeos [sic]. *Bolm Saúde*, Porto Alegre 11: 8-11.
- Stallings, T., M. S. Cupp & E. W. Cupp, 2002. Orientation of *Onchocerca lienalis* Stiles (Filarioidea: Onchocercidae) microfilariae to black fly saliva. *J. med. Ent.* 39 (6): 908-914.
- Strieder, M. N., 1986. Ocorrência de Simuliidae (Diptera: Nematocera) no conteúdo estomacal de peixes do arroio Feitoria, Picada Verão, Sapiranga, Rio Grande do Sul – Brasil. *Acta biol. leopold.* 8 (1): 167-176.
- Tada, I., 1987. A comparative study on onchocerciasis between South and Central America. Shinoda Printing Co., Ltd., Matsubase, Shimomashiki-gun, Kumamoto.
- Takaoka, H., 1980. Pathogens of blackfly larvae in Guatemala and their influence on natural populations of three species of onchocerciasis vectors. *Amer. J. trop. Med. Hyg.* 29 (3): 467-472.
- Takaoka, H., 1981. Seasonal occurrence of *Simulium ochraceum*, the principal vector of *Onchocerca volvulus* in the southeastern endemic area of Guatemala. *Amer. J. trop. Med. Hyg.* 30 (5): 1121-1132.
- Takaoka, H., 1982. Observation on the bionomics of larval and man-biting female populations of *Simulium horacioi*, a new potential vector of *Onchocerca volvulus* in Guatemala. *Japan. J. trop. Med. Hyg.* 10: 49-62.
- Takaoka, H., J. O. Ochoa, E. L. Juárez & K. M. Hansen, 1982. Effects of temperature on development of *Onchocerca volvulus* in *Simulium ochraceum*, and longevity of the simuliid vector. *J. Parasitol.* 68 (3): 478-483.
- Takaoka, H., H. Suzuki, S. Noda, J. O. Ochoa A. & I. Tada, 1984. The intake, migration and development of *Onchocerca volvulus* microfilariae in *Simulium haematopotum* in Guatemala. *Japan. J. sanit. Zool.* 35 (2): 121-127.
- Takaoka, H., H. Suzuki, S. Noda, I. Tada, M. G. Basáñez & L. Yarzábal, 1984. Development of *Onchocerca volvulus* larvae in *Simulium pintoi* in the Amazonas region of Venezuela. *Amer. J. trop. Med. Hyg.* 33 (3): 414-419.
- Takaoka, H., I. Tada, M. Baba, M. Shimada, R. F. Lazos, J. Rumbea G, R. Farias D., R. H. Guderian & M. Amunarriz, 1988. Comparative studies on three anthropophilic blackfly species in Ecuador as the vector of human onchocerciasis. *Japan. J. Parasitol.* 37 (2): 76-83.
- Takaoka, H., I. Tada, Y. Hashiguchi, M. Baba, M. Korenaga, J. O. Ochoa A. & J. Convit, 1986. Experimental infections of three Guatemalan blackfly species with north Venezuelan *Onchocerca volvulus*. *Japan. J. sanit. Zool.* 4: 319-323.
- Tanaka, I., Y. Hashiguchi, T. Okazawa, J. O. Ochoa A. & I. Tada, 1980. Duration of blood feeding of *Simulium ochraceum* in relation to intake of *Onchocerca volvulus* microfilariae. *Japan. J. sanit. Zool.* 31 (3): 209-214.
- Tidwell, M. A., P. Muñoz de Hoyos & A. Corredor, 1980. *Simulium exiguum*, the vector of *Onchocerca volvulus* on the rio Micay, Colombia. *Amer. J. trop. Med. Hyg.* 29: 377-381.
- Tidwell, M. A., B. V. Peterson, J. Ramírez Pérez, M. A. Tidwell & L. A. Lacey, 1980. Notas y claves preliminares de los jejenes neotropicales pertenecientes a los grupos *Simulium amazonicum* y *S. sanguineum* (Diptera: Simuliidae) incluyendo los vectores de *Onchocerca volvulus* y *Mansonella ozzardi*. *Boln Div. Malariaol. Saneam. amb.* 21 (2): 79-89.
- Tidwell, M. A. & M. A. Tidwell, 1982. Development of *Mansonella ozzardi* in *Simulium amazonicum*, *S. argentiscutum* and *Culicoides insinuatus* from Amazonas, Colombia. *Amer. J. trop. Med. Hyg.* 31 (6): 1137-1141.

- Tidwell, M. A., M. A. Tidwell & P. Muñoz de Hoyos, 1980. Development of *Mansonella ozzardi* in a black fly species of the *Simulium sanguineum* group from eastern Vaupés, Colombia. *Amer. J. trop. Med. Hyg.* 29 (6): 1209-1214.
- Tidwell, M. A., M. A. Tidwell, P. Muñoz de Hoyos & A. Corredor, 1980. *Simulium exiguum*, the vector of *Onchocerca volvulus* on the río Micay, Colombia. *Amer. J. trop. Med. Hyg.* 29 (3): 377-381.
- Tidwell, M. A., M. A. Tidwell, P. Muñoz de Hoyos, A. Corredor & P. Barreto, 1980. Vectores de *Onchocerca volvulus* y *Mansonella ozzardi* en Colombia. *Colombia méd.* 11 (4): 119-127.
- Torres Fernández, O., P. Muñoz de Hoyos & G. Romero de Pérez, 1991. Parasitismo en larvas de simúlidos (Diptera: Simuliidae) del río Teusaca: Microsporidios, mermítidos y hongos. *Revta Acad. colombiana Cienc.* 18 (69): 253-264.
- Travis, B. V., M. Vargas V. & F. Fallas B., 1979. Bionomics of black flies (Diptera: Simuliidae) in Costa Rica. III. Larval population dynamics in five selected streams. *Revta Biol. trop.*, San José 27 (1): 135-143.
- Travis, B. V., M. Vargas V. & J. C. Swartzwelder, 1974. Bionomics of black flies (Diptera, Simuliidae) in Costa Rica. I. Species biting man, with an epidemiological summary for the Western Hemisphere. *Revta Biol. trop.*, San José 22 (1): 187-200.
- Vargas, L., 1941. Notas sobre la importancia sanitaria de los simúlidos y de su morfología interna. *Revta Inst. Salubr. Enferm. trop.*, México 2: 213-236.
- Vargas, L., 1942. Algunas consideraciones sobre el desarrollo de *Onchocerca volvulus* en los simúlidos. *Revta Inst. Salubr. Enferm. trop.*, México 3 (1): 57-65.
- Vargas, L., 1947. Notas sobre la onchocerciasis. VI. Consideraciones sobre la biología de las larvas de simúlidos. *Gaceta méd. México* 77 (6): 346-352.
- Vargas, L., 1948a. Notas sobre oncocercosis. VII. Infección experimental de *Simulium (Lanea) mangabeirai* con *Onchocerca volvulus*. *Revta Inst. Salubr. Enferm. trop.*, México 9: 309-311.
- Vargas, L., 1948b. Notas sobre la onchocerciasis. VIII. Lineamientos entomológicos sobre el control de los simúlidos. *Revta Inst. Salubr. Enferm. trop.*, México 9 (4): 313-320.
- Vargas, L., 1948c. Los simúlidos en la transmisión de la oncocerciasis americana. *Medicina*, México 28 (555): 177-190.
- Vargas, L., 1952. Algunos aspectos de la ecología de las larvas de simúlidos en relación con la transmisión de *Onchocerca volvulus* en Chiapas. *Medicina*, México 32 (657): 353-361.
- Vargas, L. & A. Díaz Nájera, 1980. Entomologic considerations in the study of onchocerciasis transmission. *Arch. Investig. méd.*, México 11 (2): 273-279.
- Vieira, J. C., L. Brackenboro, C. H. Porter, M. G. Basáñez & R. C. Collins, 2002. Biting and infection rates of *Simulium exiguum* s. l. and *S. quadrivittatum* in two hyperendemic areas of Ecuador before the initiation of ivermectin control. *Brit. Simuliid Group Bull.* 18: 176-19.
- Vieira, J. C., L. Brackenboro, C. H. Porter, M. G. Basáñez & R. C. Collins, 2005. Spatial and temporal variation in biting rates and parasite transmission potentials of onchocerciasis vectors in Ecuador. *Trans. r. Soc. trop. Med. Hyg.* 99 (3): 178-195.
- Villalobos, L. C. de & N. B. Camino, 1997. *Limnodermis subtropicalis* n. sp. (Nematoda: Mermithidae), a parasite of larvae of *Simulium orbitale* Lutz (Diptera: Simuliidae). *Mems Inst. Oswaldo Cruz*, Rio de Janeiro 92 (3): 339-341.
- Vivas-Martínez, S., M. G. Basáñez, C. Botto, S. Rojas, M. García, M. Pacheco & C. F. Curtis, 2000. Amazonian onchocerciasis: Parasitological profiles by host-age, sex, and endemicity in southern Venezuela. *Parasitology* 121 (5): 513-525.
- Vivas-Martínez, S., M. G. Basáñez, C. Botto, L. Villegas, M. García & C. F. Curtis, 2000. Parasitological indicators of onchocerciasis relevant to ivermectin control programmes in the Amazonian focus of Southern Venezuela. *Parasitology* 121 (5): 527-534.
- Vivas-Martínez, S., M. G. Basáñez, M. E. Grillet, H. Weiss, C. Botto, M. García, N. J. Villamizar & D. C. Chavasse, 1998. Onchocerciasis in the Amazonian focus of southern Venezuela: Altitude and blackfly species composition as predictors of endemicity to select communities for ivermectin control programmes. *Trans. r. Soc. trop. Med. Hyg.* 92 (6): 613-620.
- Vojvodiæ, S., M. D. Nelder & J. W. McCreadie, 2006. Influence of fixation of the blackfly *Simulium vittatum* on morphological characters of the trichomycete *Smittium culisetae*. *Acta ent. serbica* 2006 (suppl.): 125-130.
- Wada, Y., 1982. Theoretical approach to the epidemiology of onchocerciasis in Guatemala. *Japan. J. med. Sci. Biol.* 35 (4): 183-196.
- World Health Organization (Division of Vector Biology and Control. Onchocerciasis Control Programme in the Volta River Basin), 1982. *A review of the literature concerning the feeding behaviour and general ecology of Simulium. Technical Report*, 40 pp. WHO, London.
- Wygodzinsky, P. & S. Coscarón, 1989. A revision of the blackfly genus *Gigantodax* Enderlein (Simuliidae, Diptera, Insecta). *Bull. Am. Mus. nat. Hist.*, New York 189: 1-269.
- Yamagata, Y., J. O. Ochoa, P. A. Molina, H. Sato, K. Uemoto & T. Suzuki, 1987. Chemical control of *Simulium ochraceum* Walker (Diptera: Simuliidae) larvae in an onchocerciasis endemic area of Guatemala. *Trop. Med. Parasitol.* 38 (3): 205-210.
- Yamagata, Y., T. Suzuki & G. A. García-Manzo, 1986. Geographical distribution of the prevalence of nodules of *Onchocerca volvulus* in Guatemala over the last four decades. *Trop. Med. Parasitol.* 37 (1): 28-34.
- Yarzábal, L., M. G. Basáñez, J. Ramírez-Pérez, A. Ramírez, C. Botto & A. Yarzábal, 1985. Experimental and natural infection of *Simulium sanctezi* by *Mansonella ozzardi* in the middle Orinoco region of Venezuela. *Trans. r. Soc. trop. Med. Hyg.* 79: 29-33.

Index

- Araucnephia* Wygodzinsky & Coscarón, 1973 – 5, 6
Araucnephioides Wygodzinsky & Coscarón, 1973 – 5, 6
Aspathia Wygodzinsky & Coscarón, 1973 – 8, 9, 10, 12, 13, 14, 15

bicoloratum-group, *Simulium (Ectemnaspis)* – 10, 11, 12, 14, 16
blancasi-group, *Simulium (Psilopeltmia)* – 9, 13, 14, 16
brachycladum-group, *Simulium (Hemicnetha)* – 9, 11, 12, 14, 16
brophyi-group, *Gigantodax* – 7, 8
Byssodon Enderlein, 1925 – 9, 12, 14, 16

Cerqueirellum Py-Daniel, 1983 – 10, 12, 14, 15
Chirostilbia Enderlein, 1921 – 8, 10, 11, 12, 13, 14, 15, 16
cilicinus-group, *Gigantodax* – 7, 8
Cnesia Enderlein, 1934 – 4, 5, 6
Cnesiamima Wygodzinsky & Coscarón, 1973 – 4, 5, 6
cormonsi-group, *Gigantodax* – 7, 8
Coscaroniellum Py-Daniel, 1983 – 10, 12, 14, 15

dinellii-group, *Simulium (Ectemnaspis)* – 10, 13, 15, 16

Ectemnaspis Enderlein, 1914 – 10, 11, 12, 13, 14, 15, 16
escomeli-group, *Simulium (Psilopeltmia)* – 10
Eusimulium Roubaud, 1906 – 8, 11, 14, 16

Gigantodax Enderlein, 1925 – 4, 6

Hearlea Vargas, Martínez Palacios & Díaz Nájera, 1946 – 9, 12, 13, 15
Hemicnetha Enderlein, 1934 – 9, 11, 12, 13, 14, 15

igniculus-group, *Gigantodax* – 7, 8
Inaequalium Coscarón & Wygodzinsky, 1984 – 8, 11, 13, 14, 16
incrustatum-group, *Simulium (Psaroniocompsa)* – 12

*Kempf**simulium* Py-Daniel & Mello, 1982 – 4, 5, 7

Lutzsimulium d'Andretta Jr. & d'Andretta, 1947 – 4, 7

Mayacnephia Wygodzinsky & Coscarón, 1973 – 6
mexicanum-group, *Simulium (Hemicnetha)* – 12, 14, 15
minor-group, *Gigantodax* – 7, 8
multifilis-group, *Gigantodax* – 7, 8

nemorale-group, *Simulium (Pternaspatha)* – 11
Nevermannia Enderlein, 1921 – 8, 11, 14, 16
nigristriatum-group, *Simulium (Pternaspatha)* – 11
Notolepria Enderlein, 1930 – 9, 12, 14, 16

oviedo-group, *Simulium (Hemicnetha)* – 9, 12, 14, 15

Paraustrosimulium Wygodzinsky & Coscarón, 1964 – 4, 6
paynei-group, *Simulium (Hemicnetha)* – 11, 12, 13, 15
Pedrowygomyia Coscarón & Miranda Esquivel, 1998 – 4, 6
perflavum-group, *Simulium (Ectemnaspis)* – 10, 13, 14, 15, 16
pertinax-group, *Simulium (Chirostilbia)* – 11, 16
Psaroniocompsa Enderlein, 1934 – 10, 12, 13, 14, 16
Psilopeltmia Enderlein, 1934 – 9, 10, 13, 14, 15, 16
Psilozia Enderlein, 1936 – 9, 12, 14, 15, 16
Pternaspatha Enderlein, 1930 – 9, 11, 14, 16

quadrifidum-group, *Simulium (Coscaroniellum)* – 10
quadrivittatum-group, *Simulium (Coscaroniellum)* – 10, 12

romanai-group, *Simulium (Ectemnaspis)* – 10, 11, 12, 14, 16

Simulium Latreille, 1802 – 3, 5, 6, 8
siolii-group, *Simulium (Psaroniocompsa)* – 12
subpallidum-group, *Simulium (Chirostilbia)* – 10, 11, 12, 13, 14, 16

Thrysopelma Enderlein, 1934 – 9, 11, 12, 15
Tlalocomyia Wygodzinsky & Díaz Nájera, 1970 – 4, 5, 6
Trichodagmia Enderlein, 1934 – 9, 11, 13, 14, 15

wrighti-group, *Gigantodax* – 7, 8

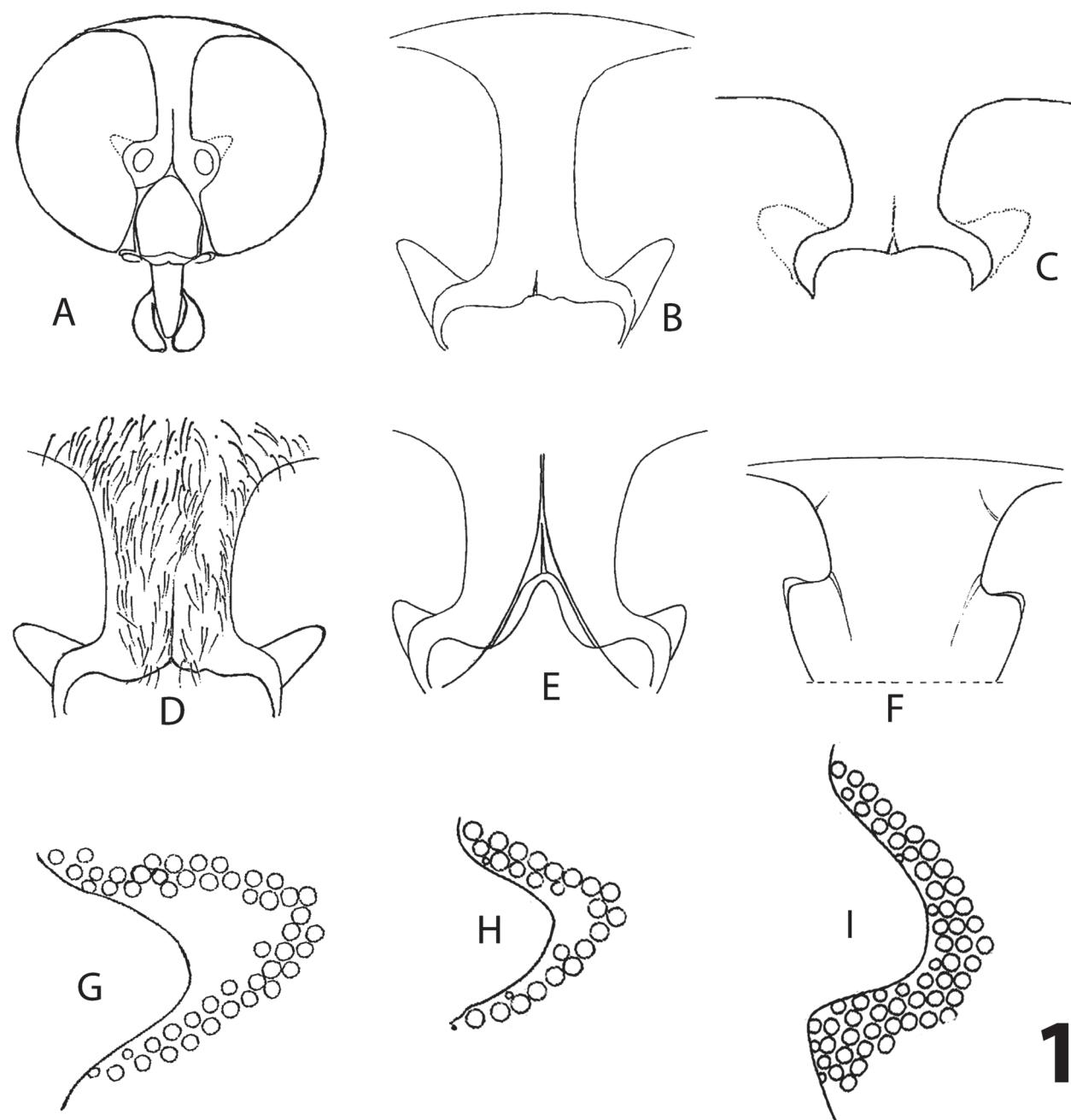


Figure 1. A. *Gigantodax brophyi* (Edwards, 1931). Head. B-F. Frons. G-I. Fronto-ocular triangle. B. *Mayacnephia aguirrei* (Dalmat, 1949). C. *Araucnephia montana* (Philippi, 1865). D. *Lutzsimulium hirticosta* (Lutz, 1909). E. *Kempfsimulium simplicicolor* (Lutz, 1910). F. *Simulium (Coscaroniellum) quadrifidum* Lutz, 1917. G. *Simulium (Pternaspatha) philippii* Coscarón, 1976. H. *Simulium (Cerqueirellum) delponteanum* Wygodzinsky, 1961. I. *Simulium (Cerqueirellum) chaquense* Coscarón, 1971.

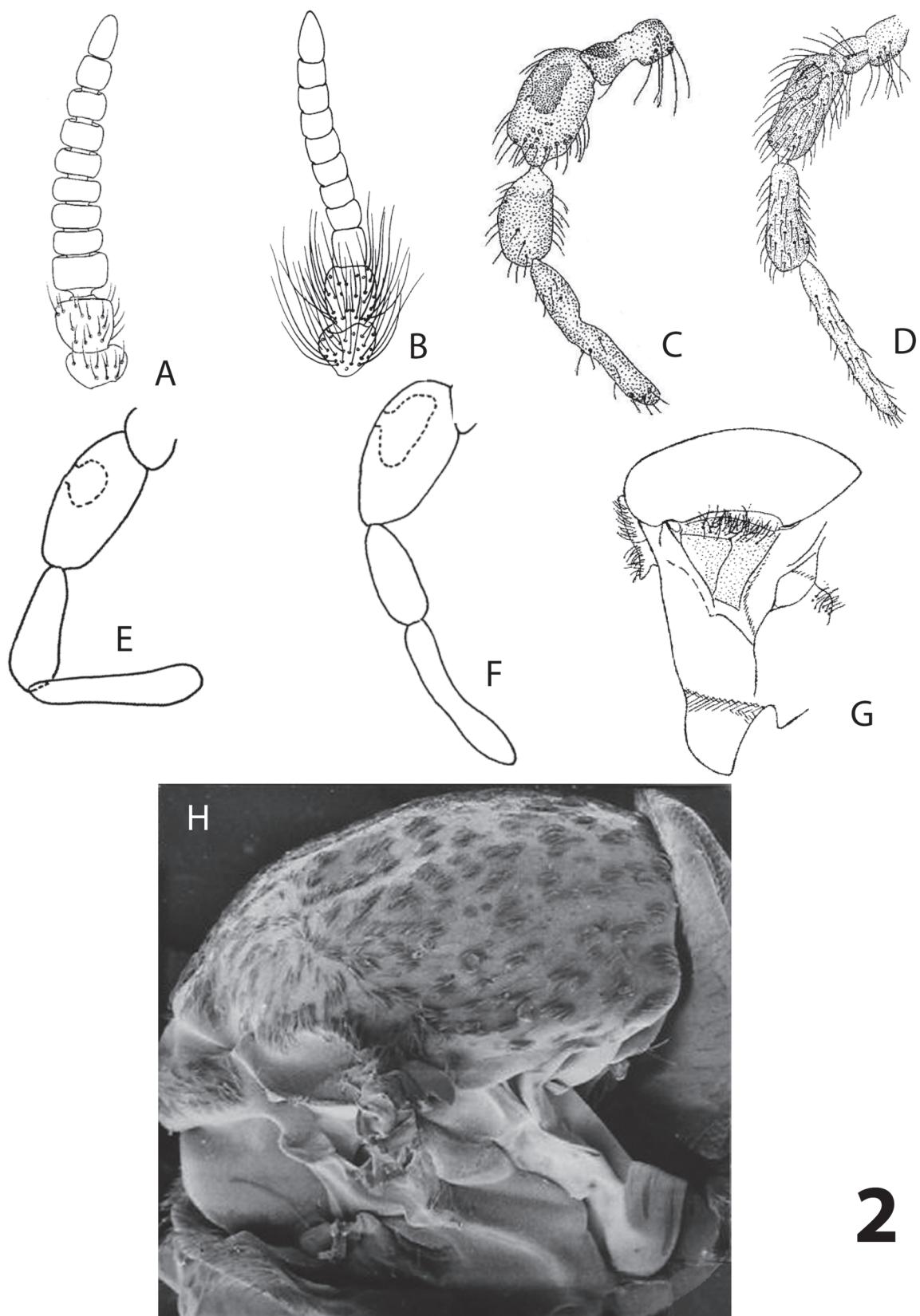


Figure 2. A-B. Antennae. C-F. Maxillary palpi. A. *Araucnephioides schlingeri* Wygodzinsky & Coscarón, 1973. B. *Pedrowygomyia cortesi* (Wygodzinsky & Coscarón, 1989). C. *Mayacnephia aguirrei* (Dalmat, 1949). D. *Araucnephia montana* (Philippi, 1865). E. *Gigantodax multifilis* Wygodzinsky & Coscarón, 1989. F. *Gigantodax antarcticus* (Bigot, 1888). G. *Tlalocomyia revelata* Wygodzinsky & Díaz Nájera, 1970: thorax, lateral view. H. *Simulium (Notolepria) exiguum* Roubaud, 1906: groups of hairs of scutum.

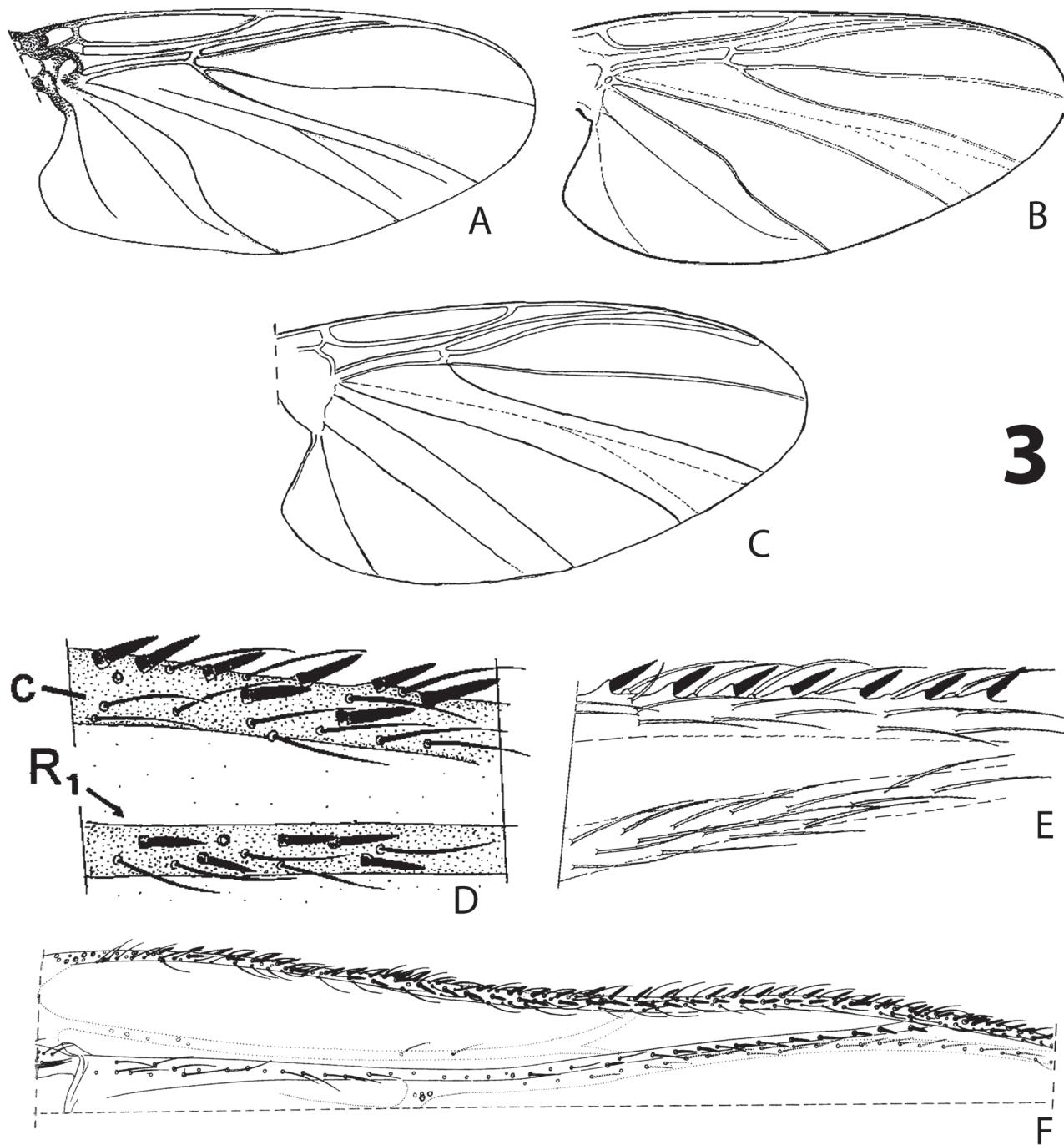


Figure 3. A-C. Wings. D-E. Portion of anterior margin of wing, showing veins C and R₁. A. *Mayacnephia aguirrei* (Dalmat, 1949). B. *Cnesia dissimilis* (Edwards, 1931). C. *Gigantodax dryadicaudicis* Wygodzinsky & Coscarón, 1989. D. *Araucnephia montana* (Philippi, 1865). E. *Cnesia dissimilis* (Edwards, 1931). F. *Simulium (Chirostilbia) subpallidum* Lutz, 1910.

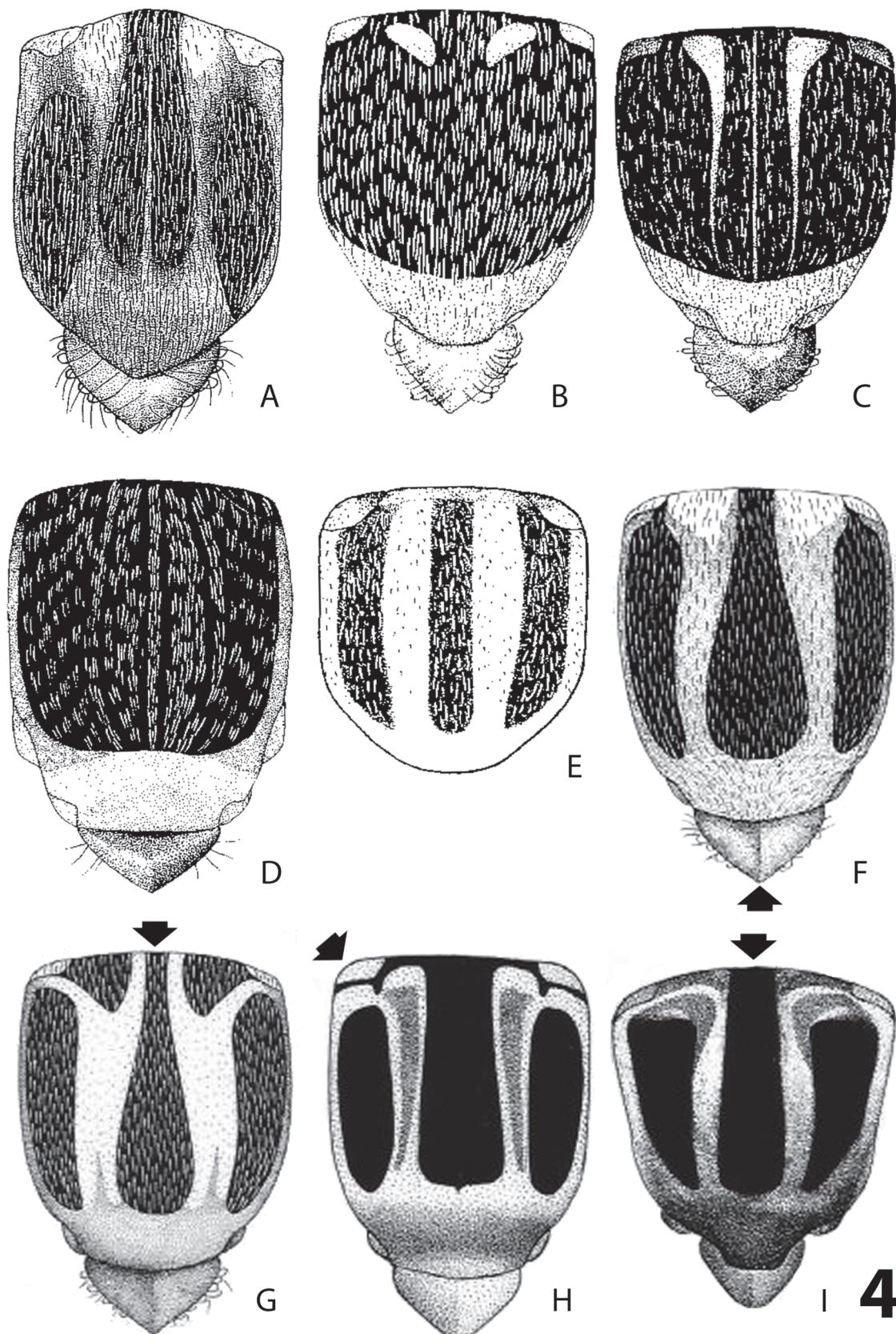


Figure 4. Scutum and scutellum [arrows indicate angle of incidence of light]. A. *Simulium (Pternaspatha) diamantinum* Coscarón & Coscarón Arias, 1996. B. *Simulium (Psaroniocompsa) incrassatum* Lutz, 1910. C. *Simulium (Psaroniocompsa) limbatum* Knab, 1915. D. *Simulium (Psaroniocompsa) auristriatum* Lutz, 1910. E. *Simulium (Psaroniocompsa) damascenoi* Py-Daniel, 1988. F-G *Simulium (Cerqueirellum) minusculum* Lutz, 1910. H-I. *Simulium (Cerqueirellum) oyapockense* Floch & Abonnenc, 1946.

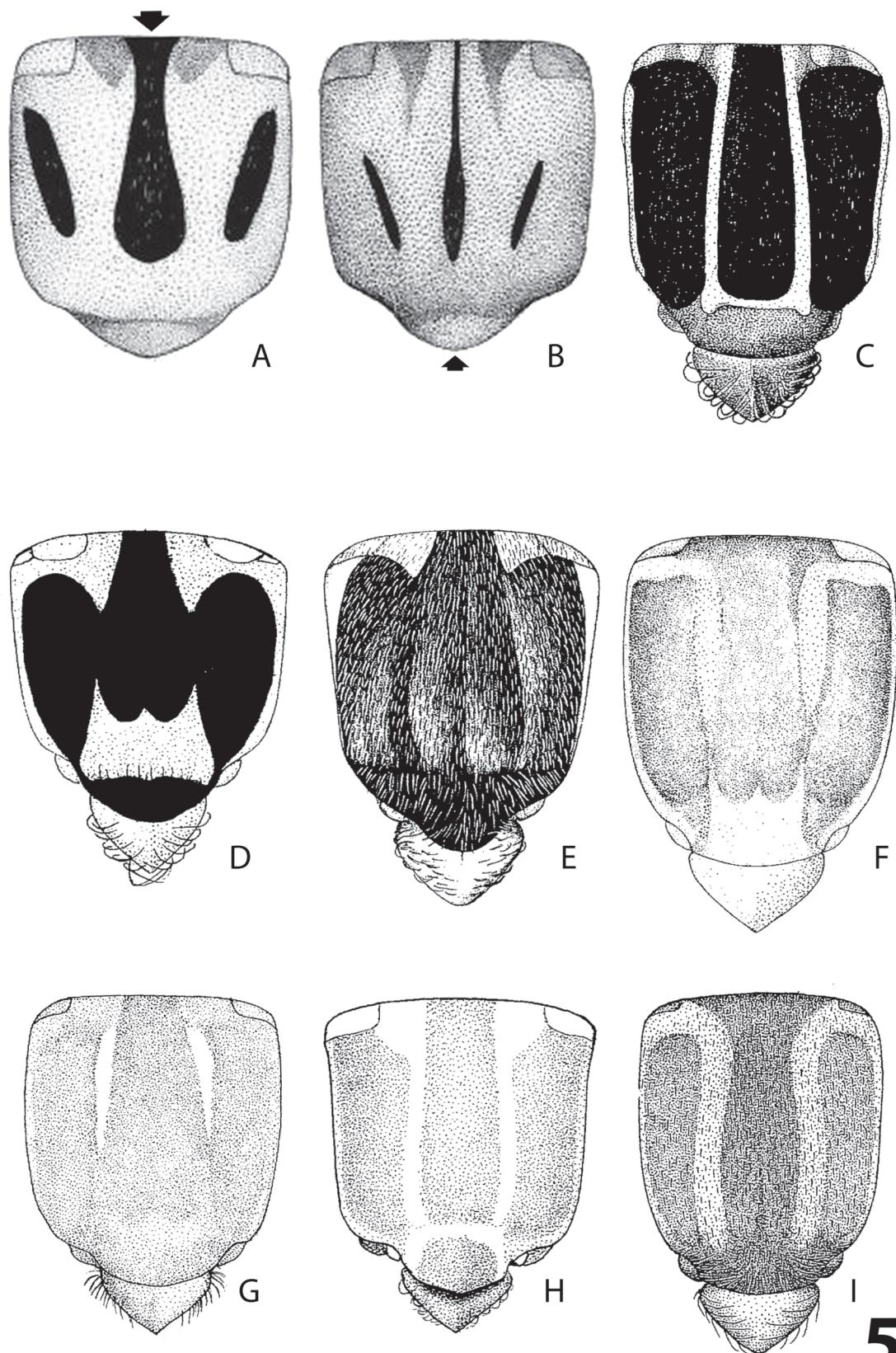


Figure 5. Scutum and scutellum [arrows indicate angle of incidence of light]. A. *Simulium (Cerqueirellum) amazonicum* Goeldi, 1905. B. *Simulium (Cerqueirellum) argentiscutum* Shelley & Dias, 1980. C. *Simulium (Coscaroniellum) quadrivittatum* Loew, 1862. D. *Simulium (Ectemnaspis) bicoloratum* Malloch, 1912. E. *Simulium (Ectemnaspis) mayuchuspi* Coscarón, 1990. F. *Simulium (Ectemnaspis) antillarum* Jennings, 1915. G. *Simulium (Ectemnaspis) furcillatum* Wygodzinsky & Coscarón, 1982. H. *Simulium (Psilopeltmia) samboni* Jennings, 1915. I. *Simulium (Psilopeltmia) escomeli* Roubaud, 1909.

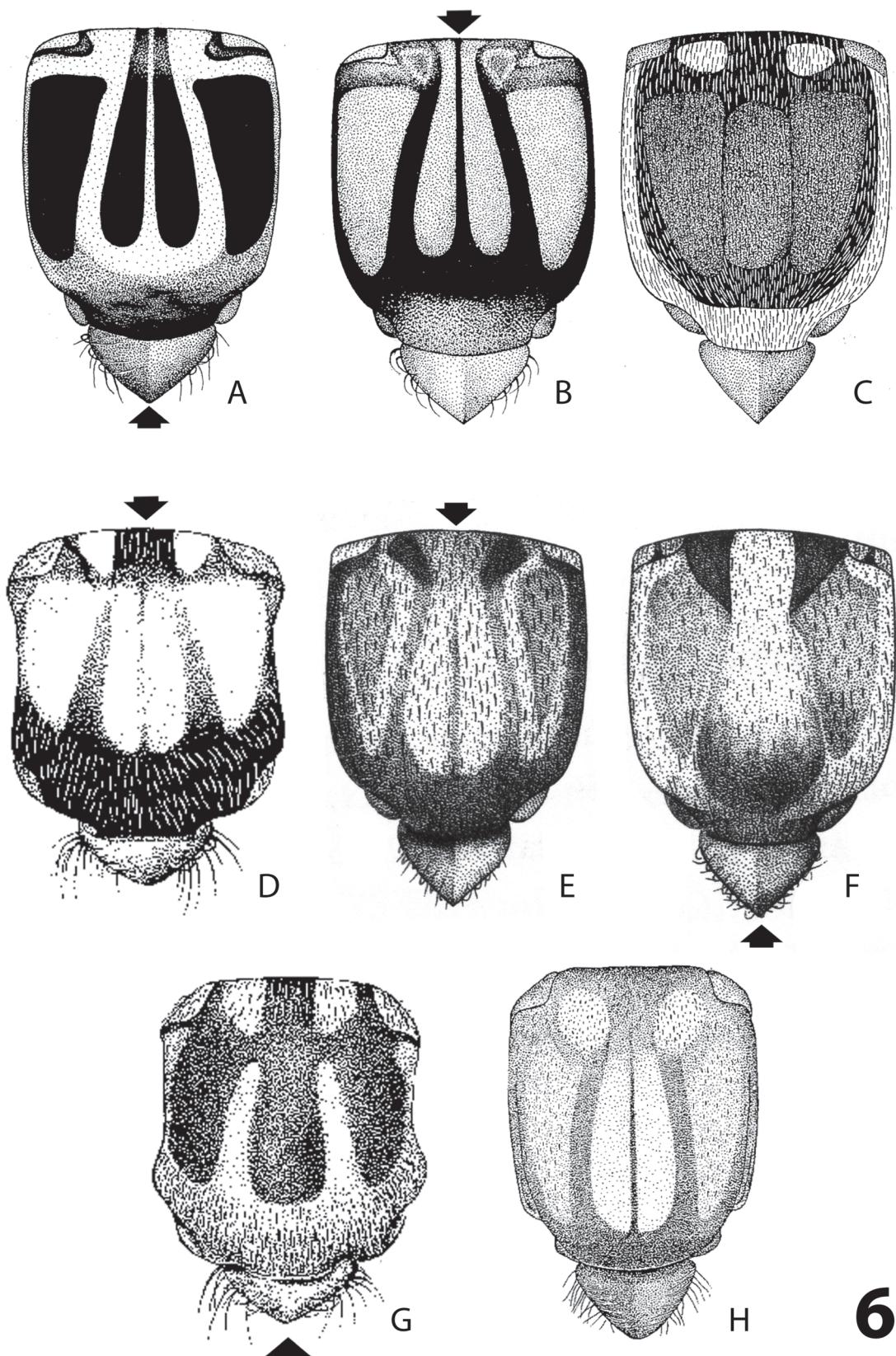


Figure 6. Scutum and scutellum [arrows indicate angle of incidence of light]. A-B. *Simulium (Aspathia) metallicum* Bellardi, 1859. C. *Simulium (Aspathia) putre* Cocarón & Matta, 1982. D-E. *Simulium (Hearlea) microbranchium* Dalmat, 1949. F-G. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. H. *Simulium (Trichodagmia) lahillei* (Paterson & Shannon, 1927).

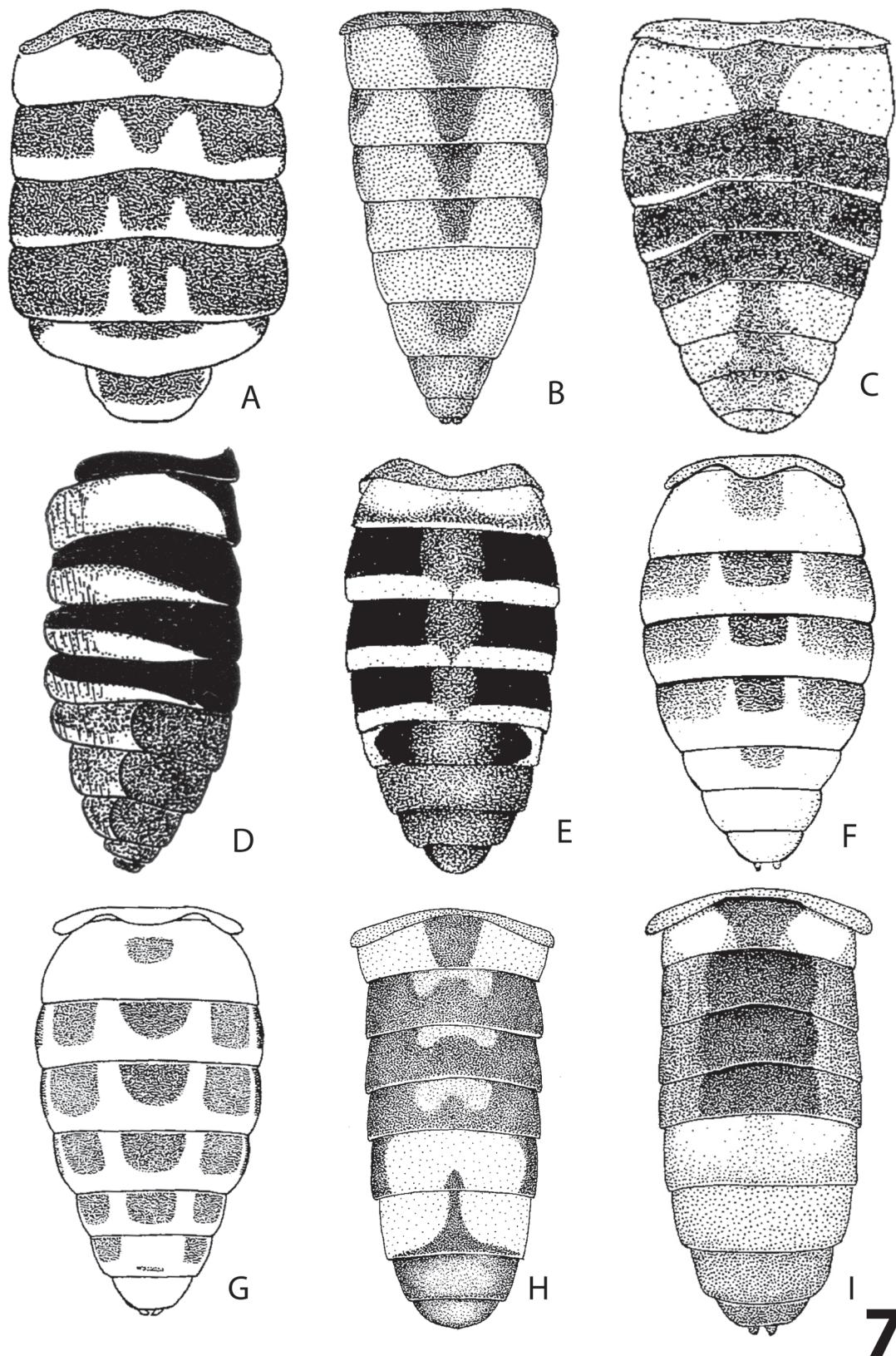


Figure 7. Abdomen, (A-C, E-I) dorsal view, (D) lateral view. A. *Simulium (Pternaspatha) nigristrigatum* (Enderlein, 1930). B. *Simulium (Pternaspatha) diamantinum* Coscarón & Coscarón Arias, 1996. C. *Simulium (Pternaspatha) annulatum* Philippi, 1865. D. *Simulium (Cerqueirellum) chaquense* Coscarón, 1971. E. *Simulium (Coscaroniellum) quadrivittatum* Loew, 1862. F. *Simulium (Psilopelmia) dugesi* Vargas, Martínez Palacios & Díaz Nájera, 1946. G. *Simulium (Psilopelmia) escomeli* Roubaud, 1909. H. *Simulium (Psilopelmia) blancasi* Wygodzinsky & Coscarón, 1970. I. *Simulium (Psilopelmia) tenuipes* Knab, 1914.

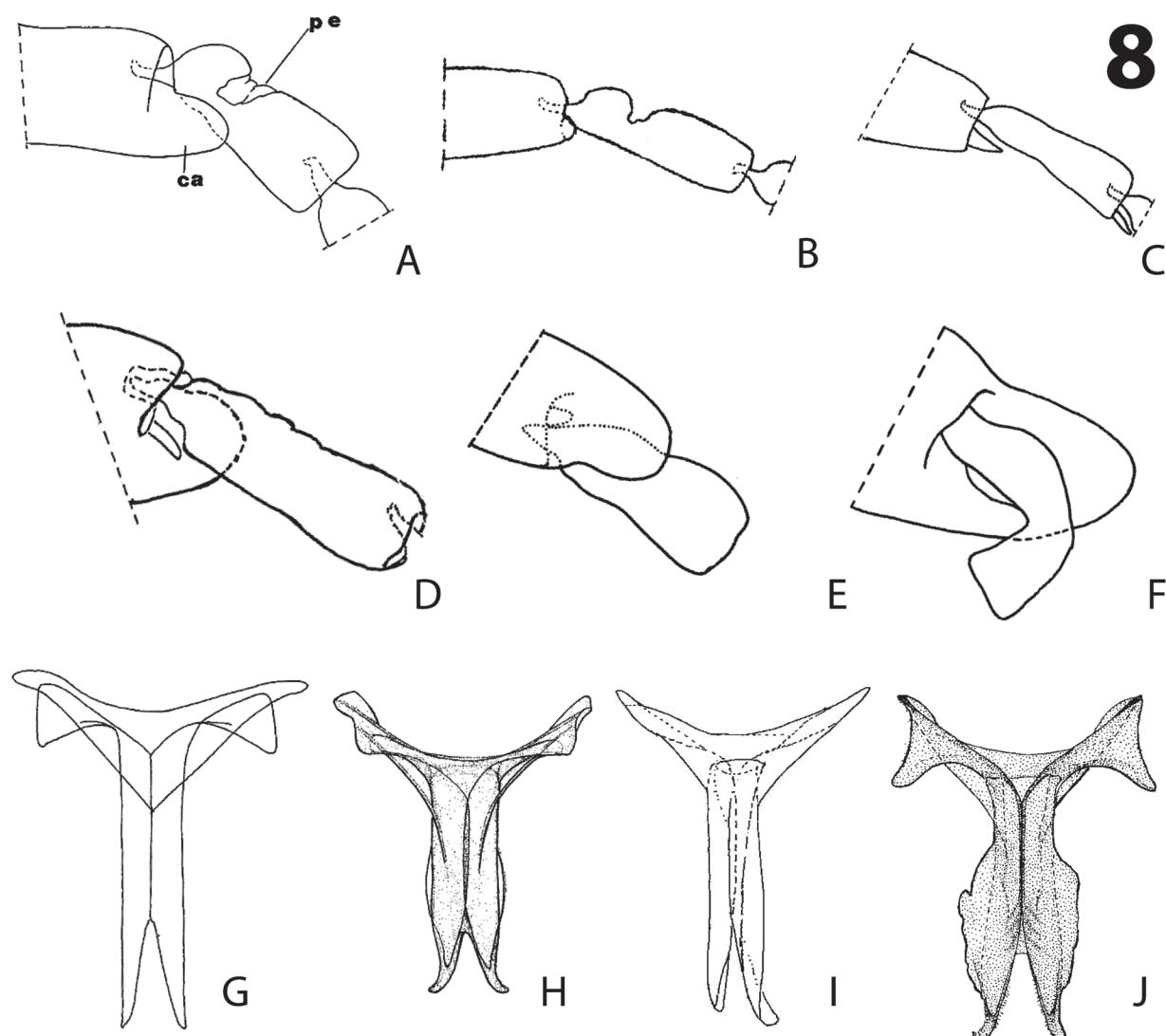


Figure 8. A-F. Apex of hind basitarsus and second tarsomere. A. *Simulium sp.* (ca: calcipala; pe: pedisulcus). B. *Simulium (Pternaspatha) philippii* Coscarón, 1976. C. *Pedrowygomyia punapi* (Wygodzinsky & Coscarón), 1989. D. *Gigantoidax igniculus* Coscarón & Wygodzinsky, 1962. E. *Gigantodax antarcticus* (Bigot, 1888). F. *Gigantodax multifilis* Wygodzinsky & Coscarón, 1989. G-J. Furcasternum. G. *Mayacnephia aguirrei* (Dalmat, 1949). H. *Paraustrosimulium anthracinum* (Bigot, 1888). I. *Kempfsimulium simplicicolor* (Lutz, 1910). J. *Lutzsimulium pernigrum* (Lutz, 1910).

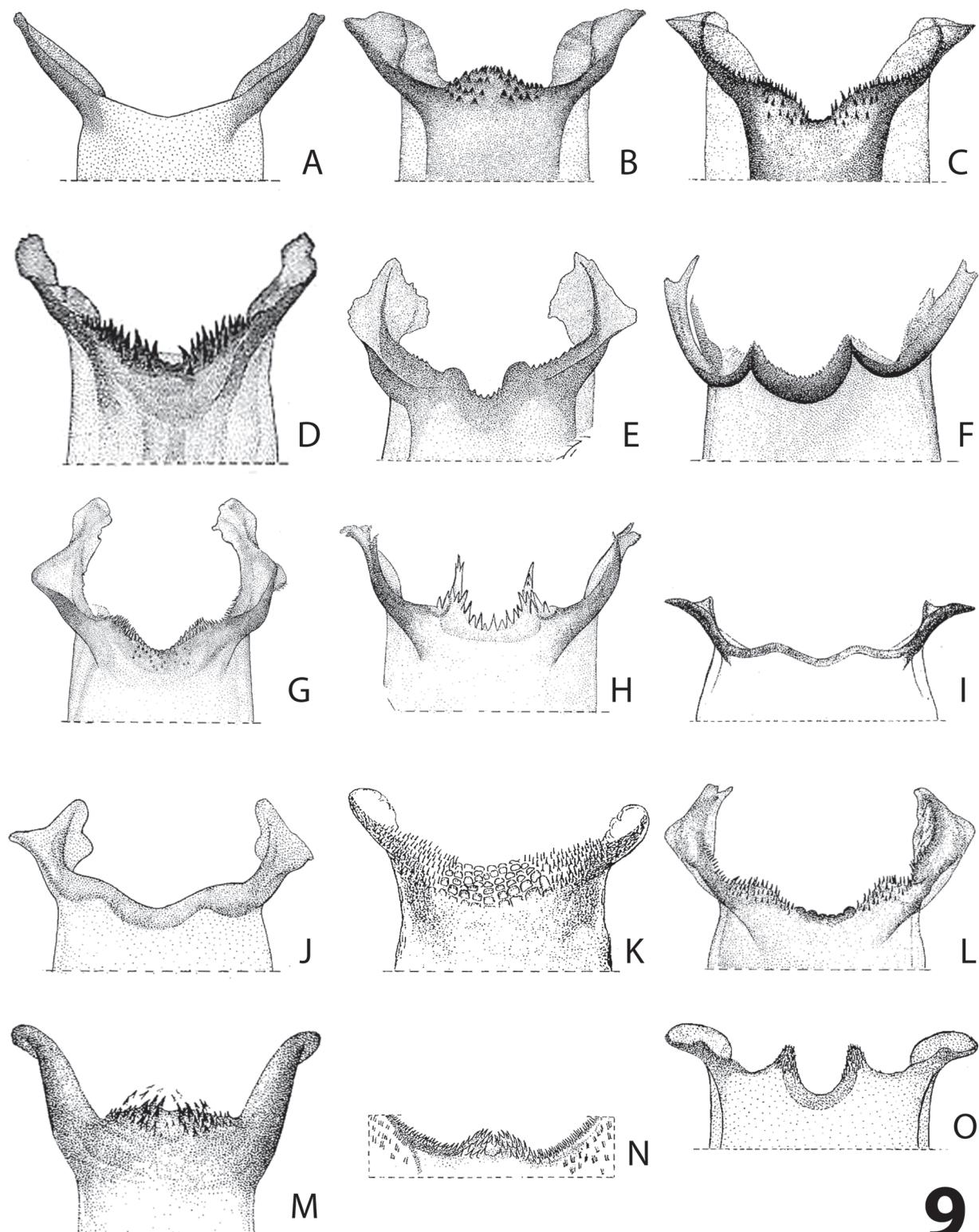


Figure 9. A-O. Basal portion of cibarium. A. *Simulium (Aspathia) putre* Coscarón & Matta, 1982. B. *Simulium (Inaequalium) subclavibranchium* Lutz, 1910. C. *Simulium (Psaroniocompsa) incrustatum* Lutz, 1910. D. *Simulium (Cerqueirellum) cuneatum* (Enderlein, 1936). E. *Simulium (Coscaroniellum) quadrifidum* Lutz, 1917. F. *Simulium (Coscaroniellum) cerradense* Coscarón, Cerqueira, Sato & La Salvia, 1992. G. *Simulium (Coscaroniellum) goeldii* Cerqueira & Mello, 1967. H. *Simulium (Coscaroniellum) quadrivittatum* Loew, 1862. I. *Simulium (Ectemnaspis) rubiginosum* (Enderlein, 1934). J. *Simulium (Ectemnaspis) arcabucense* Coscarón, 1990. K. *Simulium (Ectemnaspis) lutzianum* Pinto, 1931. L. *Simulium (Ectemnaspis) romanai* Wygodzinsky, 1951. M. *Simulium (Ectemnaspis) kabayanense* Ramírez Pérez & Vulcano, 1973. N. *Simulium (Ectemnaspis) perflavum* (Roubaud, 1906). O. *Simulium (Ectemnaspis) dinellii* (Joan, 1912).

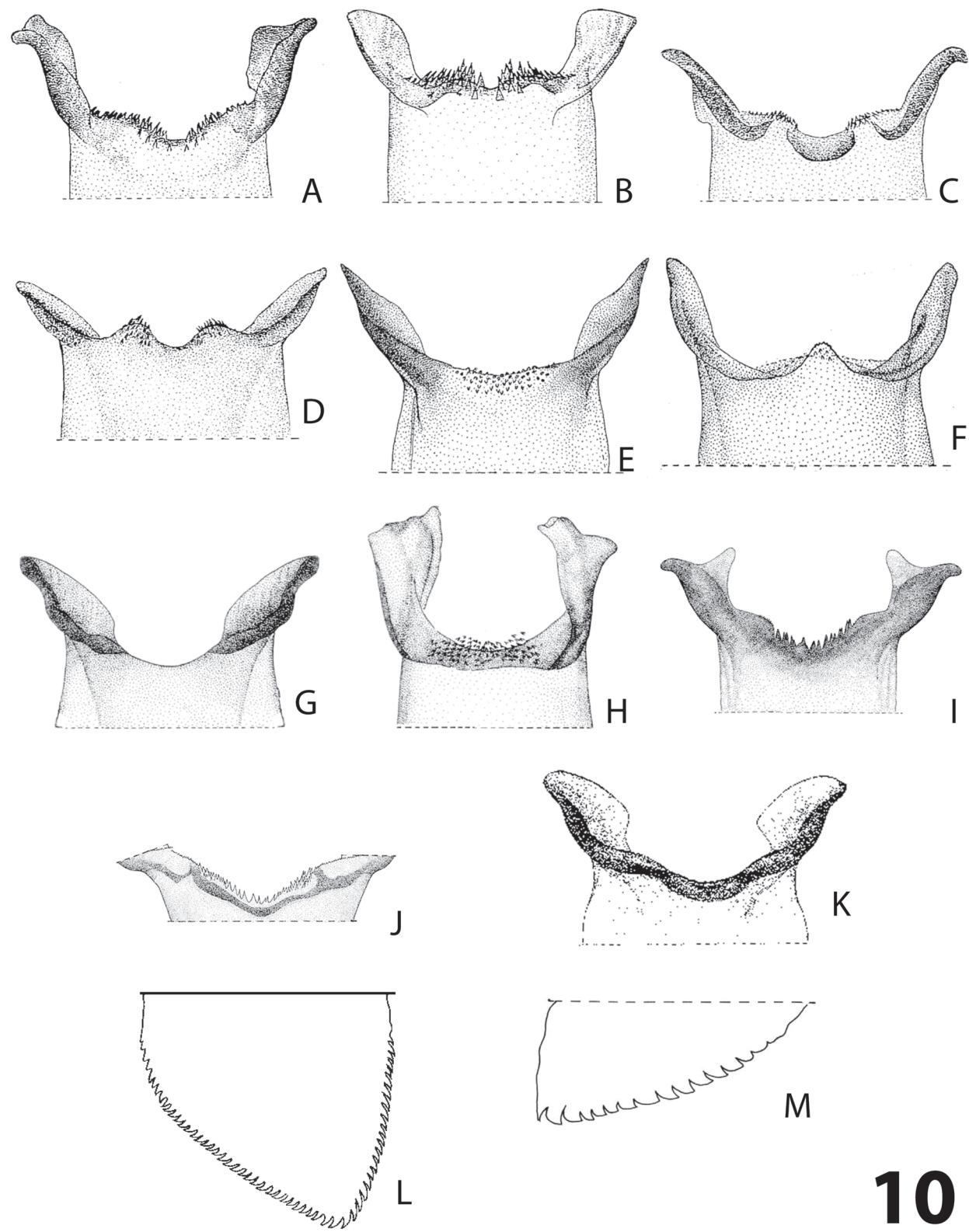
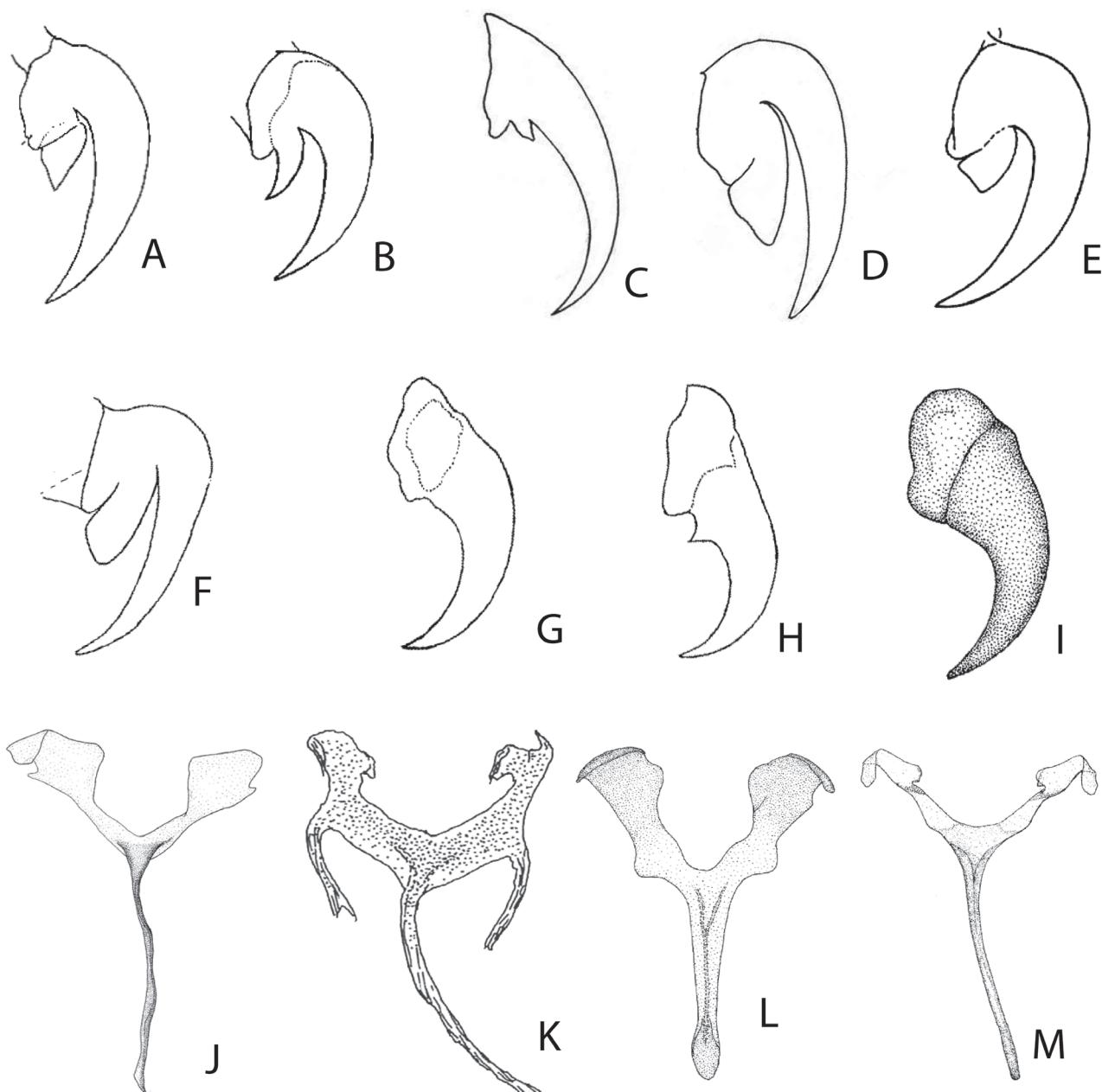


Figure 10. A-K. Basal portion of cibarium. A. *Simulium (Psilopelmia) dugesi* Vargas, Martínez Palacios & Díaz Nájera, 1946. B. *Simulium (Psilopelmia) panamense* Fairchild, 1940. C. *Simulium (Psilopelmia) trivittatum* Malloch, 1914. D. *Simulium (Psilopelmia) dandrettae* Vargas, Martínez Palacios & Díaz Nájera, 1946. E. *Simulium (Aspathia) matteabranchia* Anduze, 1947. F. *Simulium (Aspathia) bustosi* Vargas, Martínez Palacios & Díaz Nájera, 1946. G. *Simulium (Hemicnetha) rubri thorax* Lutz, 1909. H. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. I. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. J. *Simulium (Trichodagmia) huairayacu* Wygodzinsky, 1953. K. *Simulium (Thyrsopelma) orbitale* Lutz, 1910. L-M. Mandibles. L. *Araucnephia montana* (Philippi, 1865). M. *Kempfsimulium simplicicolor* (Lutz, 1910).



11

Figure 11. A-I. Claws. A. *Araucnephia iberaensis* Coscarón & Coscarón Arias, 2002. B. *Araucnephioides schlingeri* Wygodzinsky & Coscarón, 1973. C. *Pedrowygomyia cortesi* (Wygodzinsky & Coscarón, 1989). D. *Gigantodax brophyi* Wygodzinsky & Coscarón, 1962. E. *Gigantodax multifilis* Wygodzinsky & Coscarón, 1989. F. *Cnesiamima atroparva* (Edwards, 1931). G-H. *Simulium (Notolepria) exiguum* Roubaud, 1906. I. *Simulium (Psilopeltmia) tenuipes* Knab, 1914. J-M. Genital fork. J. *Araucnephia iberaensis* Coscarón & Coscarón Arias, 2002. K. *Gigantodax rufidulus* Wygodzinsky & Coscarón, 1989. L. *Paraustrosimulium anthracinum* (Bigot, 1888). M. *Kempfsimulium simplicicolor* (Lutz, 1910).

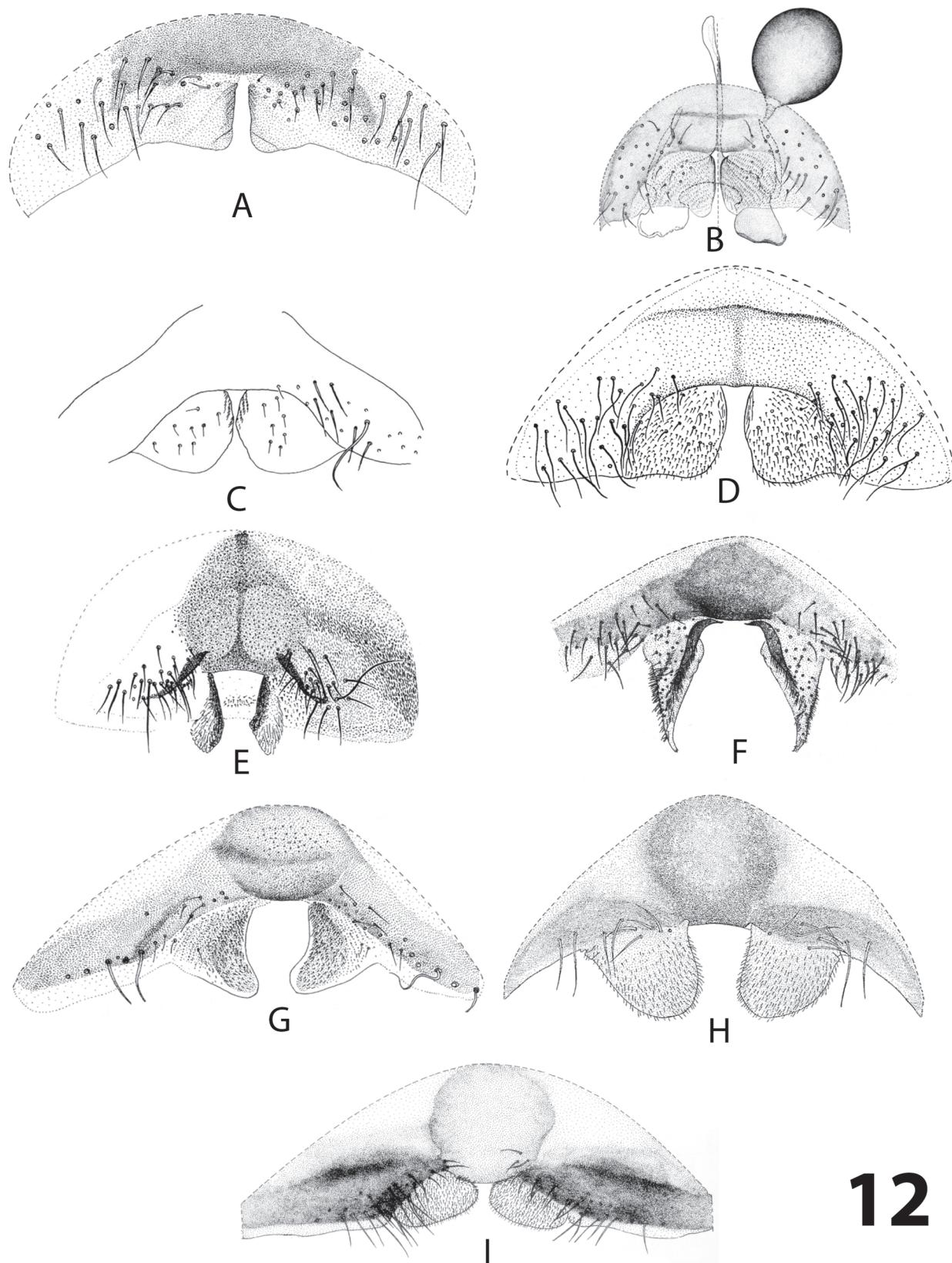


Figure 12. Sternite 8 and gonapophyses. A. *Araucnephia iberaensis* Coscarón & Coscarón Arias, 2002. B-C. *Gigantodax araucanius* (Edwards, 1931). D. *Gigantodax antarcticus* (Bigot, 1888). E. *Gigantodax igniculus* Wygodzinsky & Coscarón, 1962. F. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. G. *Simulium (Hearlea) capricorne* De Leon, 1945. H. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. I. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909.

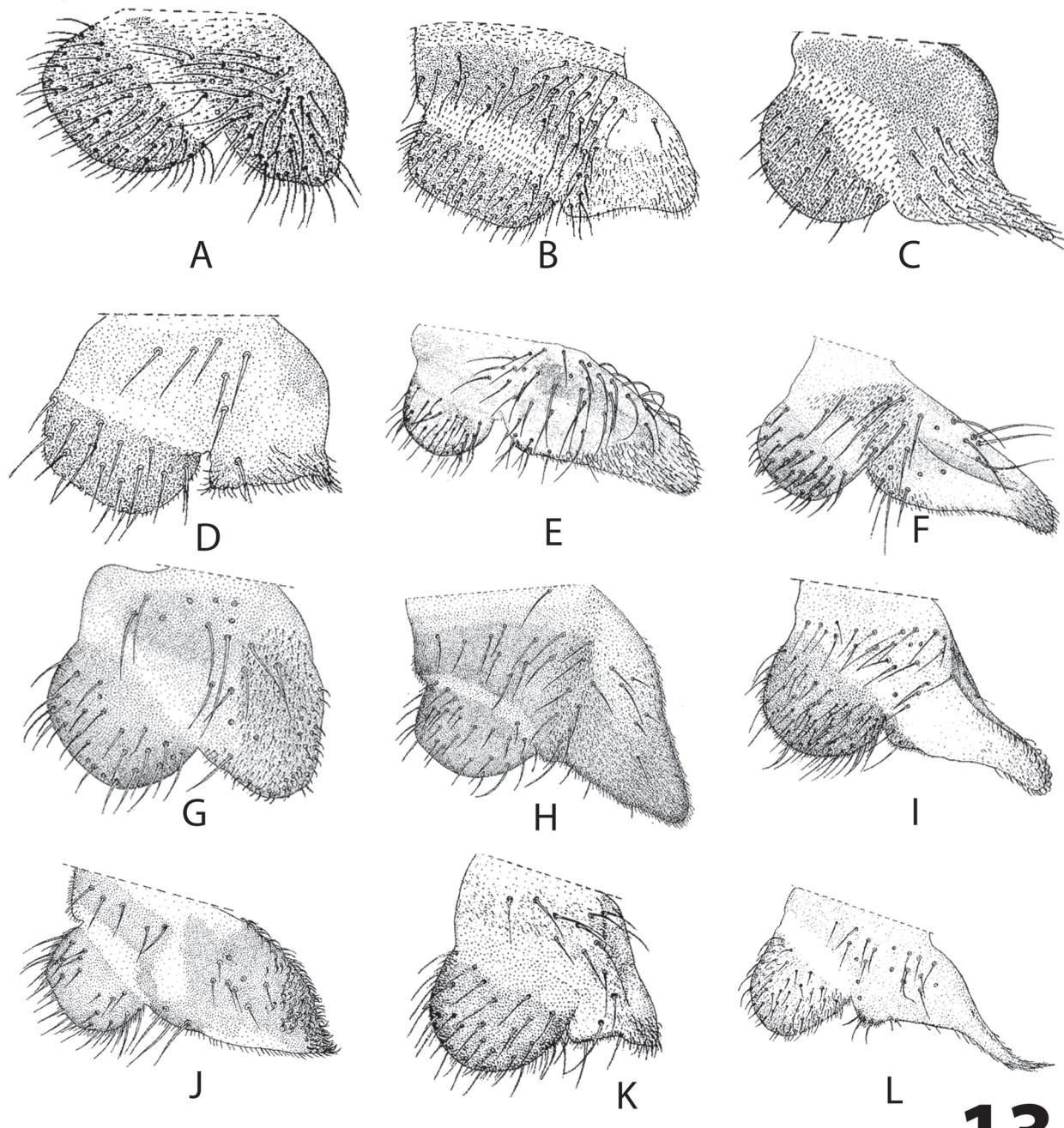
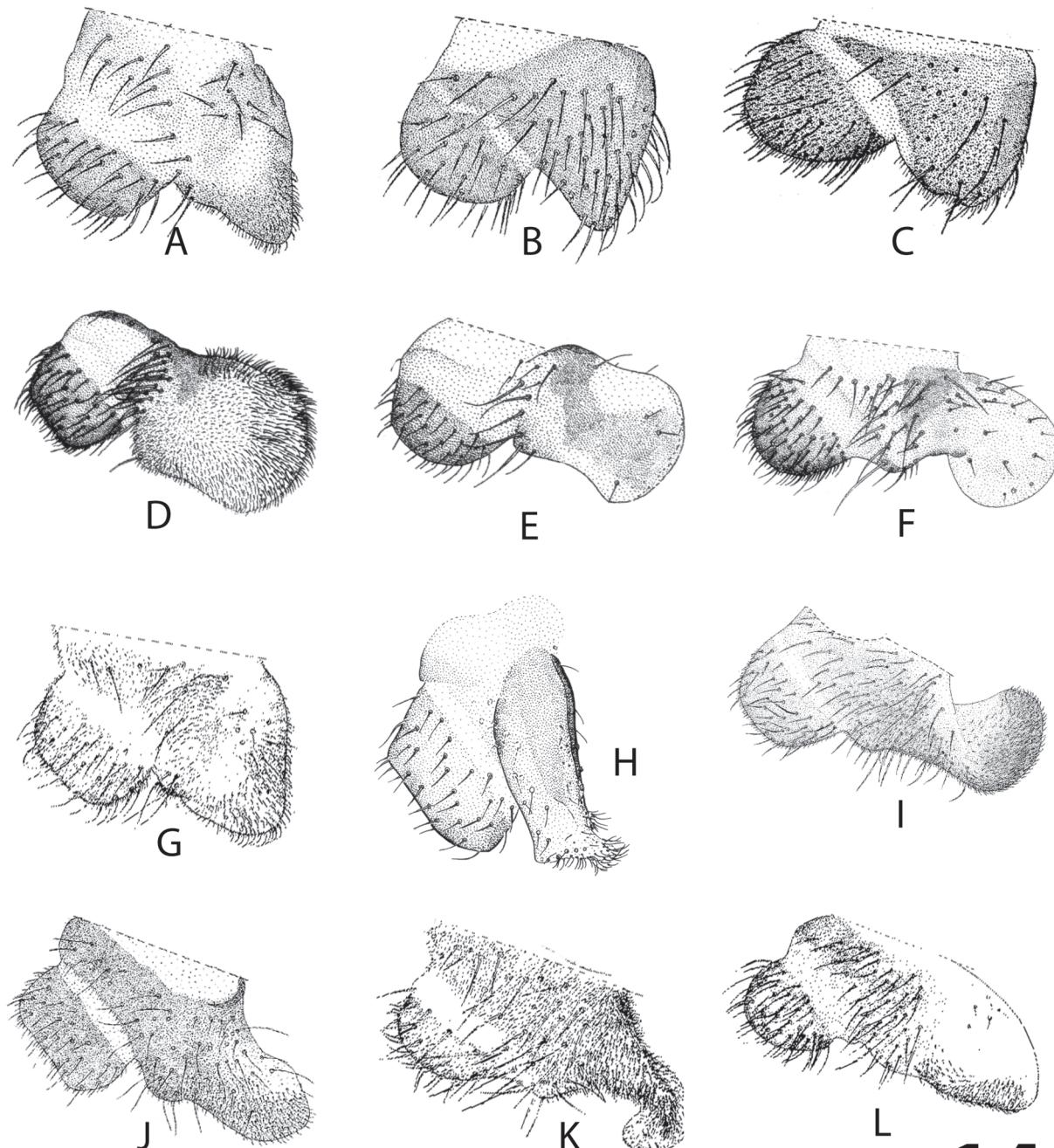
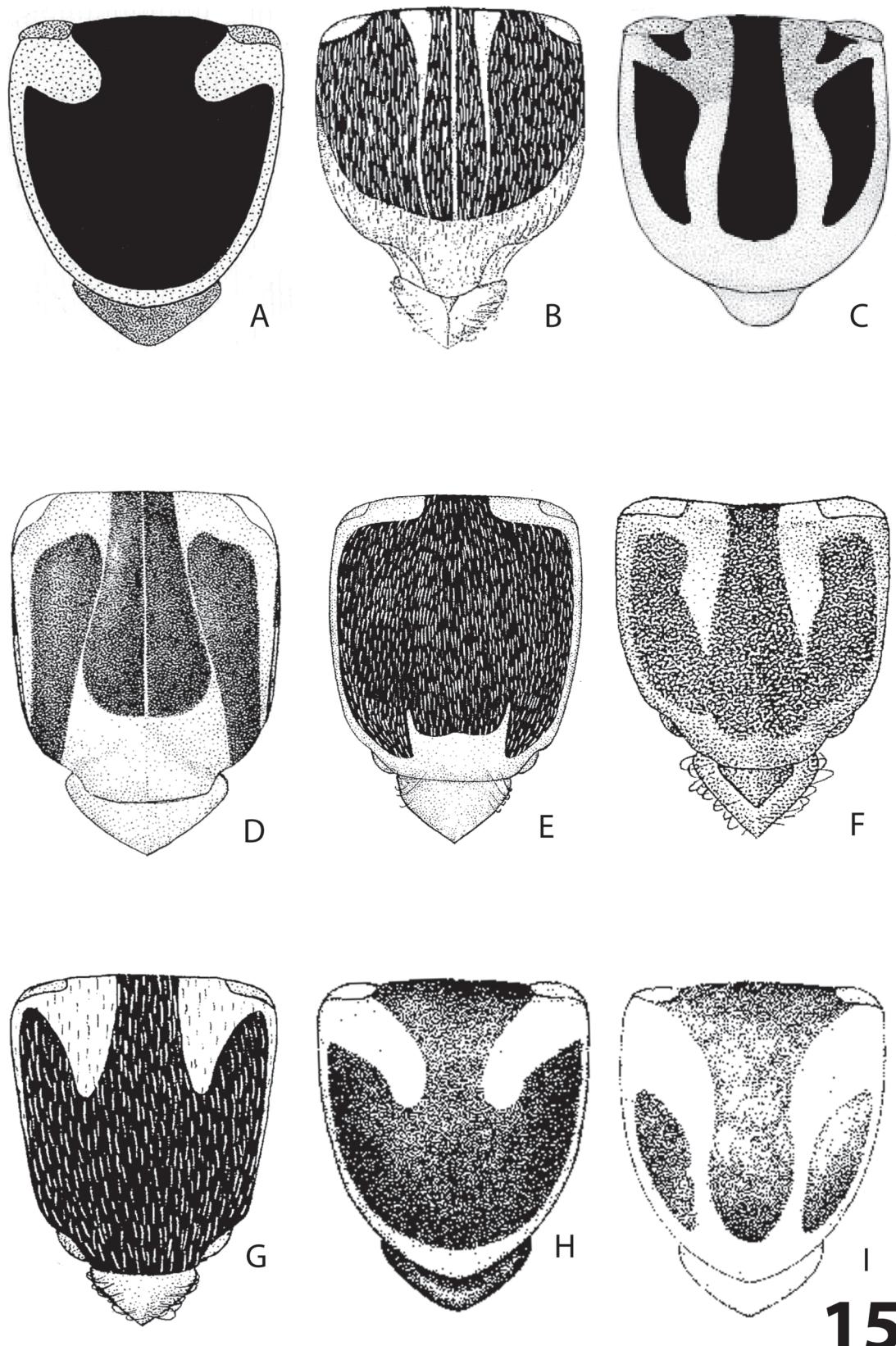


Figure 13. A-L. Cerci and paraprocts. A. *Araucnephia montana* (Philippi, 1865). B. *Simulium (Pternasptha) annulatum* Philippi, 1865. C. *Simulium (Notolepria) exiguum* Roubaud, 1906. D. *Simulium (Notolepria) gonzalezi* Vargas & Díaz Nájera, 1953. E. *Simulium (Chirostilbia) pertinax* Kollar, 1832. F. *Simulium (Chirostilbia) serratum* Coscarón, 1981. G. *Simulium (Inaequalium) travassosi* d'Andretta & d'Andretta Jr., 1947. H. *Simulium (Ectemnaspis) furcillatum* Wygodzinsky & Coscarón, 1982. I. *Simulium (Ectemnaspis) perflavum* (Roubaud, 1906). J. *Simulium (Ectemnaspis) rorotaense* Floch & Abonnenc, 1946. K. *Simulium (Psilopeltmia) dugesii* Vargas, Martínez Palacios & Díaz Nájera, 1946. L. *Simulium (Psilopeltmia) trivittatum* Malloch, 1914.



14

Figure 14. A-L. Cerci and paraprocts. A. *Simulium (Psilopelmia) tenuipes* Knab, 1914. B. *Simulium (Aspathia) metallicum* Bellardi, 1859. C. *Simulium (Aspathia) matteabranchia* Anduze, 1947. D. *Simulium (Hemicnetha) rubrithorax* Lutz, 1909. E. *Simulium (Hemicnetha) pulverulentum* Knab, 1914. F. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. G. *Simulium (Hemicnetha) oviedoii* Ramírez Pérez, 1971. H. *Simulium (Hearlea) capricornis* De León, 1945. I. *Simulium (Trichodagmia) lahillei* (Paterson & Shannon, 1927). J. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. K. *Simulium (Thrysopelma) hirtipupa* Lutz, 1910. L. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909.



15

Figure 15. A-I. Scutum and scutellum. A. *Simulium (Chirostilbia) acarayense* Coscarón & Wygodzinsky, 1972. B. *Simulium (Psaroniocompsa) limbatum* Knab, 1915. C. *Simulium (Cerqueirellum) chaquense* Coscarón, 1971. E. *Simulium (Ectemnaspis) romanai* Wygodzinsky, 1951. F. *Simulium (Psilopeltia) dugesi* Vargas, Martínez Palacios & Díaz Nájera, 1946. G. *Simulium (Psilopeltia) escomeli* Roubaud, 1909. H. *Simulium (Thrysopelma) orbitale* Lutz, 1910. I. *Simulium (Thrysopelma) guianense* Wise, 1911.

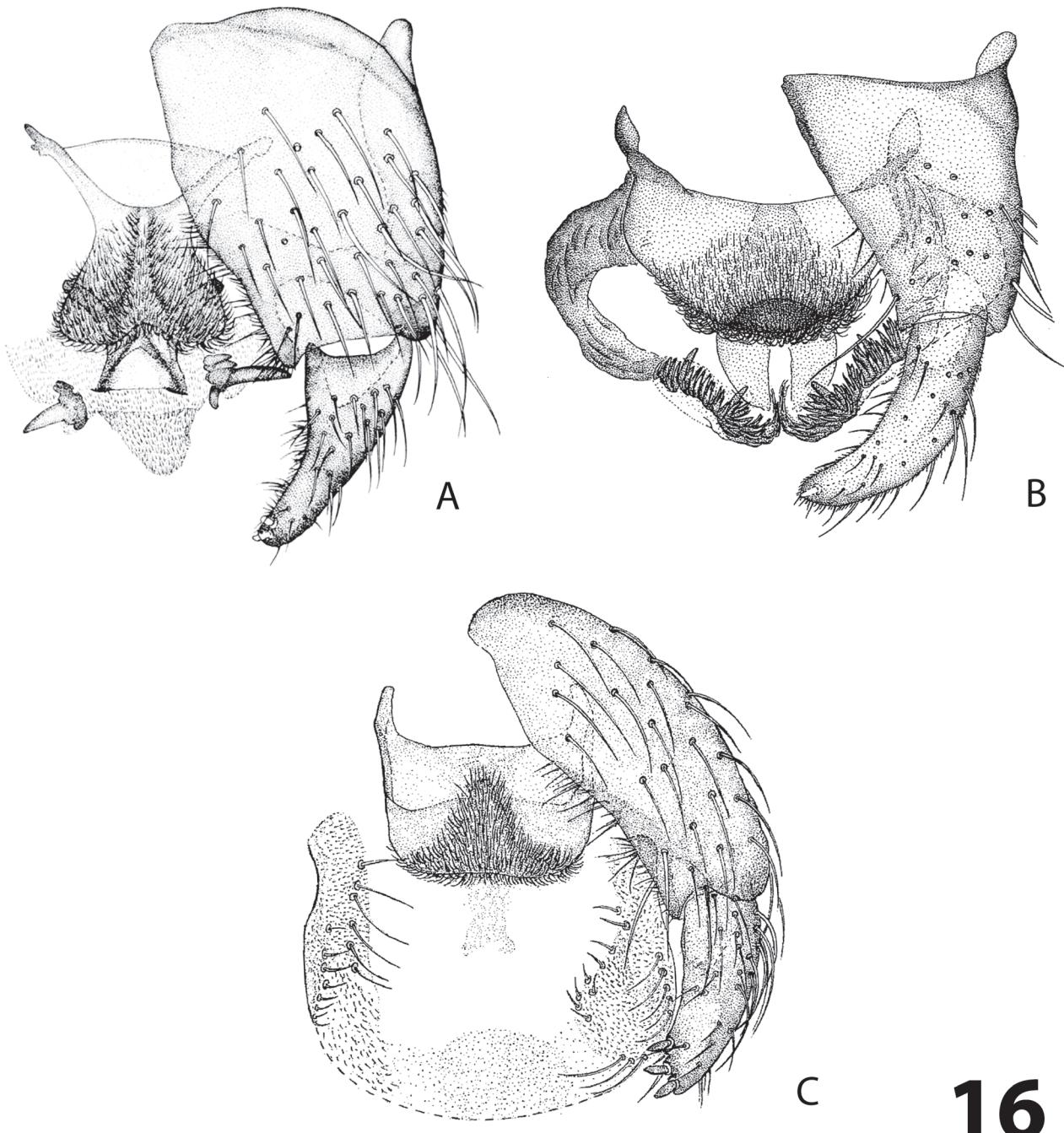


Figure 16. A-C. Terminalia, showing gonocoxite, gonostylus, ventral plate, median sclerite and endoparameres (ventral view). A. *Gigantodax araucanius* (Edwards, 1931). B. *Simulium (Psilopeltmia) tenuipes* Knab, 1914. C. *Lutzsimulium hirticosta* (Lutz, 1909).

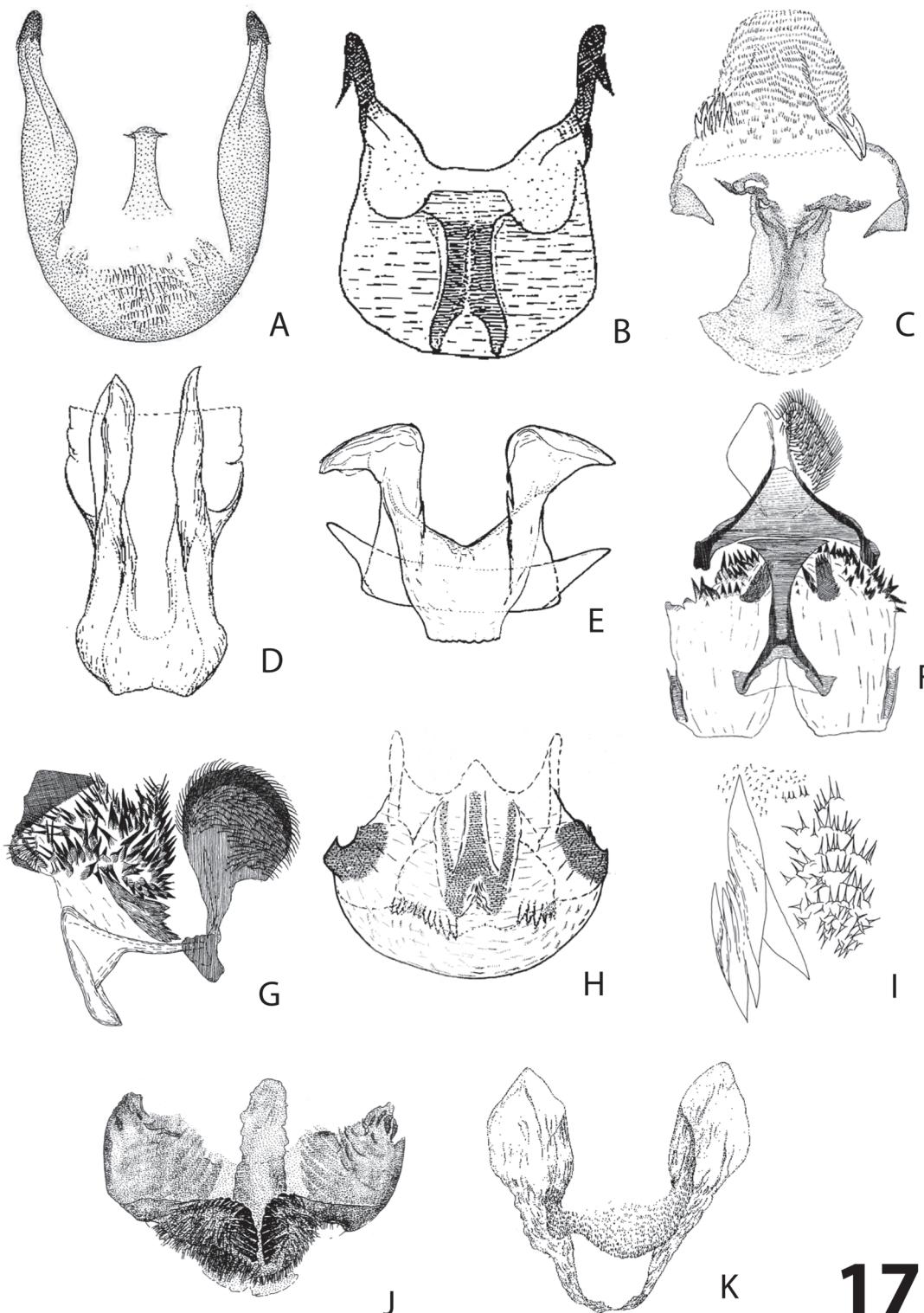


Figure 17. A. *Mayacnephia aguirrei* (Dalmat, 1949), endoparameres with portion of median sclerite and aedeagal membrane. B. *Mayacnephia roblesi* (De León, 1943), median sclerite positioned on ventral plate. C. *Araucnephia iberaensis* Coscarón & Coscarón Arias, 2002, median sclerite, emdoparameres with hooks and aedeagal membrane. D. *Araucnephia montana* (Philippi, 1865), median sclerite. E. *Araucnephioides schlingeri* Wygodzinsky & Coscarón, 1973, median sclerite. F-G. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962, aedeagus with spinules, ventral plate and median sclerite (F) and same in ventral view (G). H. *Paraustrosimilium anthracinum* (Bigot, 1888), aedeagus with median sclerite, endoparameres and ventral plate, visible through it. I. *Gigantodax araucanius* (Edwards, 1931), endoparameres with small spines on aedeagus. J. *Simulium (Hemicnetha) seriatum* Knab, 1914, endoparameres and median sclerite. K. *Simulium (Trichodagmia) lahillei* (Paterson & Shannon, 1927), endoparameres and aedeagus.

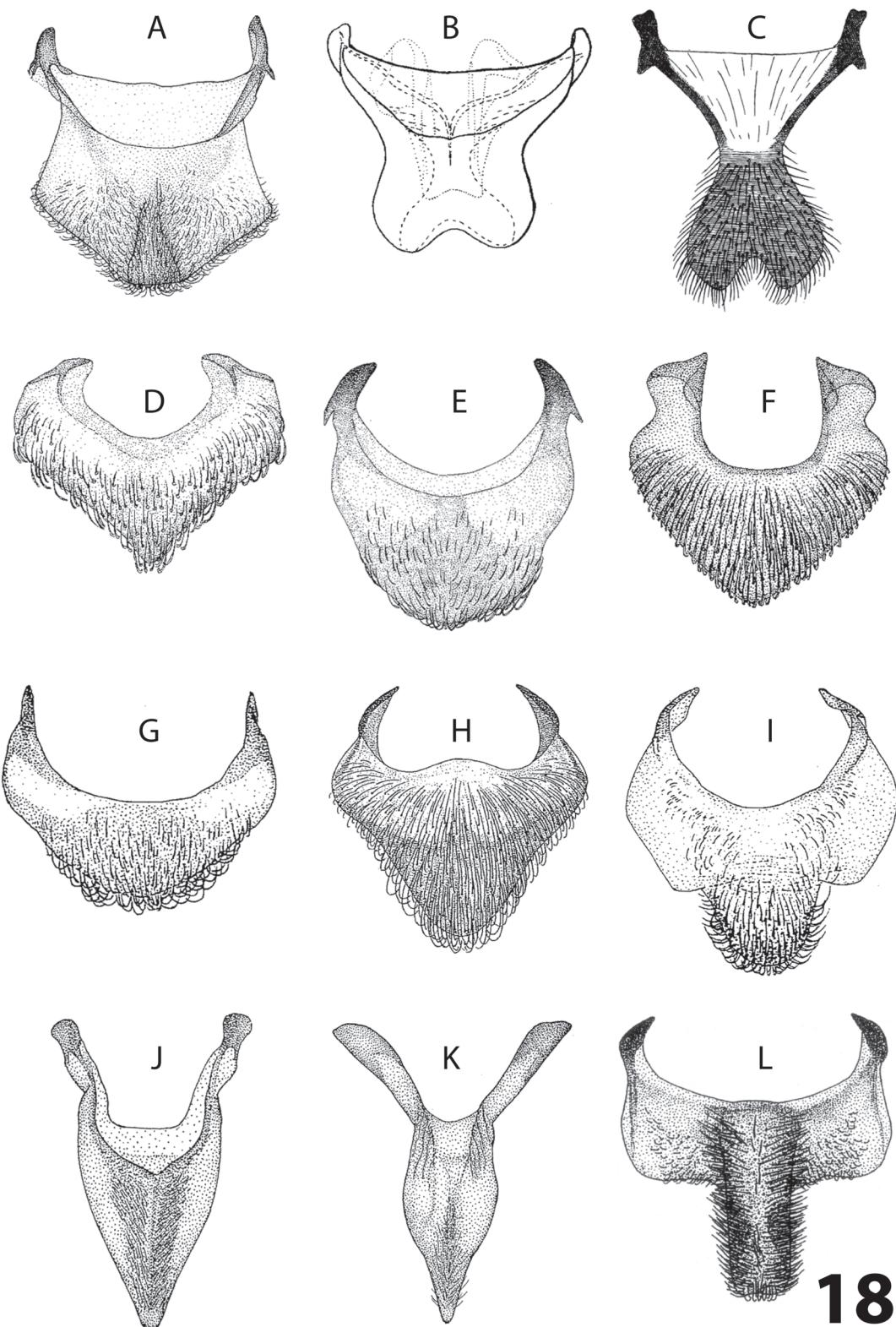
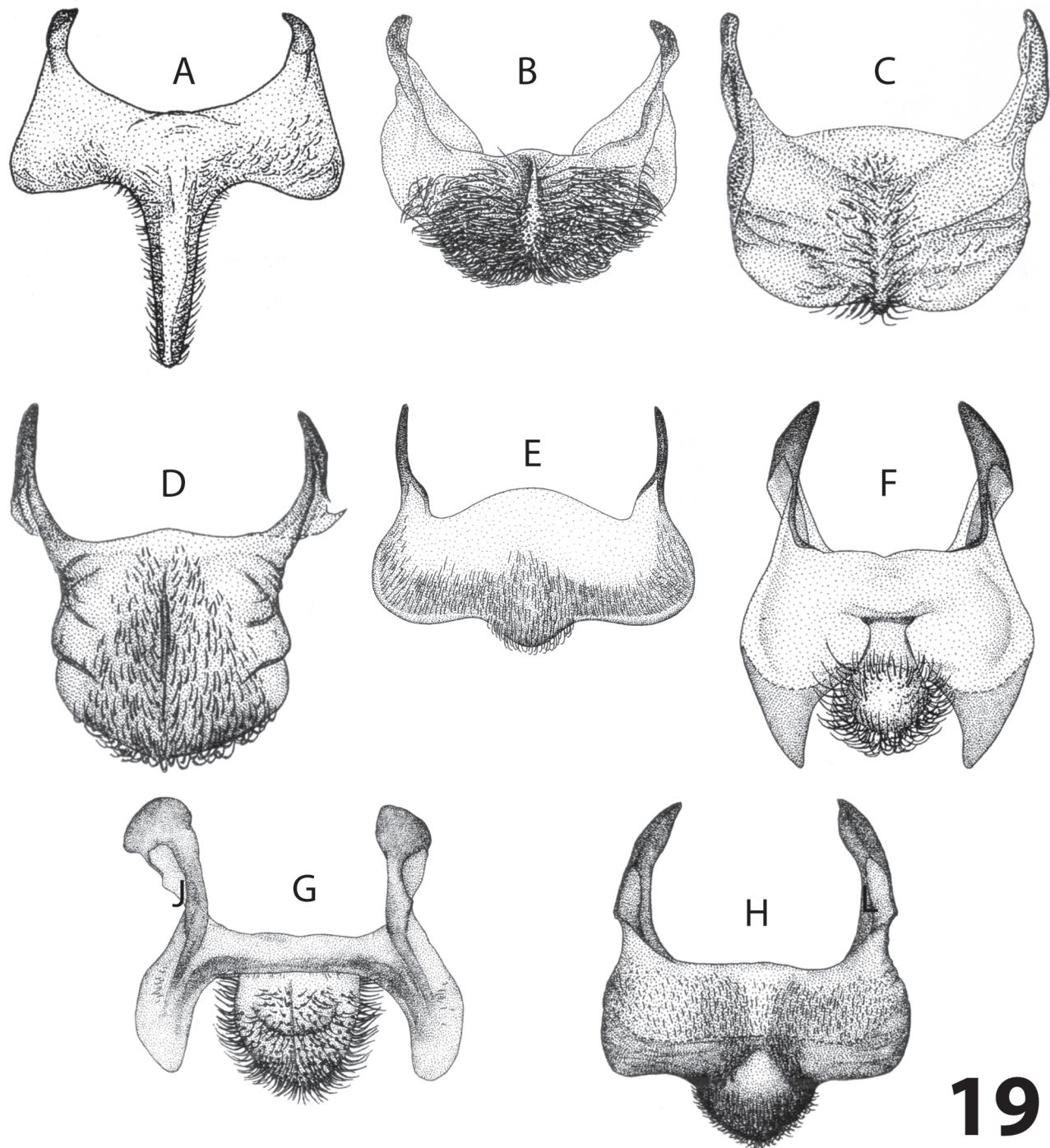


Figure 18. A-L. Male terminalia ventral plate. A. *Araucnephia iberaensis* Coscarón & Coscarón Arias, 2002. B. *Araucnephioides schlengeri* Wygodzinsky & Coscarón, 1973. C. *Gigantodax igniculus* Cocarón & Wygodzinsky, 1962. D. *Simulium (Inaequalium) subnigrum* Lutz, 1910. E. *Simulium (Inaequalium) subclavibranchium* Lutz, 1910. F. *Simulium (Inaequalium) travassosi* d'Andretta & d'Andretta Jr., 1947. G. *Simulium (Psilopeltmia) dugesi* Vargas, Martínez Palacios & Díaz Nájera, 1946. H. *Simulium (Psilopeltmia) escomeli* Roubaud, 1909. I. *Simulium (Psilopeltmia) veracruzanum* Vargas, Martínez Palacios & Díaz Nájera, 1946. J. *Simulium (Aspathia) hunteri* Malloch, 1914. K. *Simulium (Aspathia) matteabranchia* Anduze, 1947. L. *Simulium (Hemicnetha) rubrihorax* Lutz, 1909.



19

Figure 19. Male terminalia ventral. A. *Simulium (Hemicnetha) virgatum* Coquillett, 1902. B. *Simulium (Hemicnetha) seriatum* Knab, 1914. C. *Simulium (Hearlea) delatorrei* Dalmat, 1950. D. *Simulium (Hearlea) larvispinosum* De León, 1948. E. *Simulium (Trichodagnia) huairayacu* Wygodzinsky, 1953. F. *Simulium (Thrysopelma) orbitale* Lutz, 1910. G. *Simulium (Thrysopelma) guianense* Wise, 1911. H. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909.

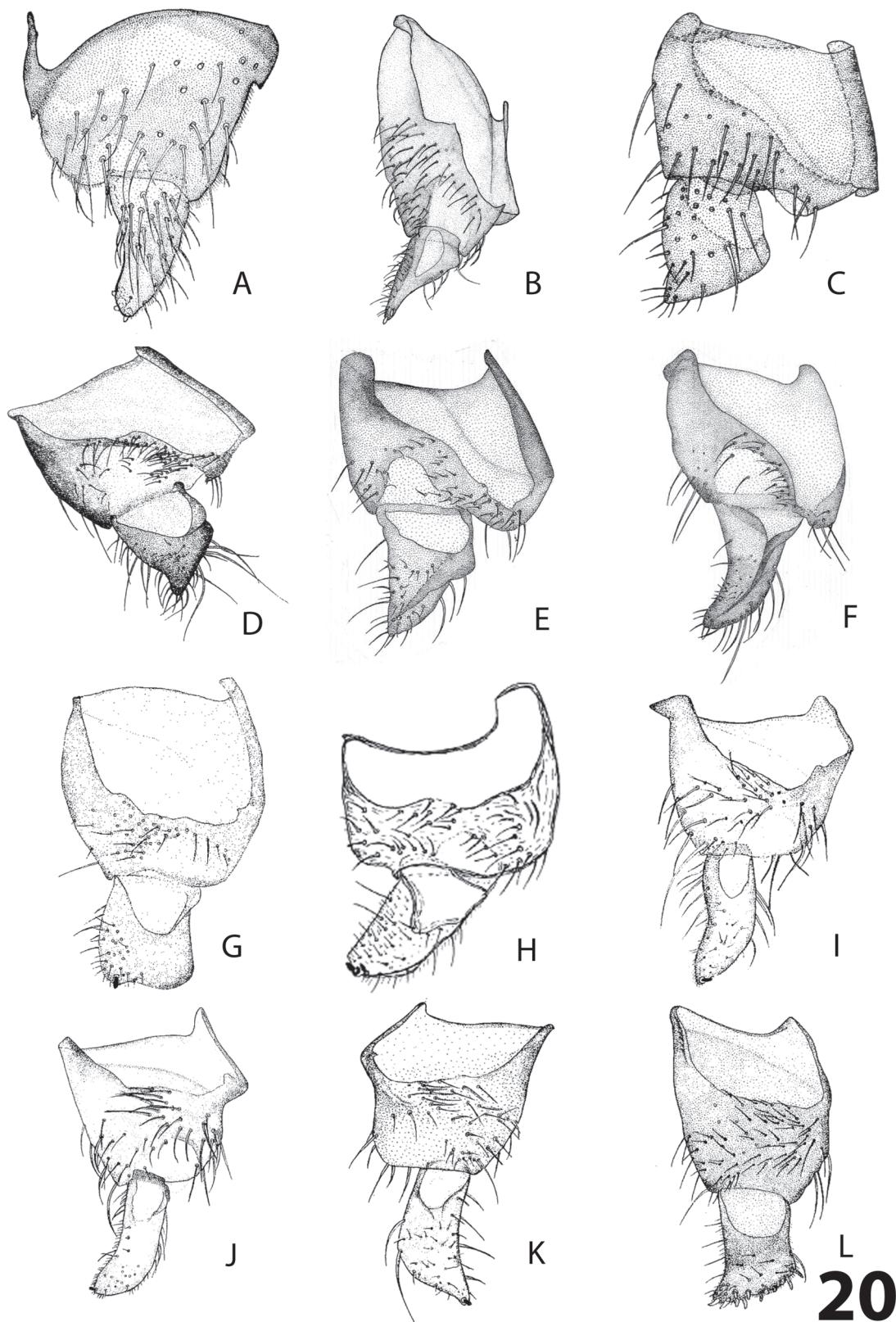


Figure 20. Male gonocoxite and gonostylus. A. *Lutzsimulium hirticosta* (Lutz, 1909). B. *Kempf simulium simplicicolor* (Lutz, 1909). C. *Simulium (Chirostilbia) pertinax* Kollar, 1832. D. *Simulium (Chirostilbia) serratum* Coscarón, 1981. E. *Simulium (Chirostilbia) acarayense* Coscarón & Wygodzinsky, 1972. F. *Simulium (Chirostilbia) subpallidum* Lutz, 1910. G. *Simulium (Psaroniocompsa) auristriatum* Lutz, 1910. H. *Simulium (Ectemnaspis) chaquense* Coscarón, 1971. I. *Simulium (Ectemnaspis) arcabucense* Coscarón, 1990. J. *Simulium (Ectemnaspis) tunja* Coscarón, 1990. K. *Simulium (Ectemnaspis) romanai* Wygodzinsky, 1951. L. *Simulium (Ectemnaspis) wolffhuegeli* (Enderlein, 1922).

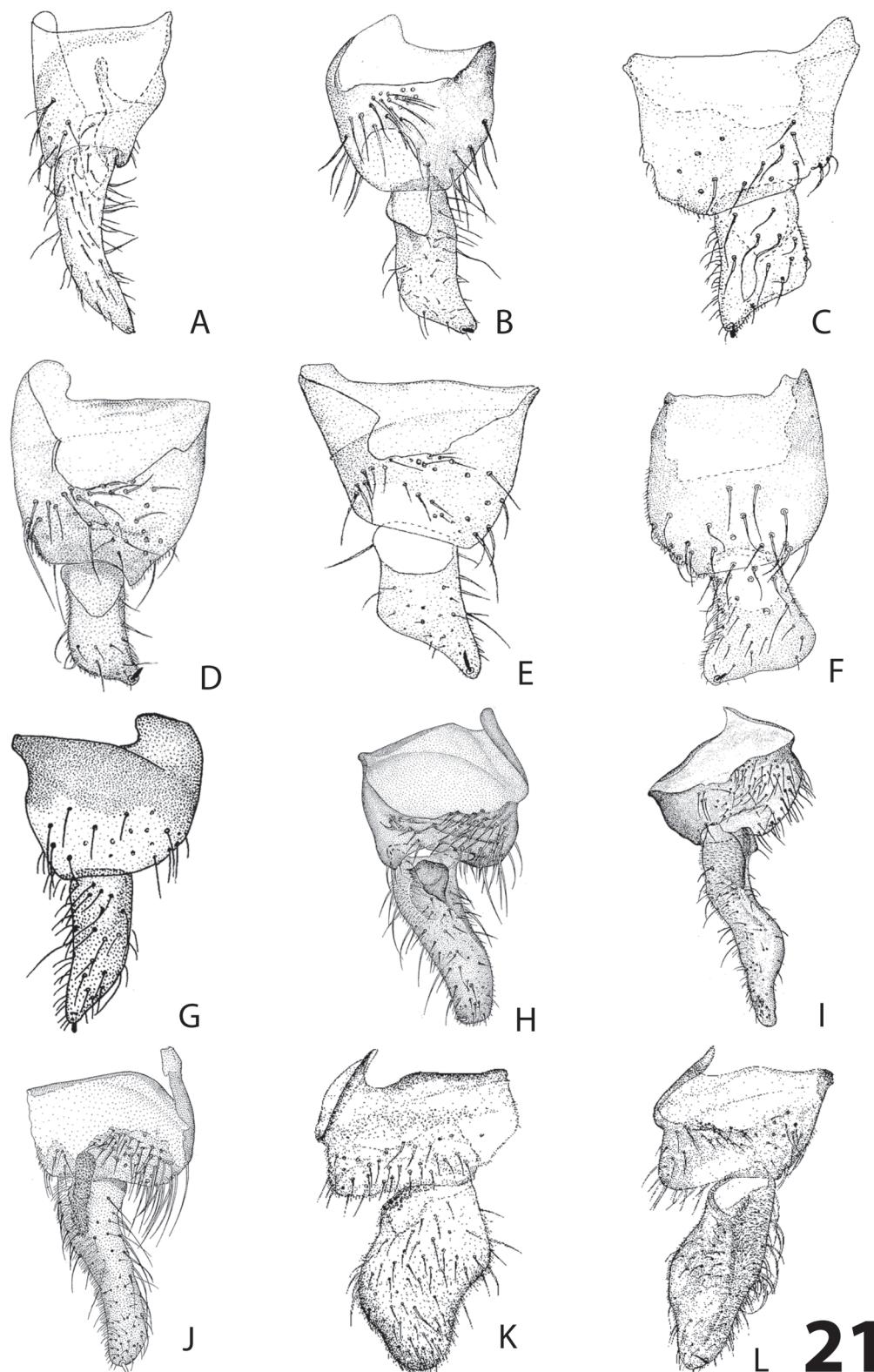


Figure 21. A-L. Male gonocoxite and gonostylus. A. *Simulium (Ectemnaspis) perflavum* (Roubaud, 1906). B. *Simulium (Ectemnaspis) rorotaense* Floch & Abonnenc, 1946. C. *Simulium (Psilopelmia) dugesi* Vargas, Martínez Palacios & Díaz Nájera, 1946. D. *Simulium (Psilopelmia) iracouboense* Floch & Abonnenc, 1946. E. *Simulium (Psilopelmia) haematopotum* Malloch, 1914. F. *Simulium (Psilopelmia) dandrettae* Vargas, Martinez Palacios & Díaz Nájera, 1946. G. *Simulium (Psilopelmia) blancasi* Wygodzinsky & Coscarón, 1970. H. *Simulium (Aspathia) metallicum* Bellardi, 1859. I. *Simulium (Aspathia) putre* Coscarón & Matta, 1982. J. *Simulium (Aspathia) wygoi* Coscarón, Ibáñez Bernal & Coscarón Arias, 1999. K. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. L. *Simulium (Hemicnetha) seriatum* Knab, 1914.

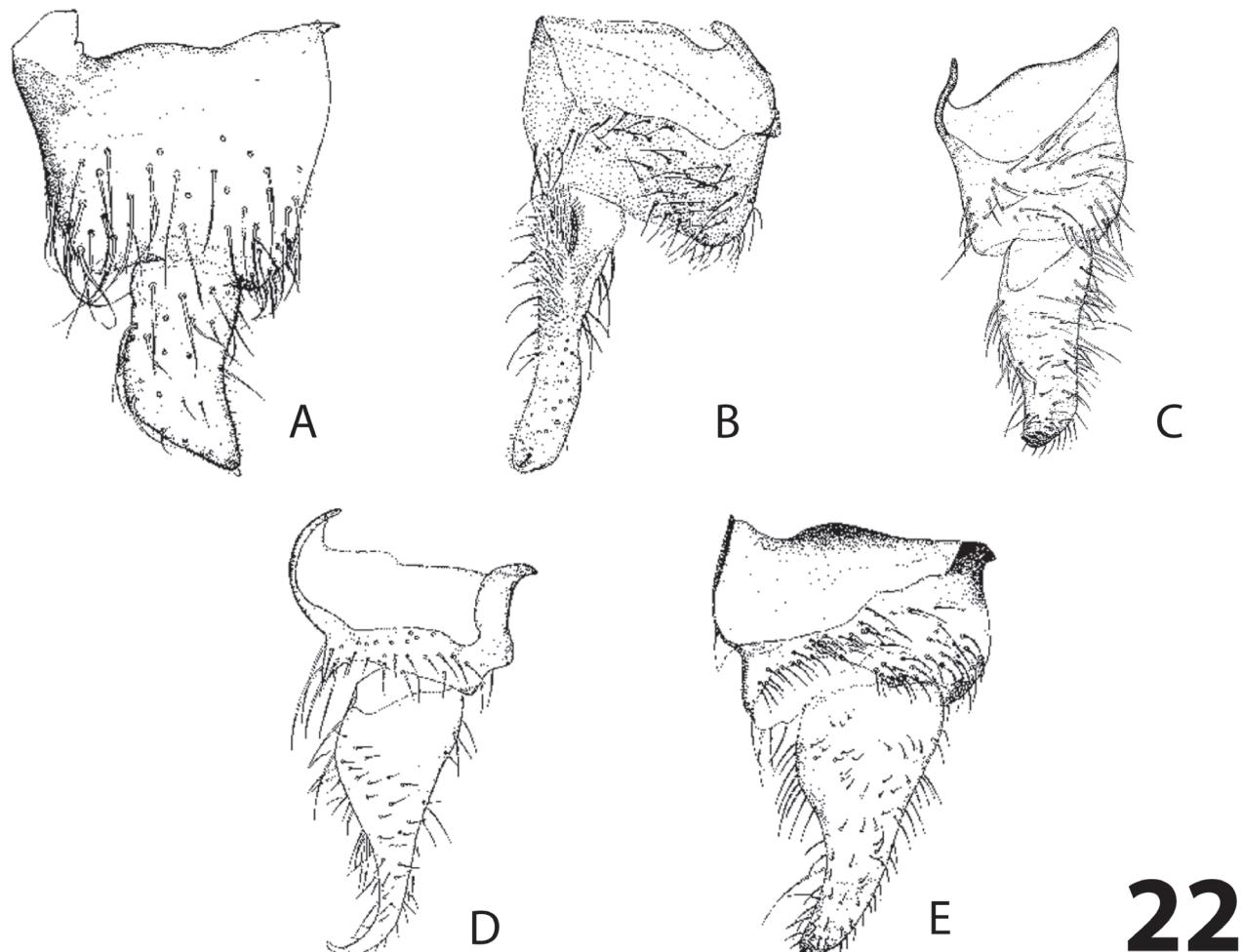


Figure 22. A-E. Male gonocoxite and gonostylus. A. *Simulium (Hemicnetha) oviedoi* Ramírez Pérez, 1971. B. *Simulium (Hearlea) delatorrei* Dalmat, 1950. C. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. D. *Simulium (Thrysopelma) orbitale* Lutz, 1910. E. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909.

22

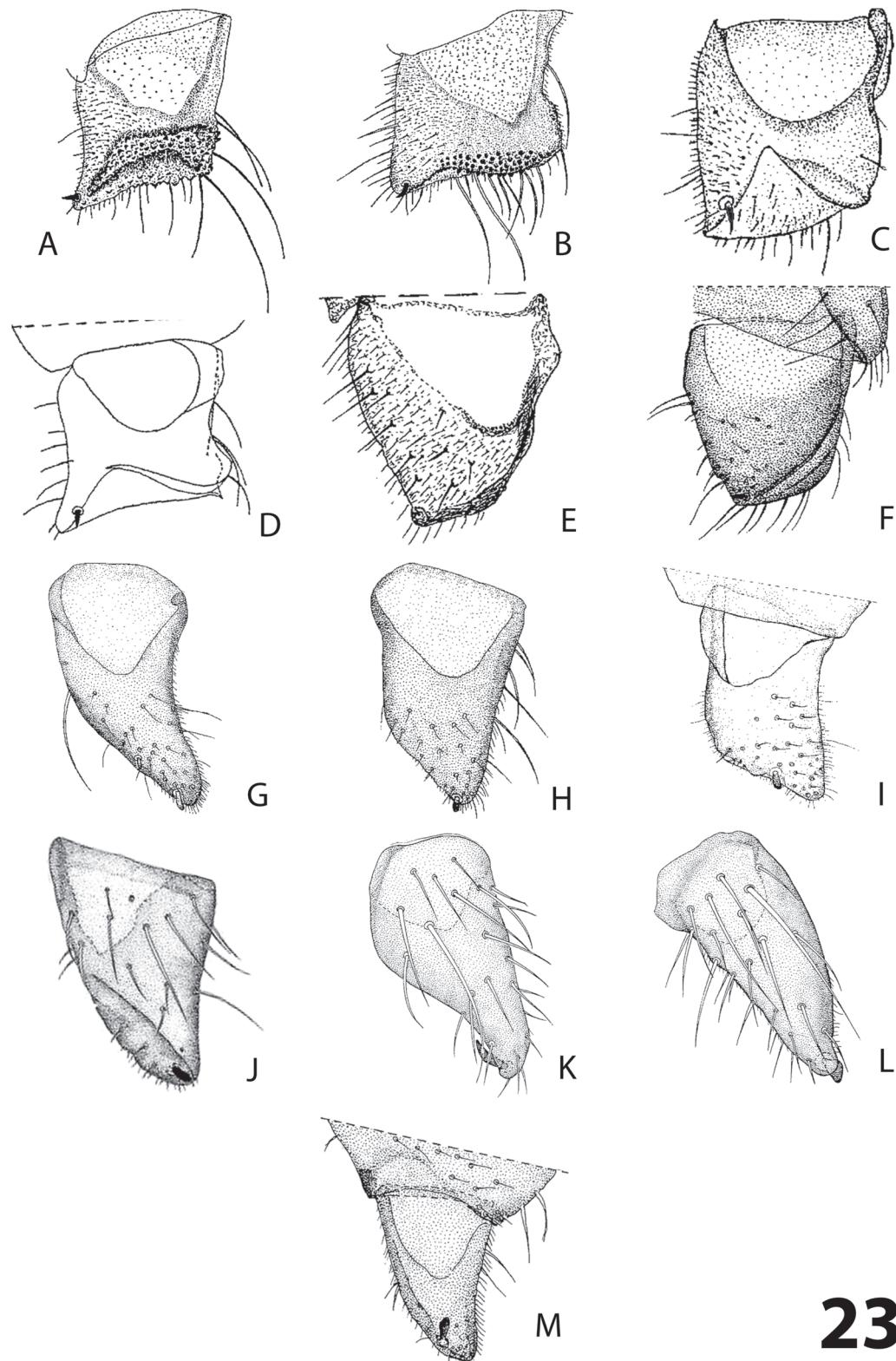
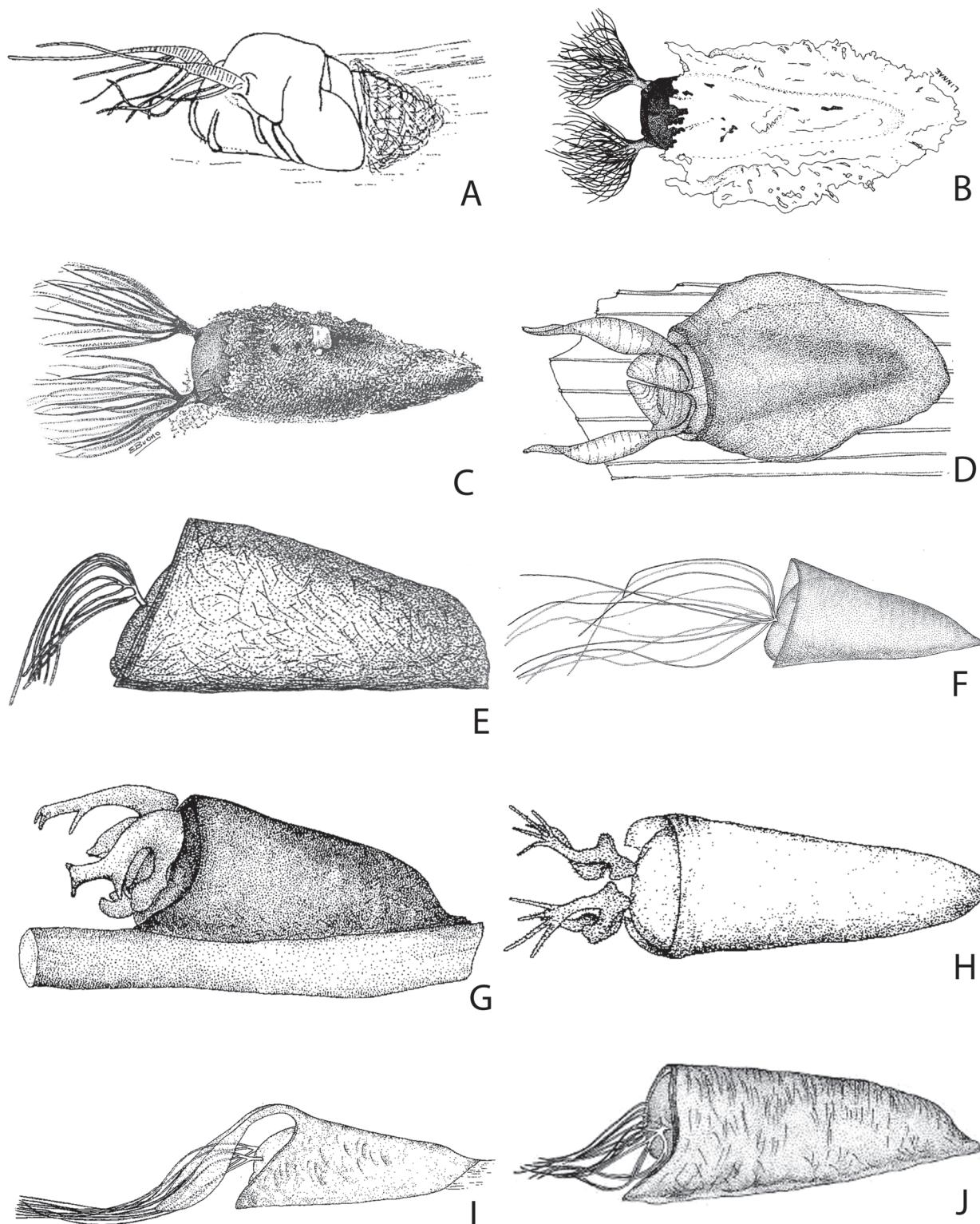
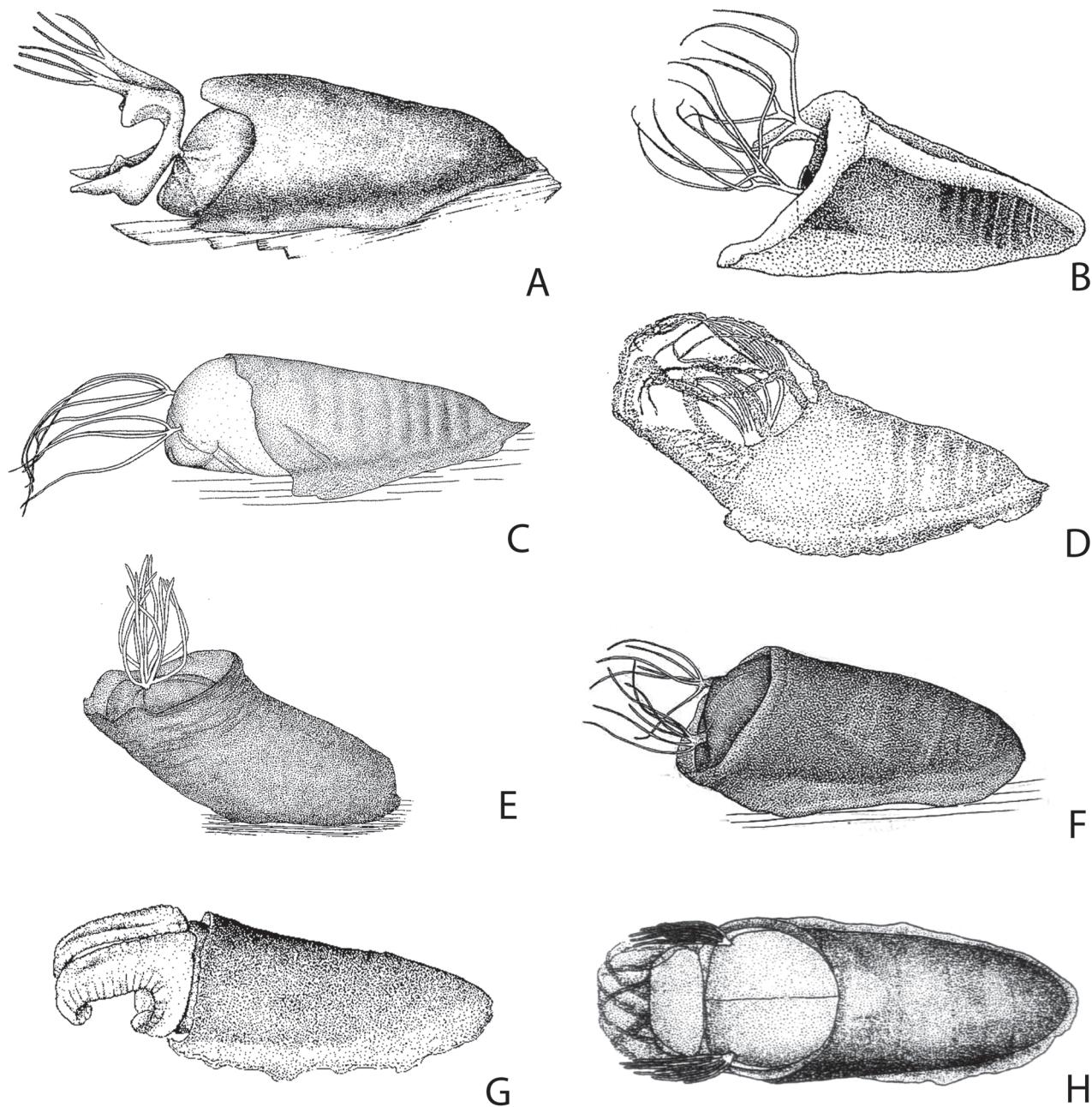


Figure 23. A-M. Male gonostylus. A. *Simulium (Pternaspatha) pichi* Wygodzinsky & Coscarón, 1967. B. *Simulium (Pternaspatha) dureti* Wygodzinsky & Coscarón, 1967. C. *Simulium (Pternaspatha) philippii* Coscarón, 1976. D. *Simulium (Pternaspatha) caprii* Wygodzinsky & Coscarón, 1967. E. *Simulium (Notolepria) exiguum* Roubaud, 1906. F. *Simulium (Notolepria) paraguayanense* Schrottky, 1909. G. *Simulium (Inaequalium) inaequale* (Paterson & Shannon, 1927). H. *Simulium (Inaequalium) subnigrum* Lutz, 1910. I. *Simulium (Psaroniocompsa) limbatum* Knab, 1915. J. *Simulium (Cerqueirellum) cuneatum* (Enderlein, 1936). K. *Simulium (Coscaroniellum) quadrifidum* Lutz, 1917. L. *Simulium (Coscaroniellum) cerradense* Coscarón, Cerqueira, Sato & La Salvia, 1992. M. *Simulium (Coscaroniellum) quadrivittatum* Loew, 1862.



24

Figure 24. Pupae. A. *Tlalocomyia revelata* Wygodzinsky & Díaz Nájera, 1970. B. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962. C. *Gigantodax brophyi* Wygodzinsky & Coscarón, 1989. D. *Paraustrosimulum anthracinum* (Bigot, 1888). E. *Simulium (Chirostilbia) papaveroi* Coscarón, 1982. F. *Simulium (Inaequalium) inaequale* (Paterson & Shannon, 1927). G. *Simulium (Inaequalium) botulibranchium* Lutz, 1910. H. *Simulium (Inaequalium) petropoliense* Coscarón, 1980. I. *Simulium (Psaroniocompsa) anamariae* Vulcano, 1962. J. *Simulium (Cerqueirellum) chaquense* Coscarón, 1971.



25

Figure 25. Pupae. A. *Simulium (Ectemnaspis) furcillatum* Wygodzinsky & Coscarón, 1982. B. *Simulium (Ectemnaspis) anaimense* Coscarón & Muñoz de Hoyos, 1995. C. *Simulium (Psilopeltia) tenuipes* Knab, 1914. D. *Simulium (Hemicnetha) rubrithorax* Lutz, 1909. E. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. F. *Simulium (Hemicnetha) oviedoi* Ramírez Pérez, 1971. G. *Smulium (Hearlea) ethelae* Dalmat, 1950. H. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838.

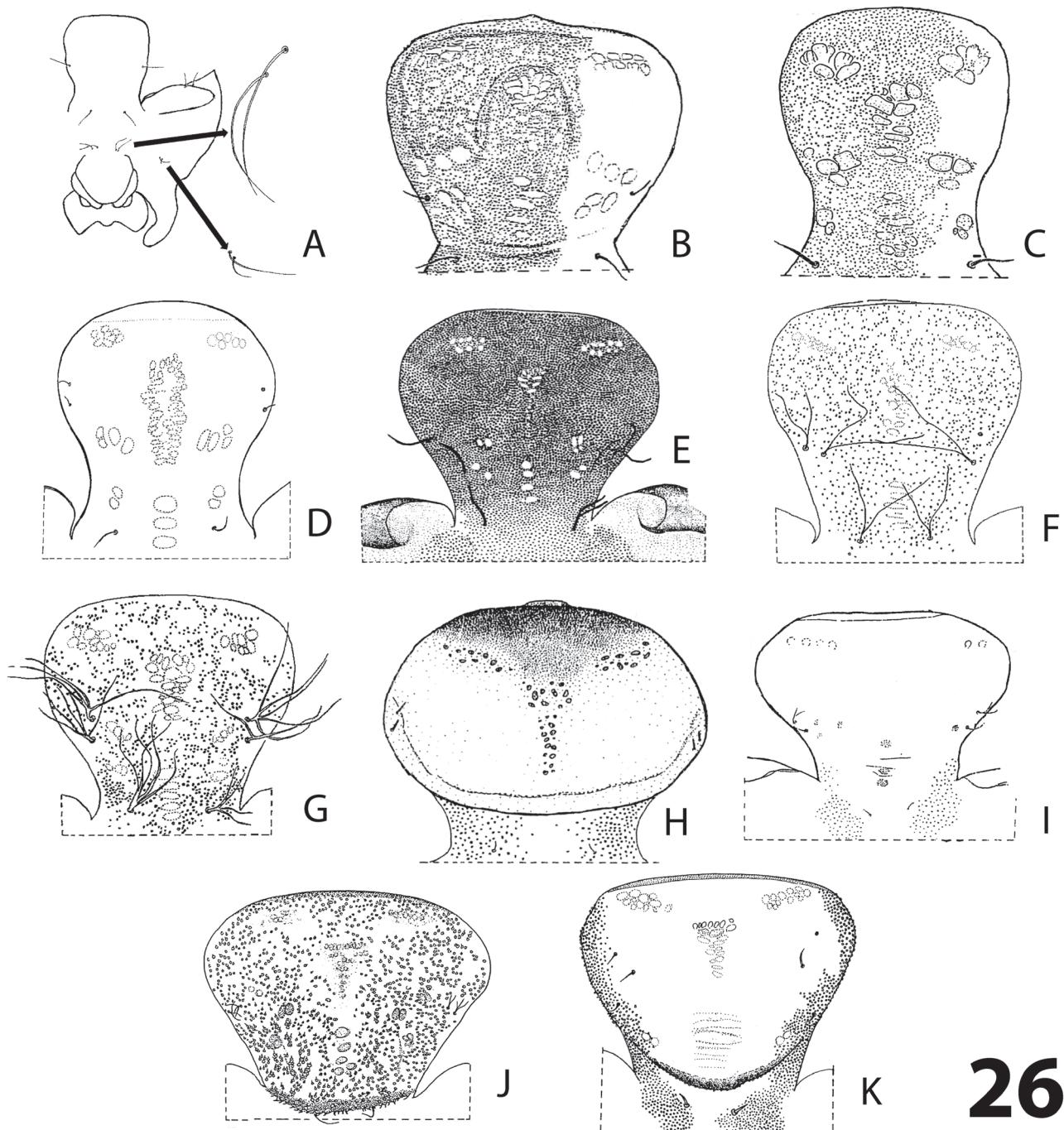


Figure 26. Frontoclypeus. A. *Araucnephia montana* (Philippi, 1865). B. *Pedrowygomyia jatunchuspi* (Wygodzinsky & Coscarón, 1989). C. *Gigantodax marginalis* (Edwards, 1931). D. *Kempfsimulium simplicicolor* (Lutz, 1910). E. *Simulium (Chirostilbia) pertinax* Kollar, 1832. F. *Simulium (Psaroniocompsa) incrustatum* Lutz, 1910. G. *Simulium (Ectemnaspis) dinellii* Joan, 1912. H. *Simulium (Hemicnetha) rubrithorax* Lutz, 1909. I. *Simulium (Hemicnetha) brachycladum* Lutz & Pinto, 1931. J. *Simulium (Hemicnetha) oviedoi* Ramírez Pérez, 1971. K. *Simulium (Hearlea) delatorrei* Dalmat, 1950.

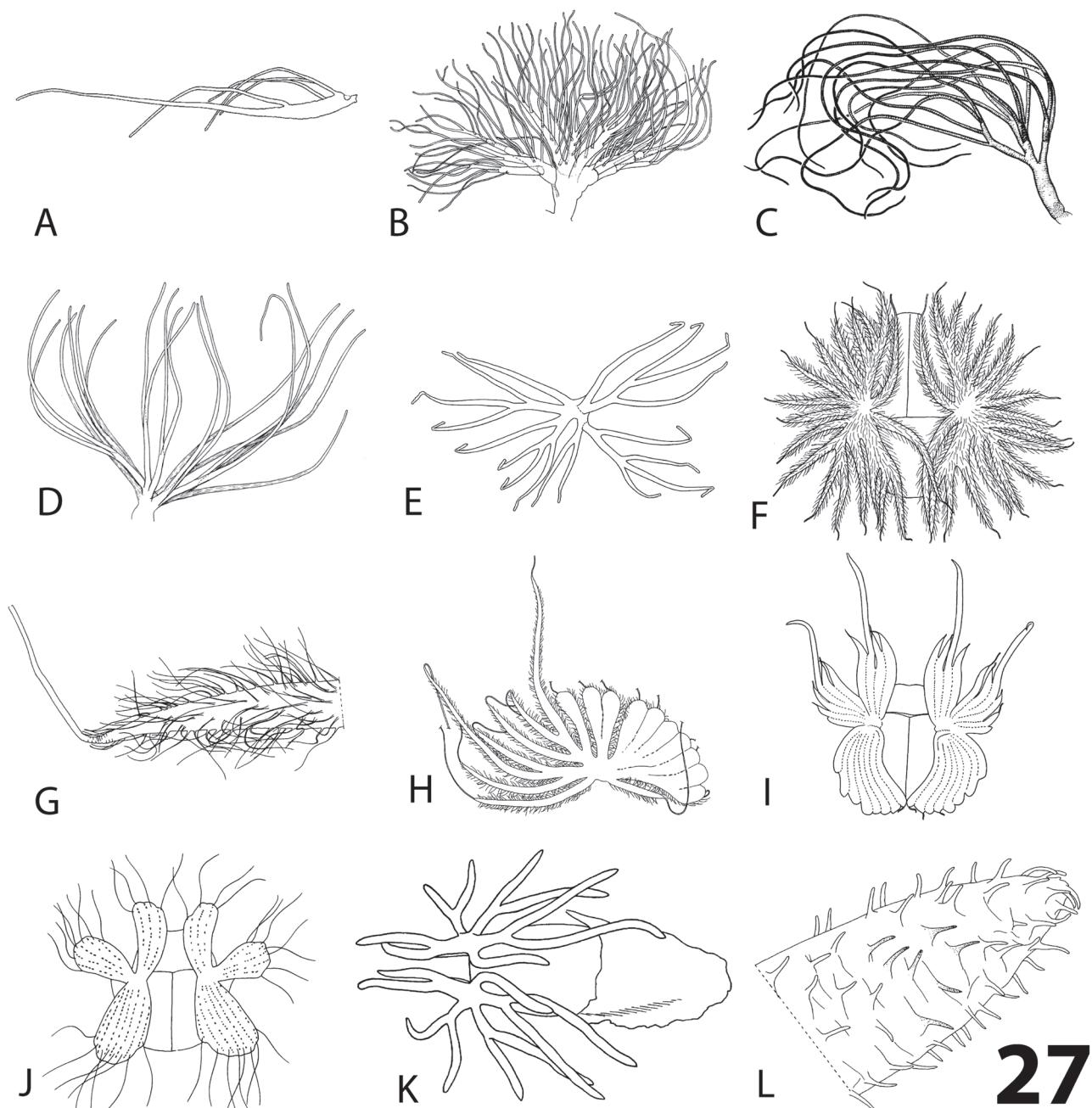


Figure 27. Gills. A. *Tlalocomyia revelata* Wygodzinsky & Díaz Nájera, 1970. B. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962. C. *Gigantodax eremicus* Wygodzinsky & Coscarón, 1989. D. *Gigantodax brophyi* (Edwards, 1931). E. *Gigantodax pennipunctus* Enderlein, 1933. F-G. *Gigantodax arrarteorum* Wygodzinsky & Coscarón, 1989 (G: apex gill branch). H. *Gigantodax brevis* Wygodzinsky & Coscarón, 1989. I. *Gigantodax cormonsi* Wygodzinsky & Coscarón, 1989. J. *Gigantodax siberianus* Wygodzinsky & Coscarón, 1989. K-L. *Gigantodax bettyae* Wygodzinsky, 1974 (L: apex of gill branch).

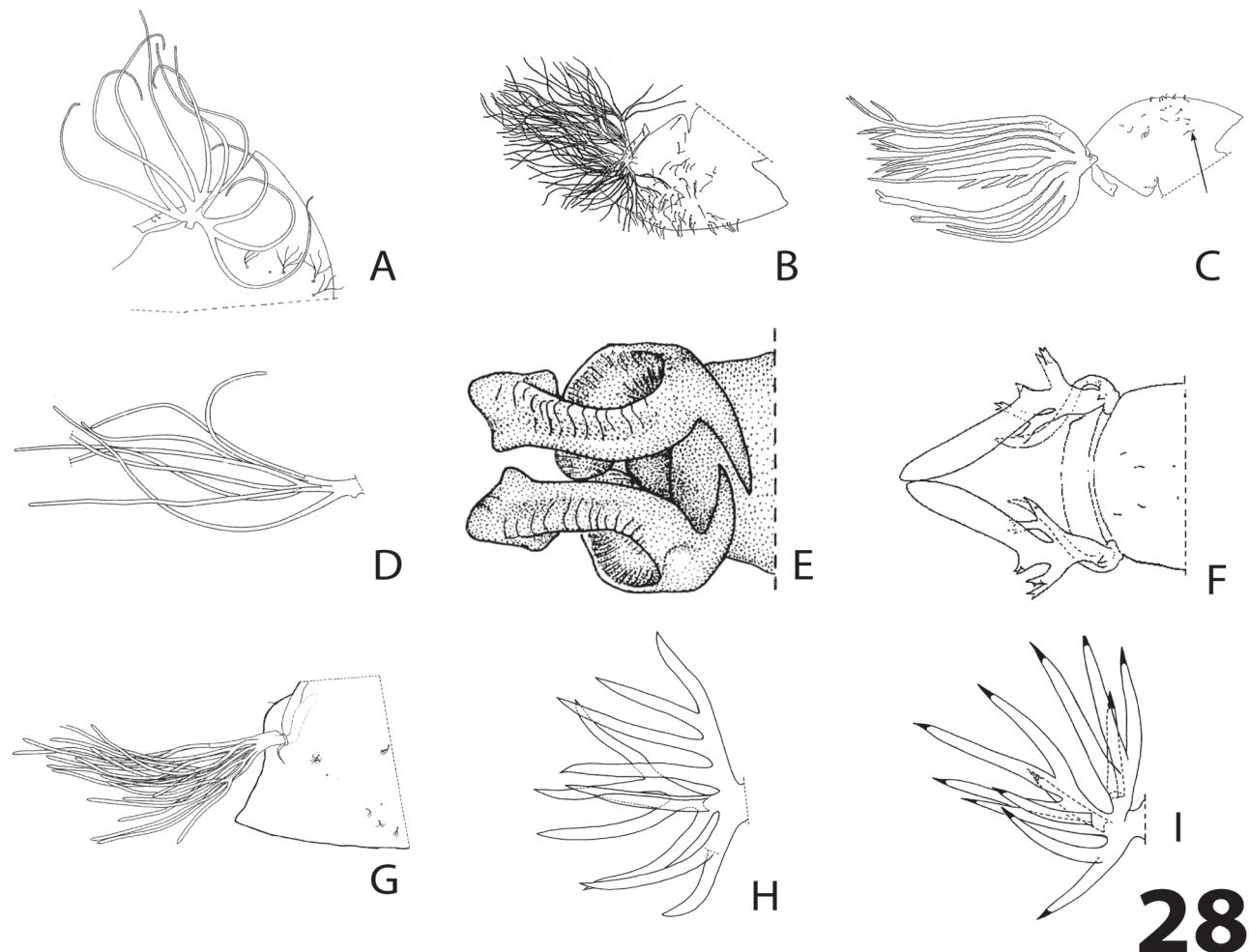
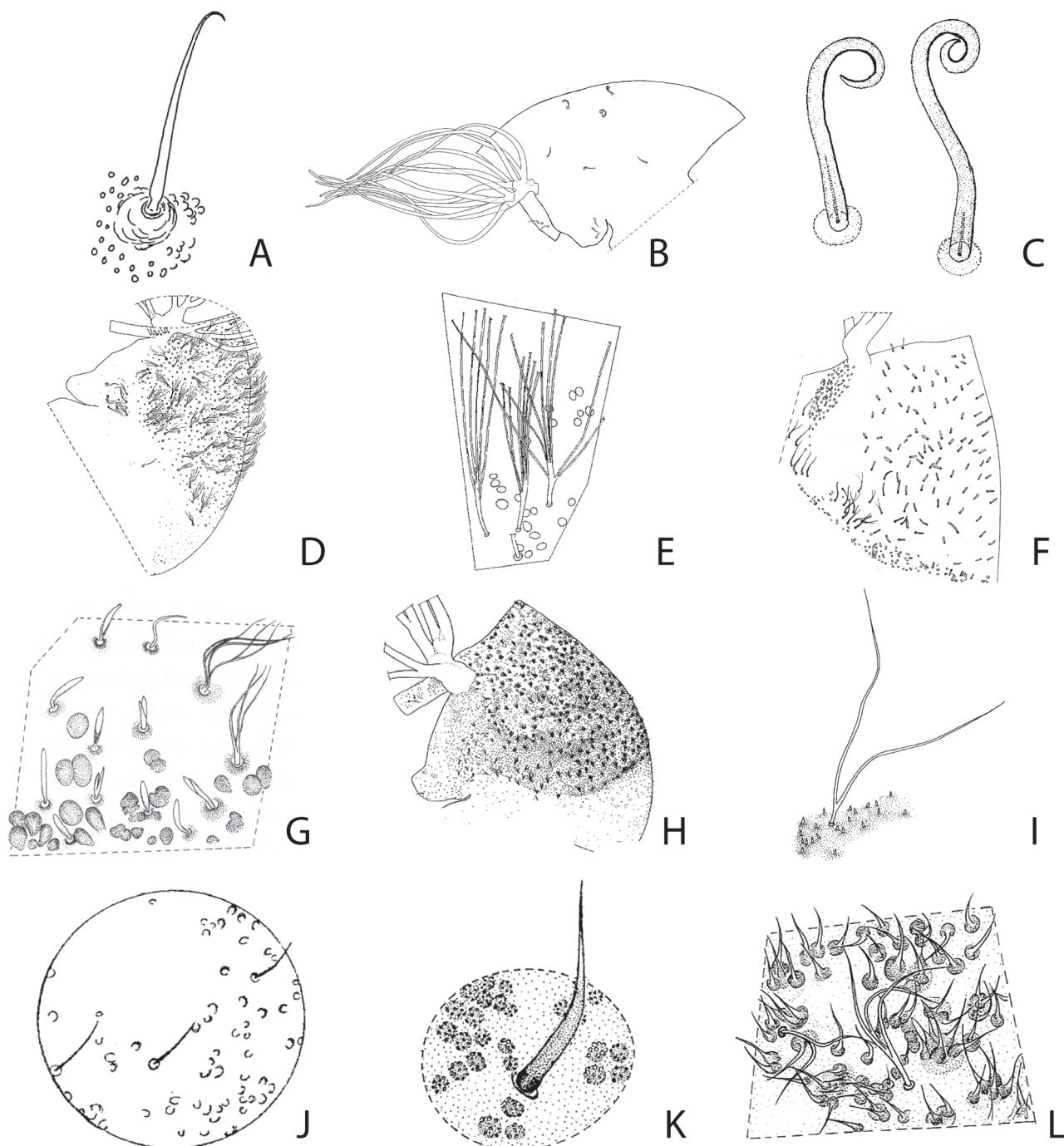


Figure 28. Gills. A. *Simulium (Chirostilbia) distinctum* Lutz, 1910. B. *Simulium (Chirostilbia) obesum* Vulcano, 1959. C. *Simulium (Aspathia) sandyi* Coscarón, Ibáñez Bernal & Coscarón Arias, 1999. D. *Simulium (Hemicnetha) rubriithorax* Lutz, 1909. E. *Simulium (Hearlea) dalmati* Vargas & Díaz Nájera, 1948. F. *Simulium (Hearlea) johnsoni* Vargas & Díaz Nájera, 1957. G. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. H. *Simulium (Thrysopelma) guianense* Wise, 1911. I. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909.



29

Figure 29. Dorsal portion of thorax and trichomes of pupa. A, C, E, G, I-K, trichomes enlarged. A. *Gigantodax minor* Wygodzinsky & Coscarón, 1989. B-C. *Kempfsimulium simplicicolor* (Lutz, 1910). D-E. *Simulium (Pternaspatha) dureti* Wygodzinsky & Coscarón, 1967. F-G. *Simulium (Pternaspatha) barbatipes* (Enderlein, 1934). H. *Simulium (Pternaspatha) stelliferum* Coscarón & Wygodzinsky, 1972. I. *Simulium (Coscaroniellum) quadrifidum* Lutz, 1917. J. *Simulium (Psilopelmia) blancasi* Wygodzinsky & Coscarón, 1970. K. *Simulium (Aspathia) sandyi* Coscarón, Ibáñez Bernal & Coscarón Arias, 1999. L. *Simulium (Hemicnetha) oviedoi* Ramírez Pérez, 1971 (acuminated tubercles and trichomes).

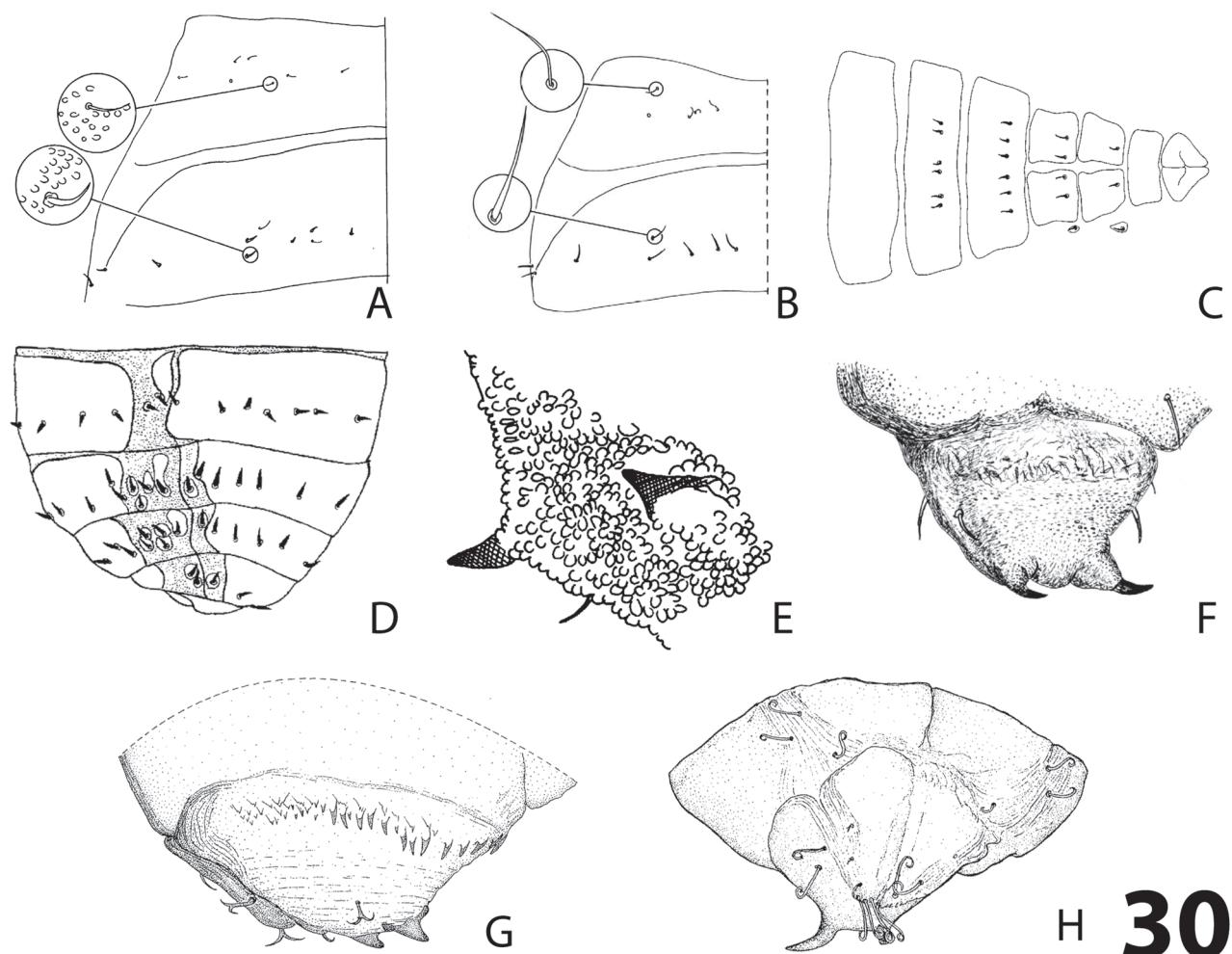


Figure 30. Chaetotaxy of abdomen of pupa. A-B. Tergites II and III. A. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962. B. *Gigantodax rufidulus* Wygodzinsky & Coscarón, 1989. C. *Cnesia dissimilis* (Edwards, 1931) (distal portion of abdomen, ventral view). D. *Tlalocomyia revelata* Wygodzinsky & Coscarón, 1970, distal portion of abdomen, lateral view. E-H. Apex of abdomen. E. *Gigantodax carmenae* Wygodzinsky & Coscarón, 1989. F. *Gigantodax brophyi* (Edwards, 1931). G. *Paraustrosimulium anthracinum* (Bigot, 1888). H. *Lutzsimulium hirticosta* (Lutz, 1909).

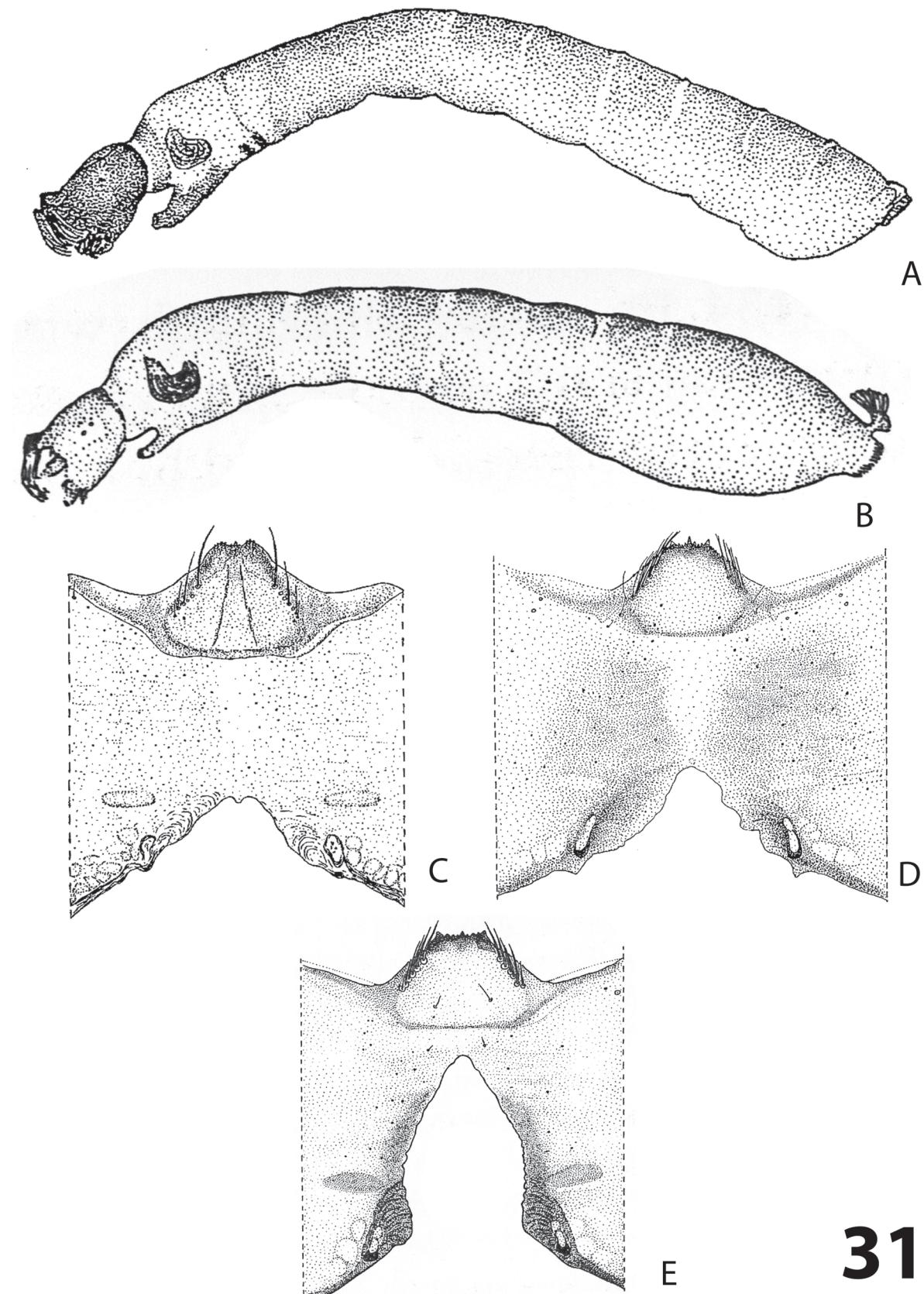


Figure 31. A-B. Larvae, lateral view. A. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. B. *Simulium (Thrysopelma) orbitale* Lutz, 1910. C-E. Larval hypostomium, postgenal bridge and gular cleft. C. *Cnesia ornata* Wygodzinsky & Coscarón, 1973. D. *Simulium (Aspathia) wygoi* Coscarón, Ibáñez Bernal & Coscarón Arias, 1999. E. *Simulium (Ectemnaspis) rorotaense* Floch & Abonnenc, 1946.

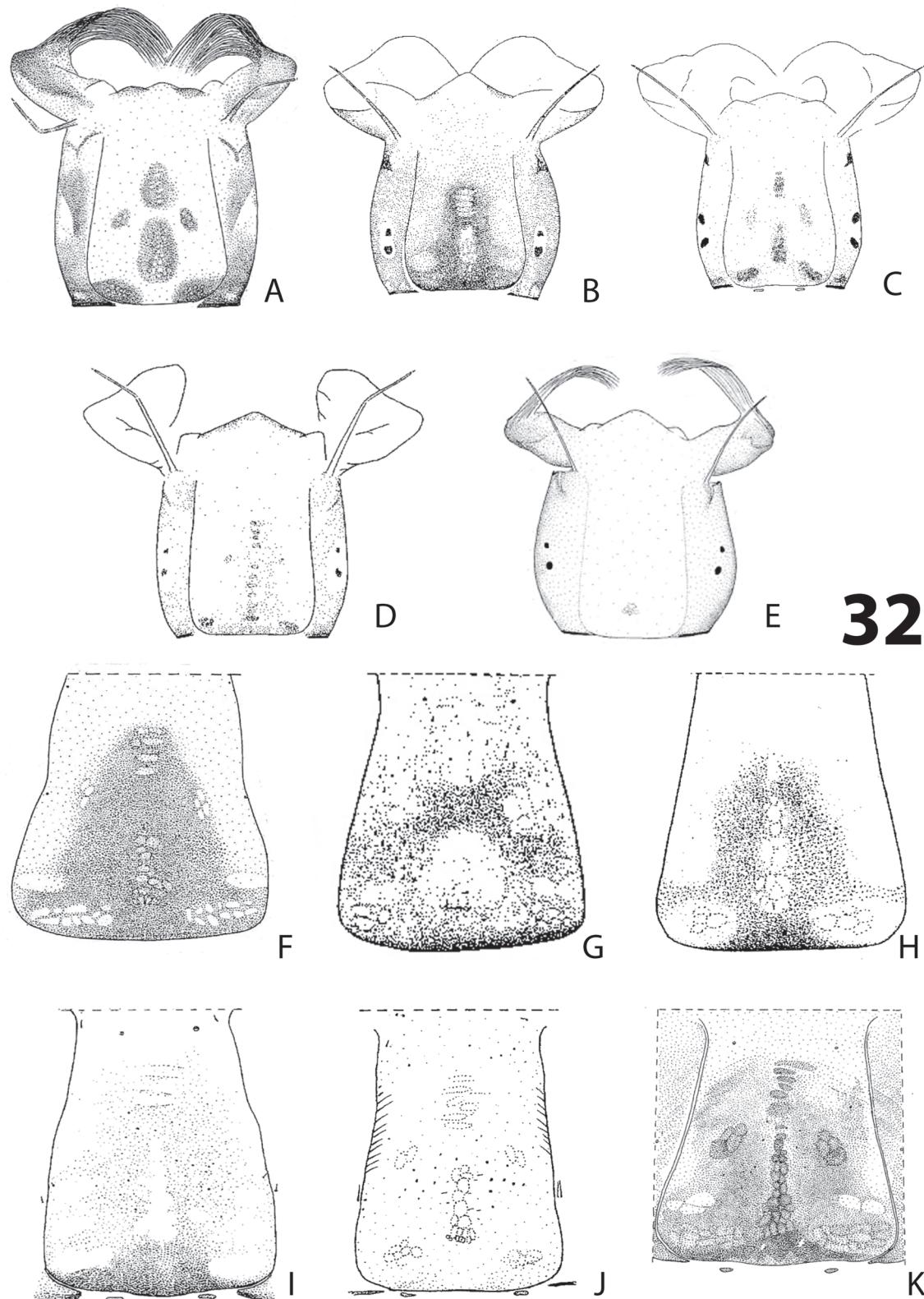
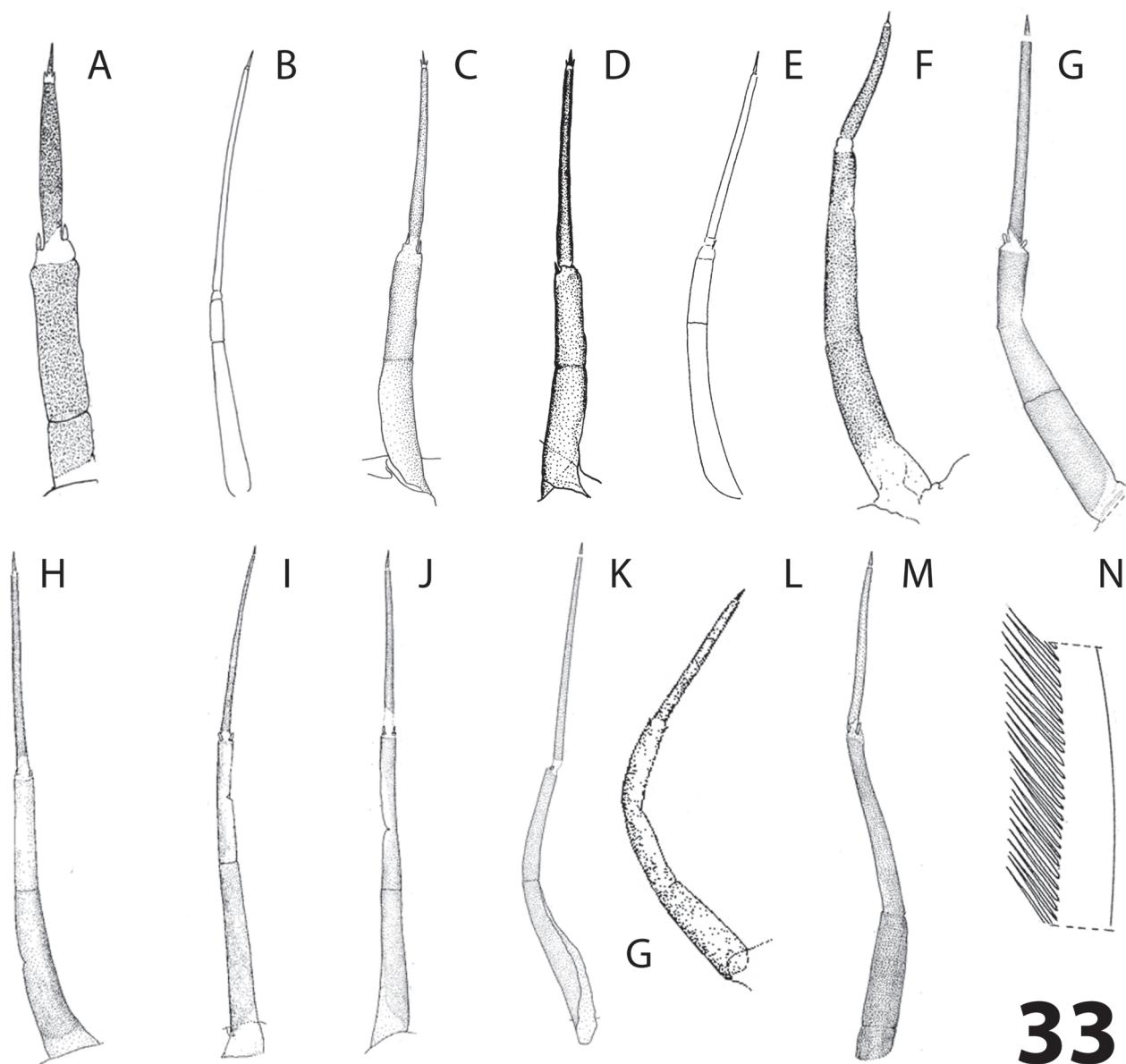
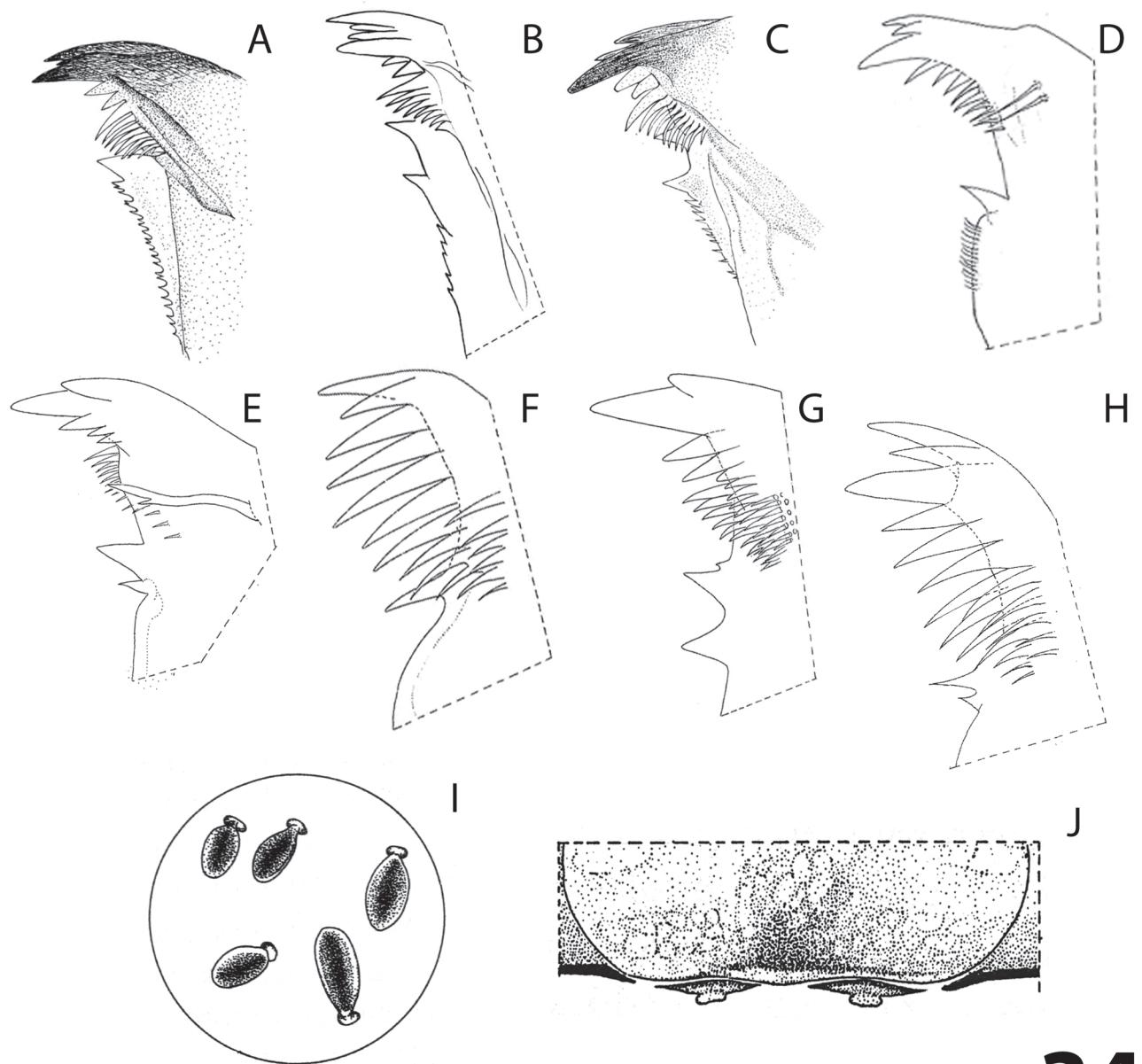


Figure 32. A-E. Larval head, dorsal view. A. *Simulium (Chirostilbia) subpallidum* Lutz, 1910. B. *Simulium (Inaequalium) subnigrum* Lutz, 1910. C. *Simulium (Psaroniocompsa) bonaerense* Coscarón & Wygodzinsky, 1984. D. *Simulium (Psaroniocompsa) auristriatum* Lutz, 1910. E. *Simulium (Cerqueirellum) cuneatum* (Enderlein, 1936). F-K. Cephalic apotome. F. *Simulium (Chirostilbia) pertinax* Kollar, 1832. G. *Simulium (Ectemnaspis) romanai* Wygodzinsky, 1951. H. *Simulium (Psilopeltia) escomeli* Roubaud, 1909. I. *Simulium (Psilopeltia) downsi* Vargas, Martínez Palacios & Díaz Nájera, 1946. J. *Simulium (Psilopeltia) zempoalense* Vargas, Martínez Palacios & Díaz Nájera, 1946. K. *Trichodagmias lahillei* (Paterson & Shannon, 1927).



33

Figure 33. Larval antennae. A. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962. B. *Gigantodax minor* Wygodzinsky & Coscarón, 1989. C. *Gigantodax rufidulus* Wygodzinsky & Coscarón, 1989. D. *Gigantodax antarcticus* (Bigot, 1888). E. *Gigantodax pennipunctus* Enderlein, 1933. F. *Simulium (Pternaspatha) limay* Wygodzinsky, 1958. G. *Simulium (Chirostilbia) subpallidum* Lutz, 1910. H. *Simulium (Inaequalium) rappae* Py-Daniel & Coscarón, 1982. I. *Simulium (Psaroniocompsa) auristriatum* Lutz, 1910. J. *Simulium (Cerqueirellum) cuneatum* (Enderlein, 1936). K. *Simulium (Psilopeltmia) quadrifidum* Lutz, 1917. L. *Simulium (Ectemnaspis) perflavum* (Roubaud, 1906). M. *Simulium (Aspathia) metallicum* Bellardi, 1859. N. *Simulium (Hemicnetha) oviedoi* Ramírez Pérez, 1971, comb of cephalic fan.



34

Figure 34. A-H. Larval mandibles. A. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962. B. *Gigantodax minor* Wygodzinsky & Coscarón, 1989. C. *Gigantodax brophyi* (Edwards, 1931). D. *Simulium (Cerqueirellum) oyapockense* Floch & Abonnenc, 1946. E. *Simulium (Coscaroniellum) cerradense* Coscarón, Cerqueira, Sato & La Salvia, 1992. F. *Simulium (Hemicnetha) seriatum* Knab, 1914. G. *Simulium (Hemicnetha) oviedoi* Ramírez Pérez, 1971. H. *Simulium (Trichodagmia) nigrimanum* Macquart, 1838. I. *Simulium (Thrysopelma) orbitale* Lutz, 1910, trichomes of body, integument. J. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909, posterodorsal view of head and cervical sclerites.

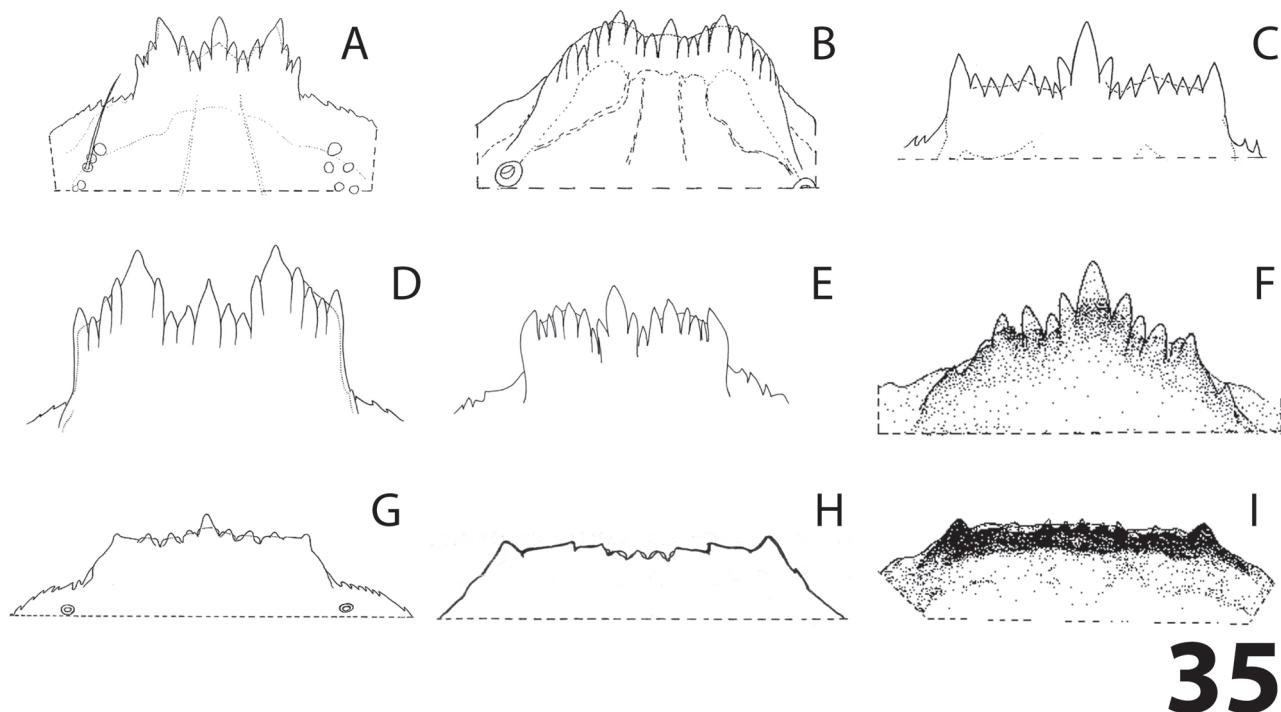


Figure 35. Larval hypostomial teeth. A. *Araucnephia montana* (Philippi, 1865). B. *Cnesia ornata* Wygodzinsky & Coscarón, 1973. C. *Pedrowygomyia cortesi* Wygodzinsky & Coscarón, 1989. D. *Gigantodax minor* Wygodzinsky & Coscarón, 1989. E. *Gigantodax igniculus* Coscarón & Wygodzinsky, 1962. F. *Simulium (Hearlea) paracarolinae* Coscarón, Miranda Esquivel, Moulton, Coscarón Arias & Ibáñez Bernal, 2004. G. *Simulium (Trichodagmia) lahillei* (Paterson & Shannon, 1927). H. *Simulium (Thrysopelma) orbitale* Lutz, 1910. I. *Simulium (Thrysopelma) scutistriatum* Lutz, 1909.

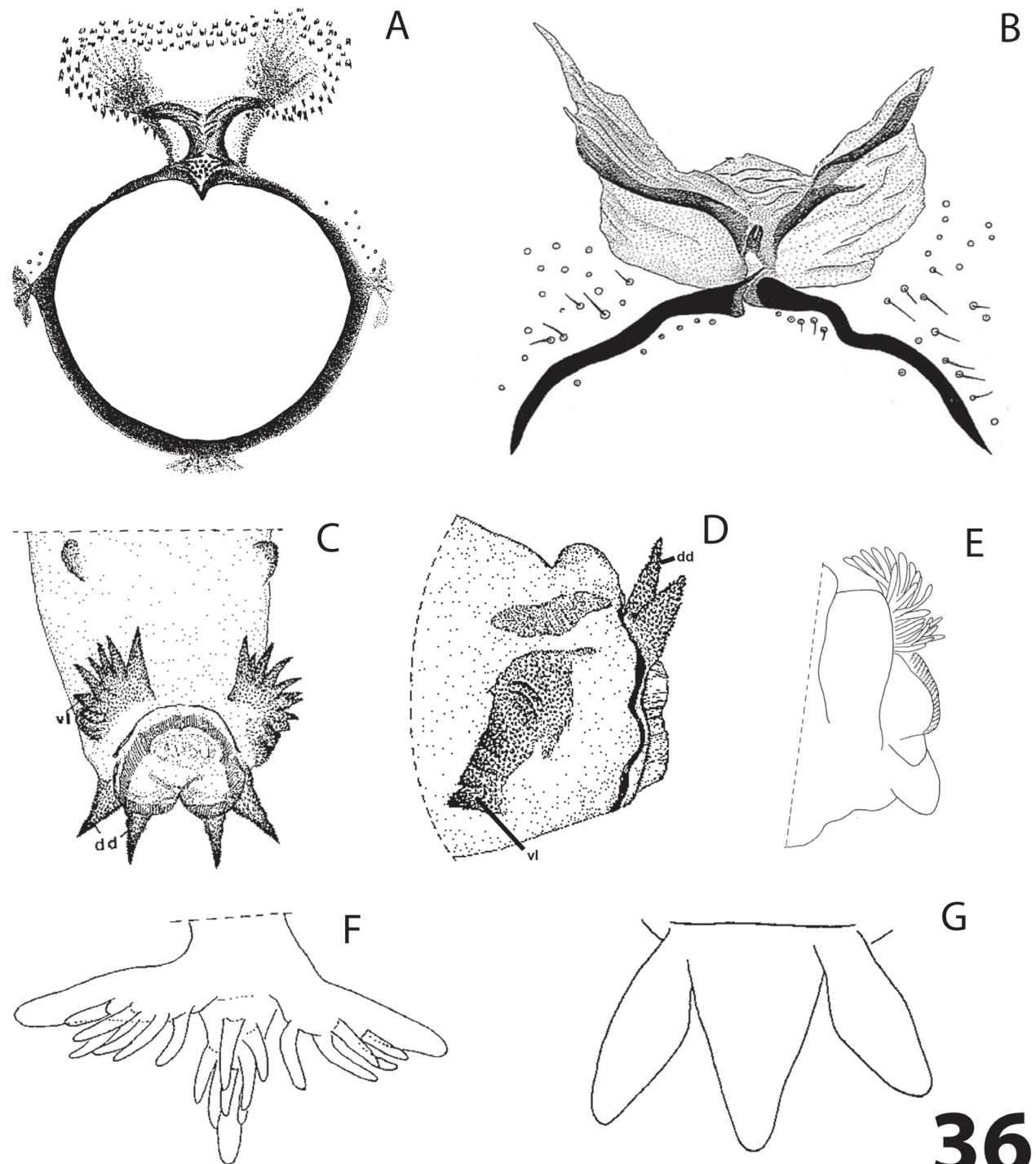


Figure 36. A-B. Larval anal sclerite. A. *Gigantodax cilicinus* Wygodzinsky & Coscarón, 1989. B. *Simulium (Chirostilbia) serranum* Coscarón, 1981. C-D. Distal portion of larval abdomen, showing anal sclerite and accessory plates (dd: dorsolateral conical accessory plate; vl: ventrolateral accessory plate). C. *Simulium (Hearlea) gorirossiae* Vargas & Díaz Nájera, 1957. D. *Simulium (Hearlea) temascalense* Díaz Nájera & Vulcano, 1962. E-G. Larval rectal papillae. E. *Gigantodax multifilis* Wygodzinsky & Coscarón, 1989 (terminal portion of abdomen, lateral view, showing rectal papillae with abundant diverticula and anal ring of crochets). F. *Simulium (Pternaspatha) diamantinum* Coscarón & Coscarón Arias, 1996 (with several diverticula on rectal lobes). G. *Simulium (Psilopeltmia) blancasi* Wygodzinsky & Coscarón, 1970 (rectal papillae with single lobes).